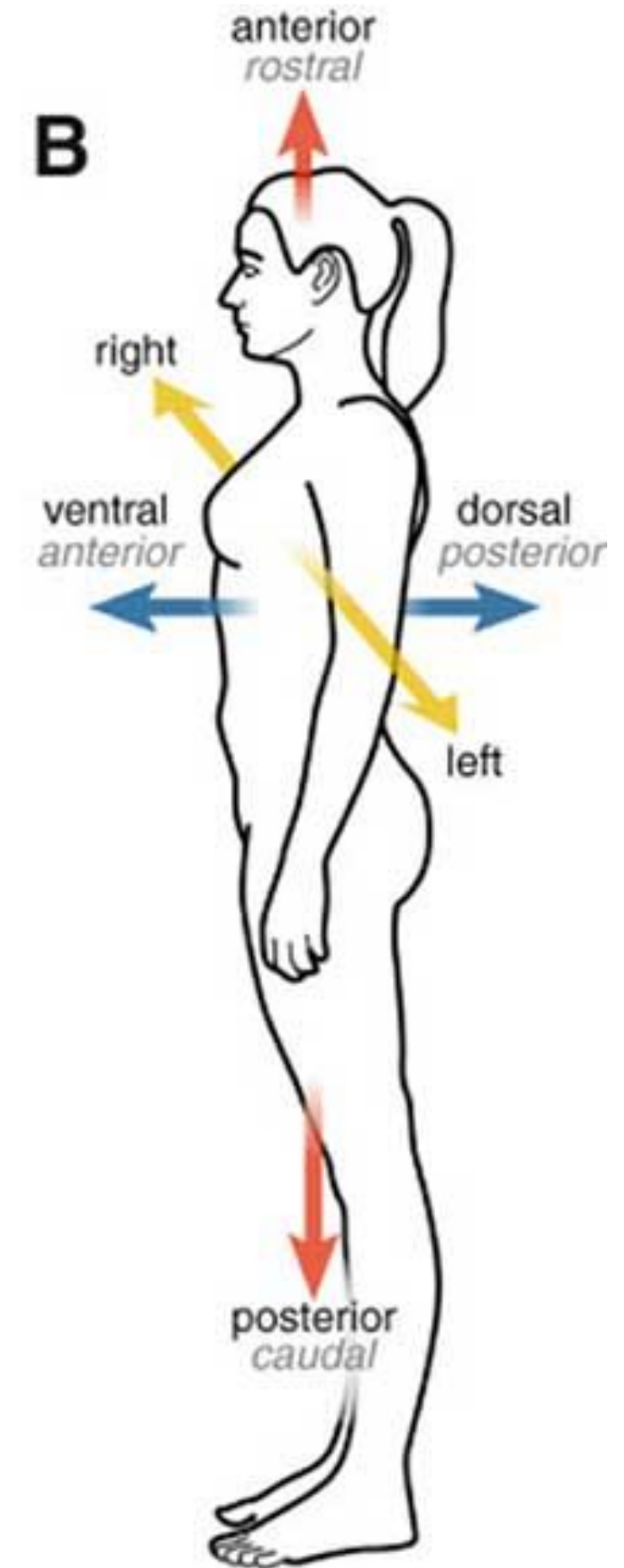
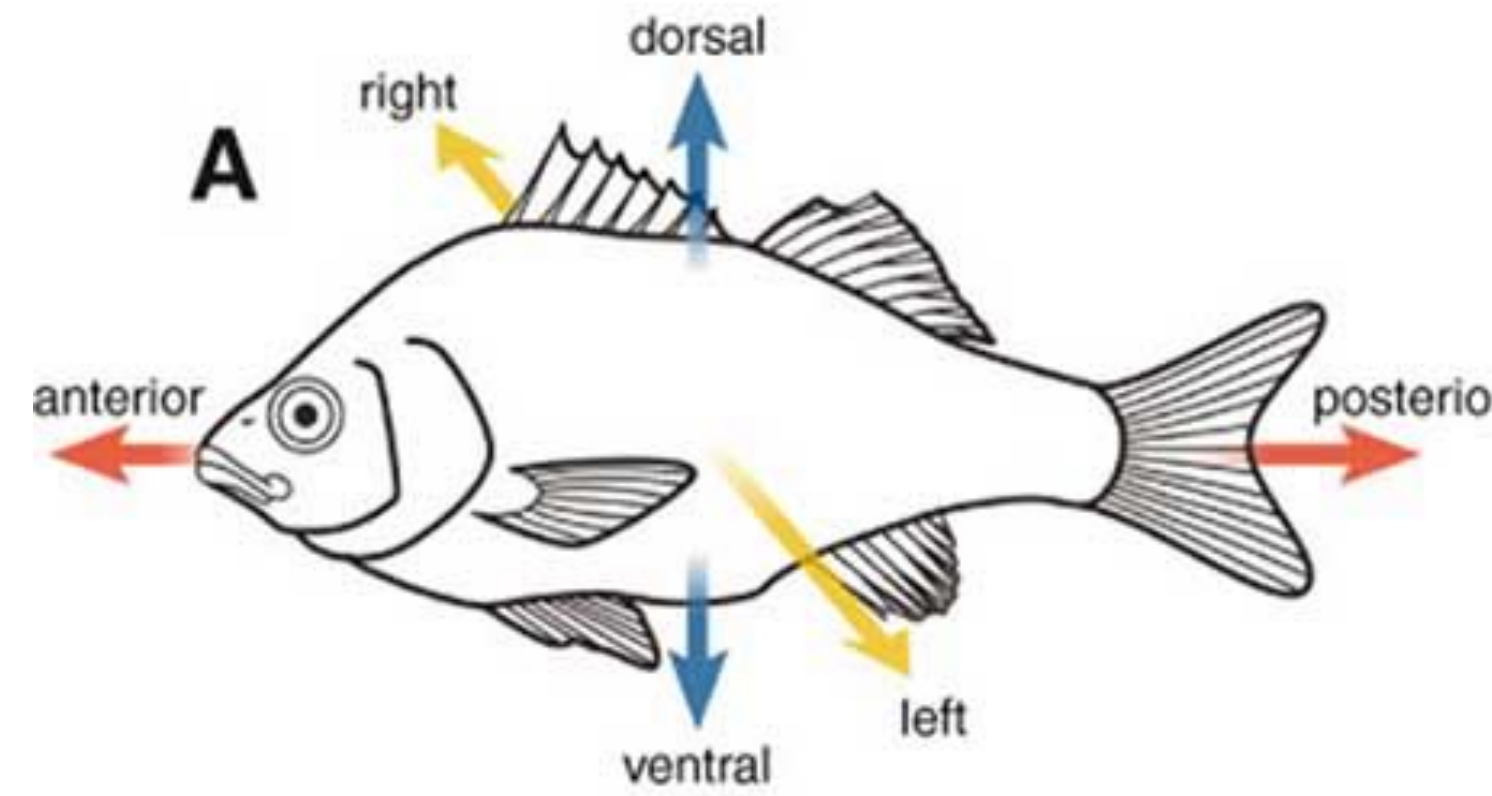




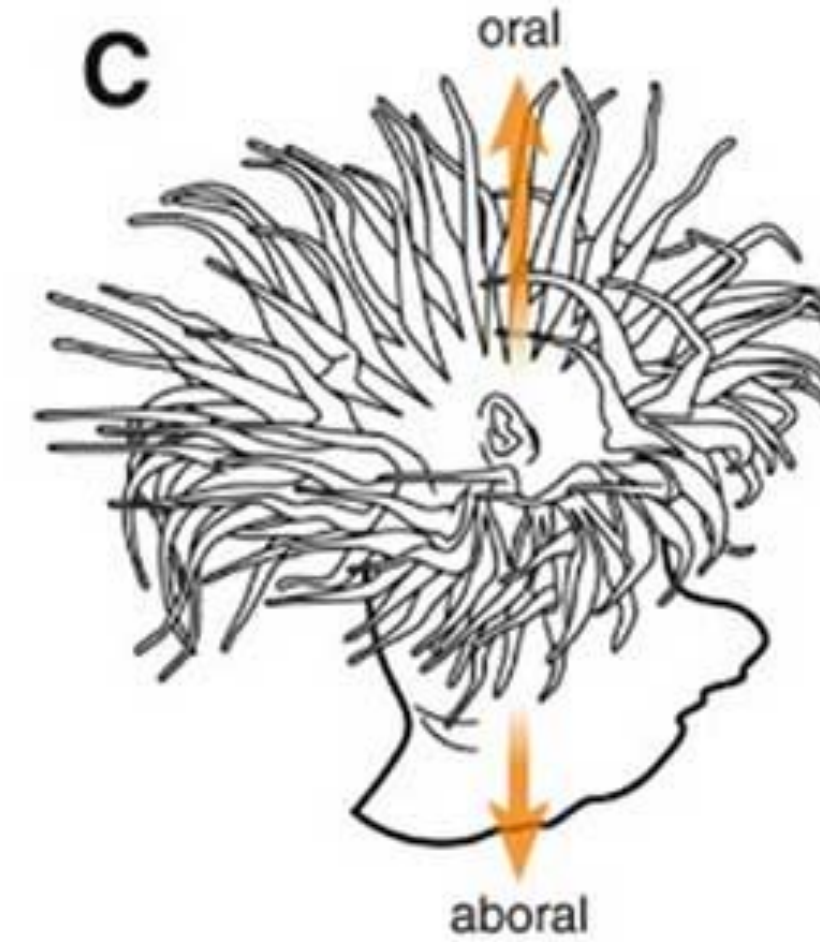
# Big questions



**Vertebrates+: all 3 spatial axes determined**



**Anenome, Hydra, jellyfish: radial symmetry**



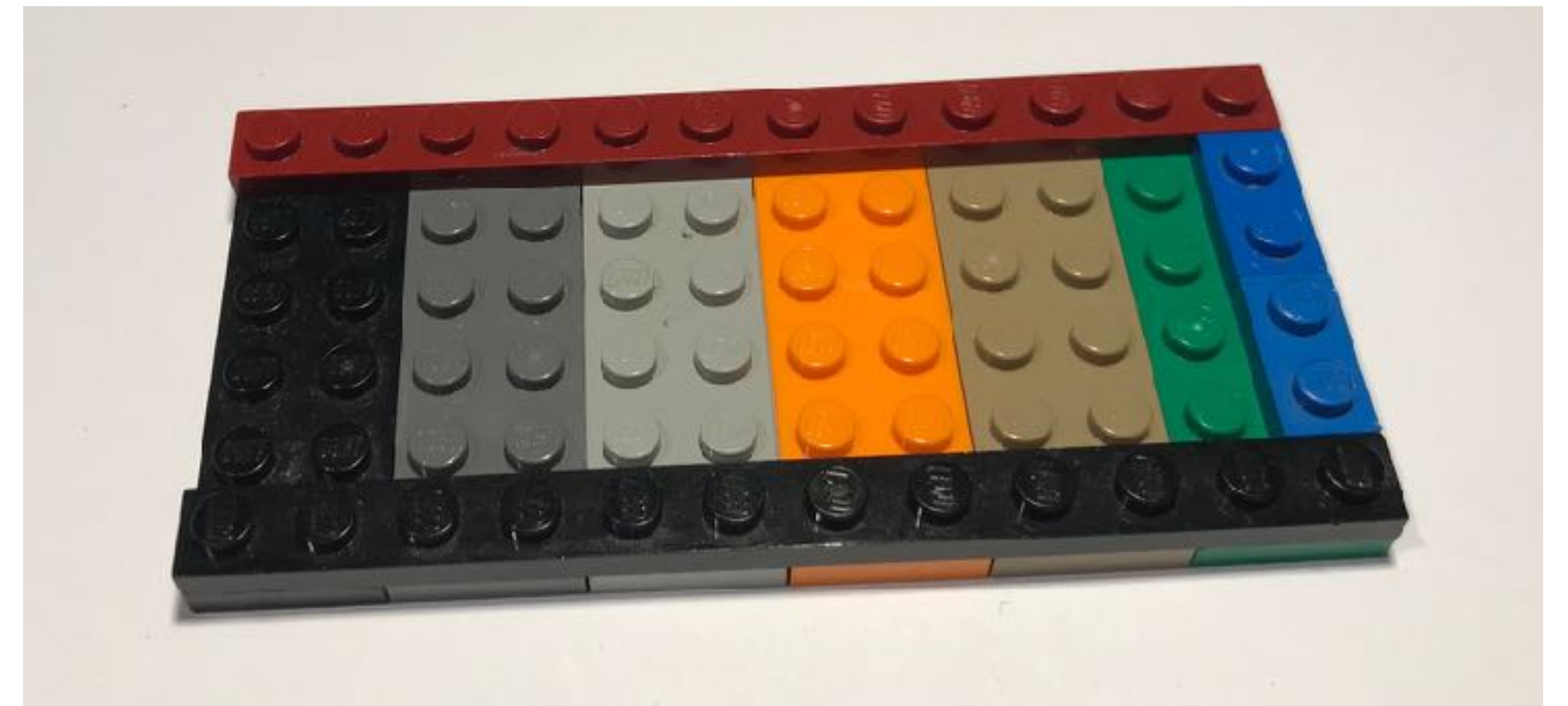
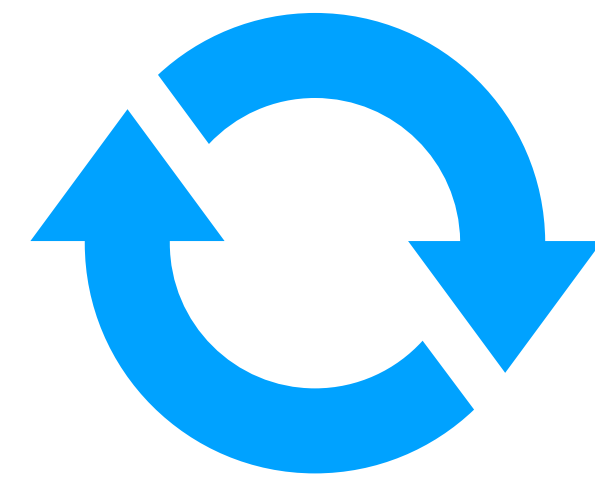
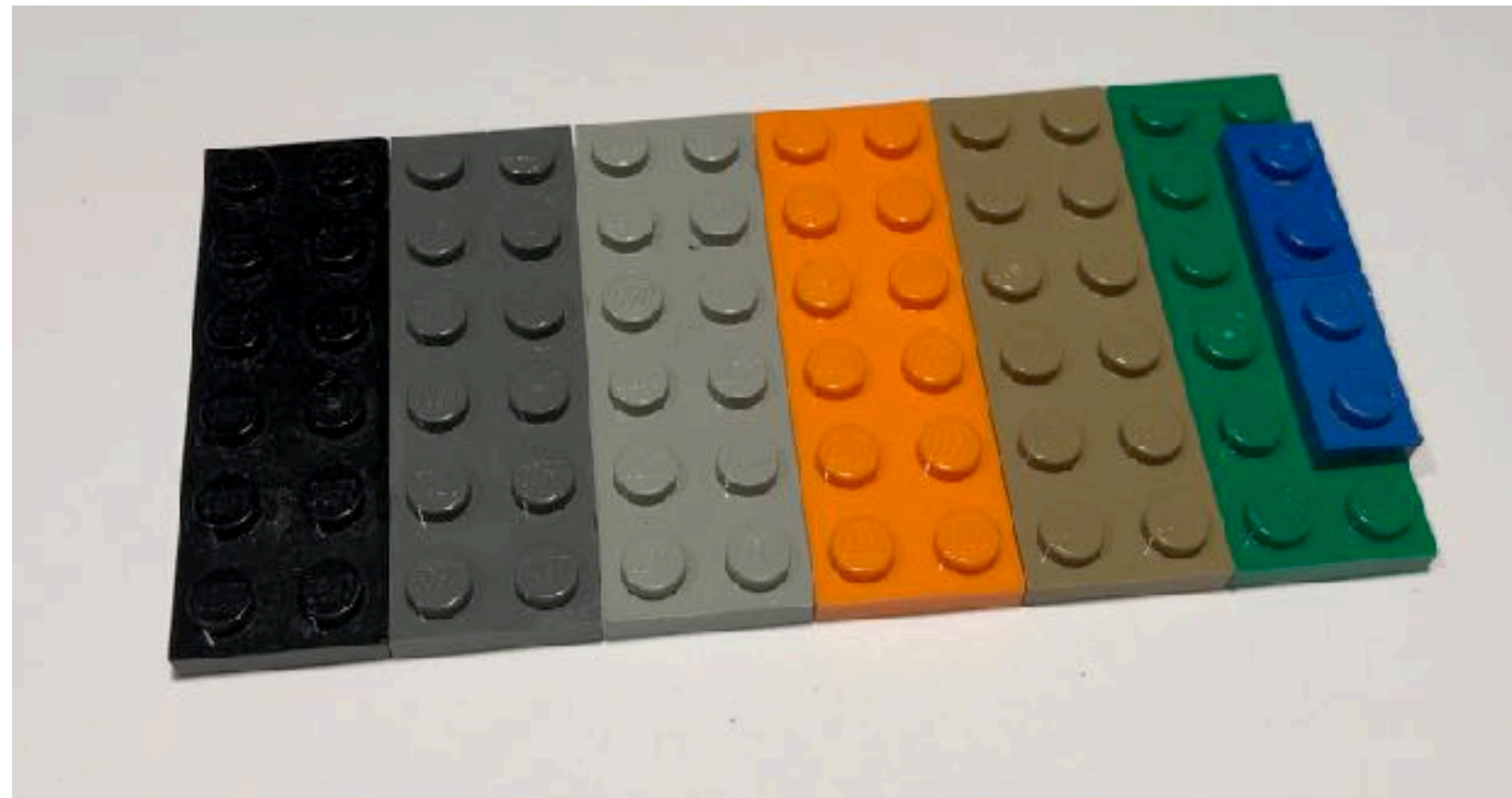
PMID: 25140222

# Today's menu - 2 courses

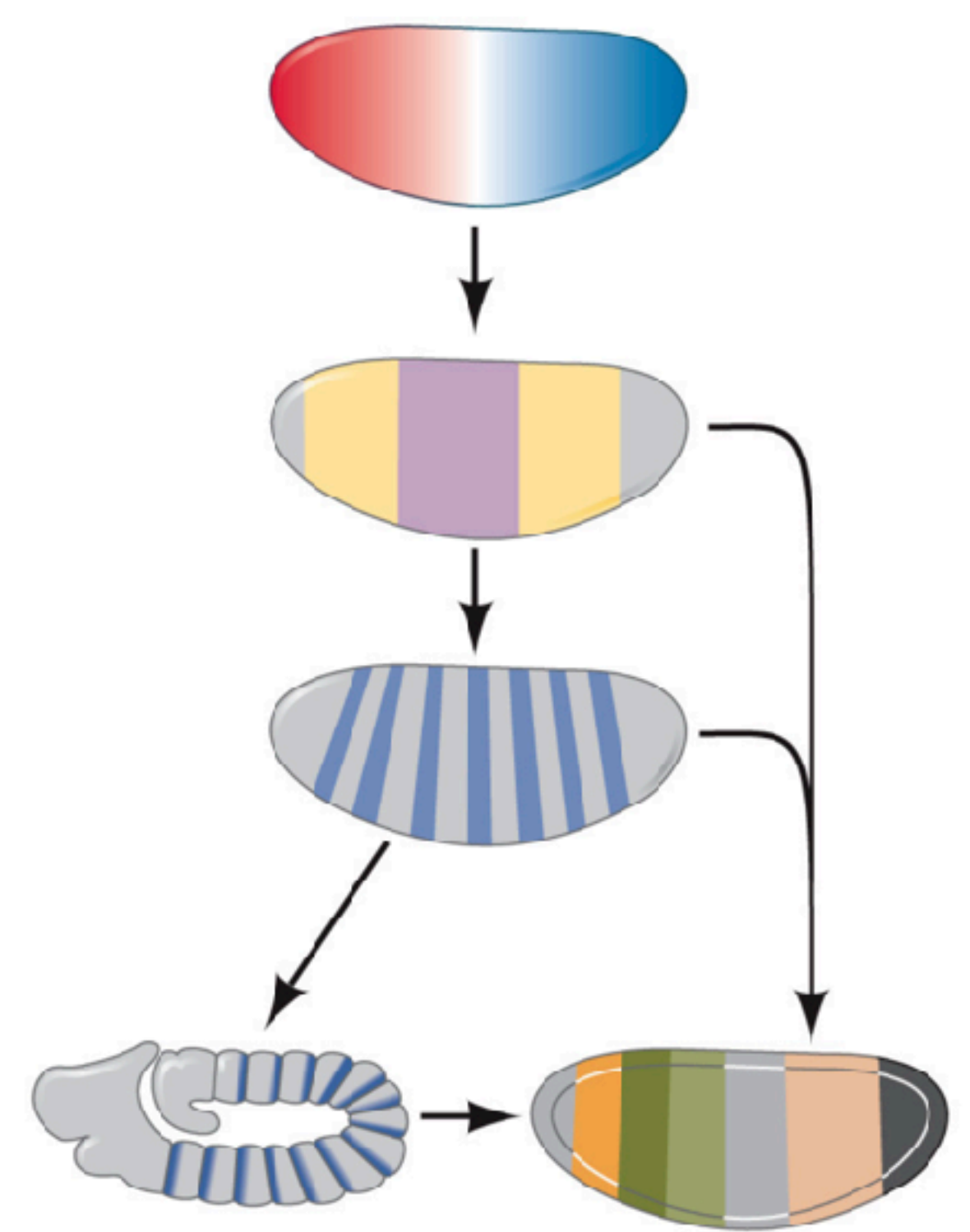
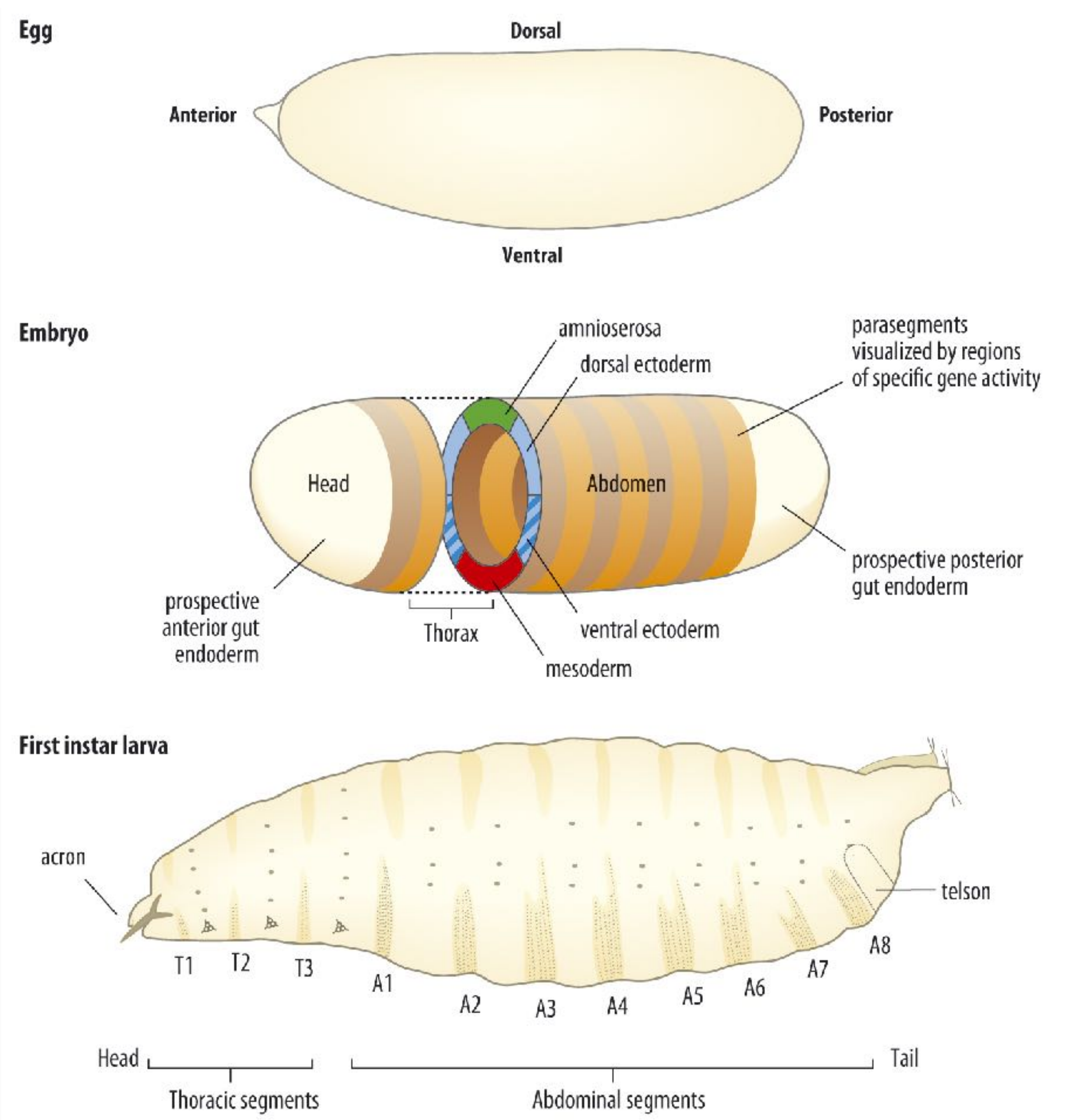
- **Front & Back, Head & Tail**
- Fate maps, specification and commitment
- Gastrulation and patterning of germ layers
- *Nobel* experiment - embryonic induction
- Integrating extracellular gradients
- *Left and Right*
- *Situs inversus*
- *Cilia in the node - directed flow*
- *Planar polarity*
- *Nodal and calcium signals to the lateral plate*

**Head (Anterior) - Tail (Posterior) axis**

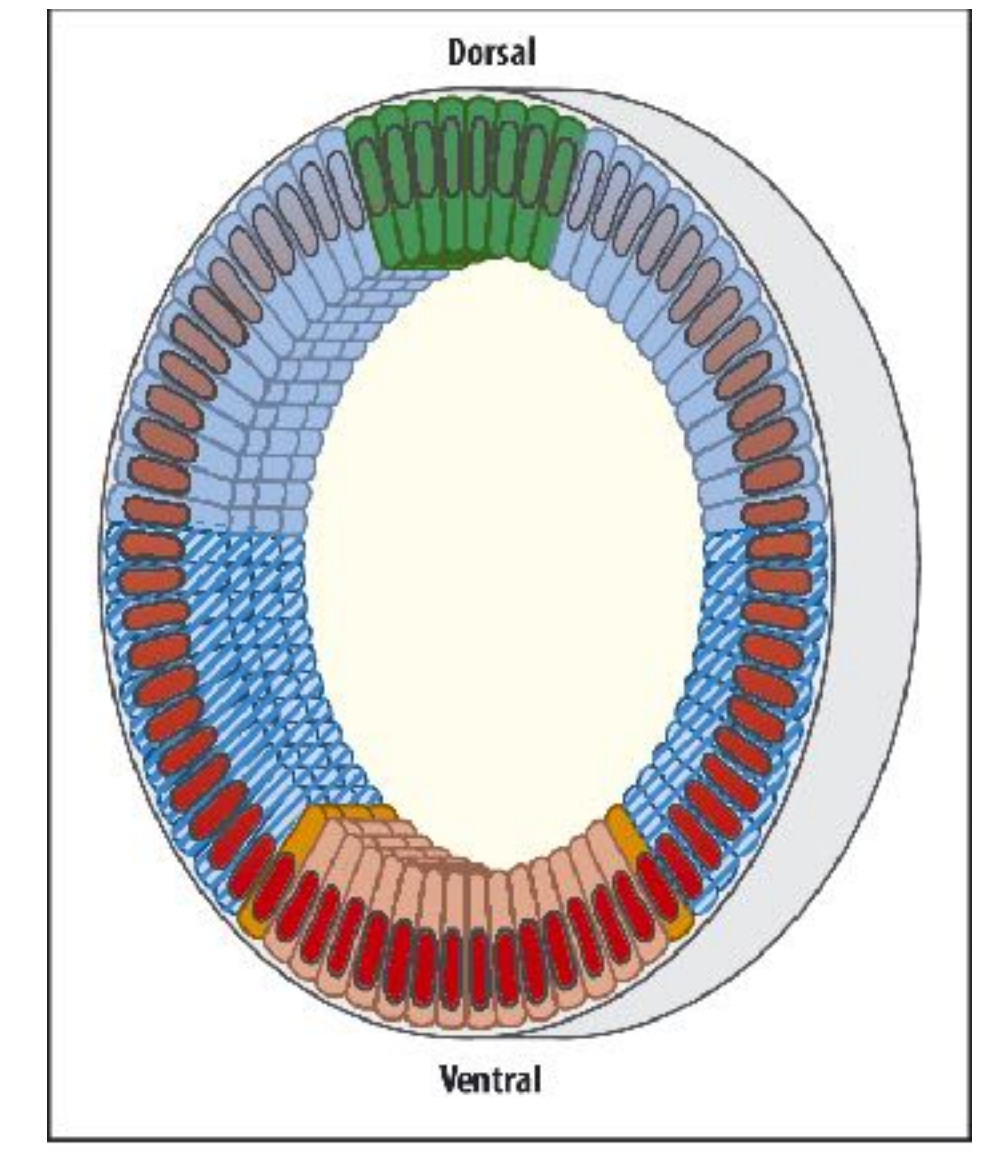
**Front (Ventral) - Back (Dorsal) axis**



# !!! Independent gene systems & orthogonal spatial mapping from egg to embryo for Anterior-Posterior and Dorsal-Ventral axes in fruit fly...



**Anterior-Posterior  
(Week 4)**



- amnioserosa (*zerknüllt*)
- dorsal ectoderm (*decapentaplegic, tolloid*)
- ▨ neurectoderm (*rhombal*)
- mesoderm (*twist, snail*)
- mesectoderm (*single-minded*)
- Dorsal gradient

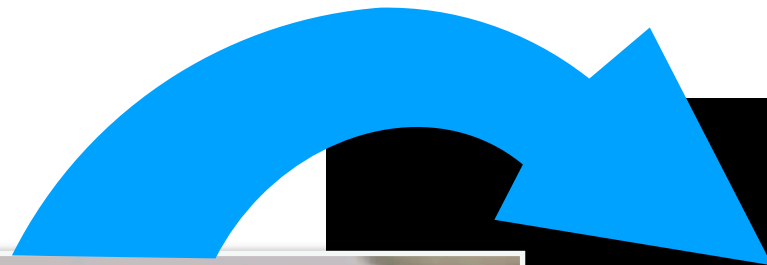
**Dorsal-Ventral  
(Week 10)**



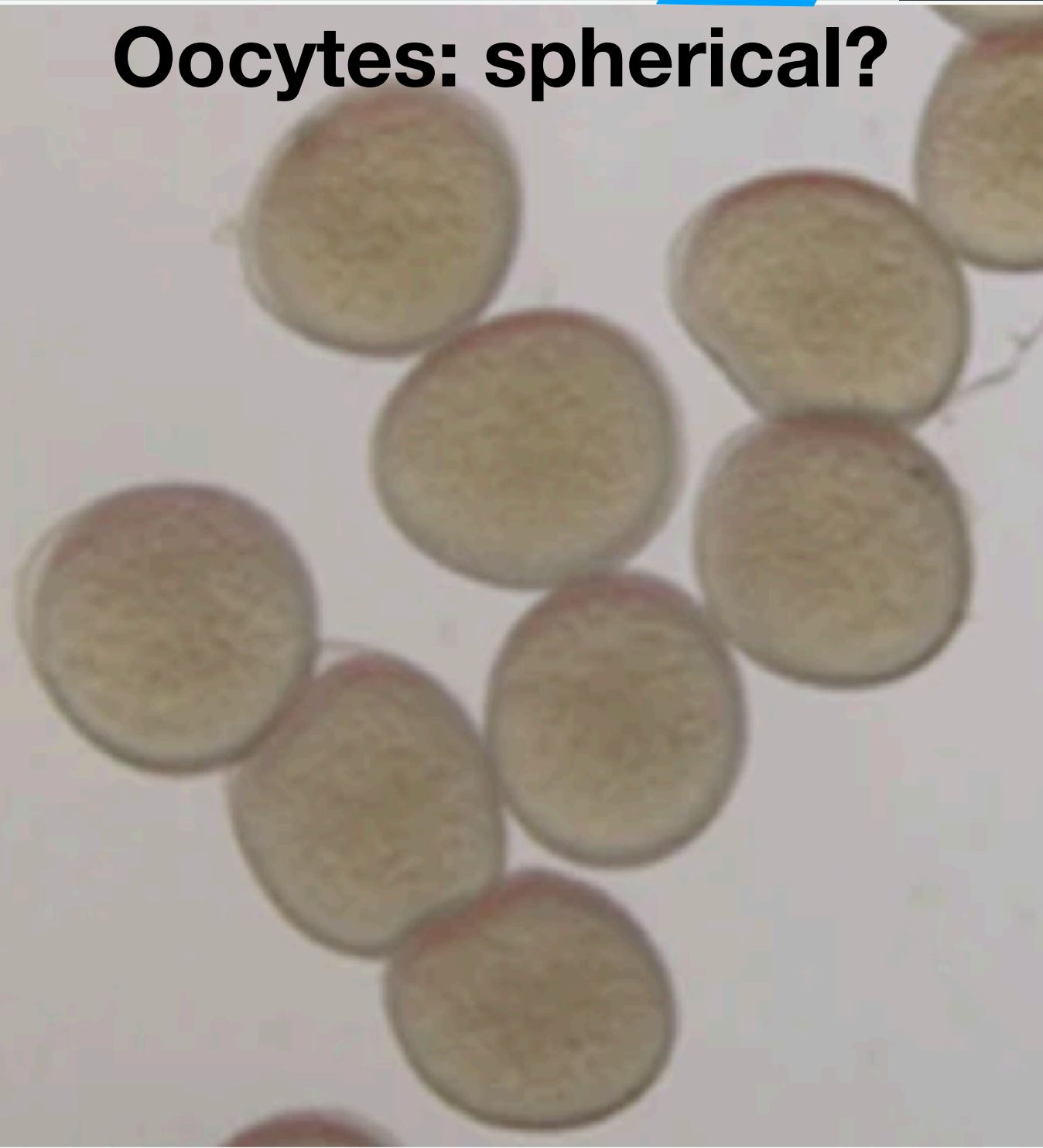
# How is symmetry broken?



Fertilization (Week 1)



Oocytes: spherical?



Zygote: radial



00:36:00

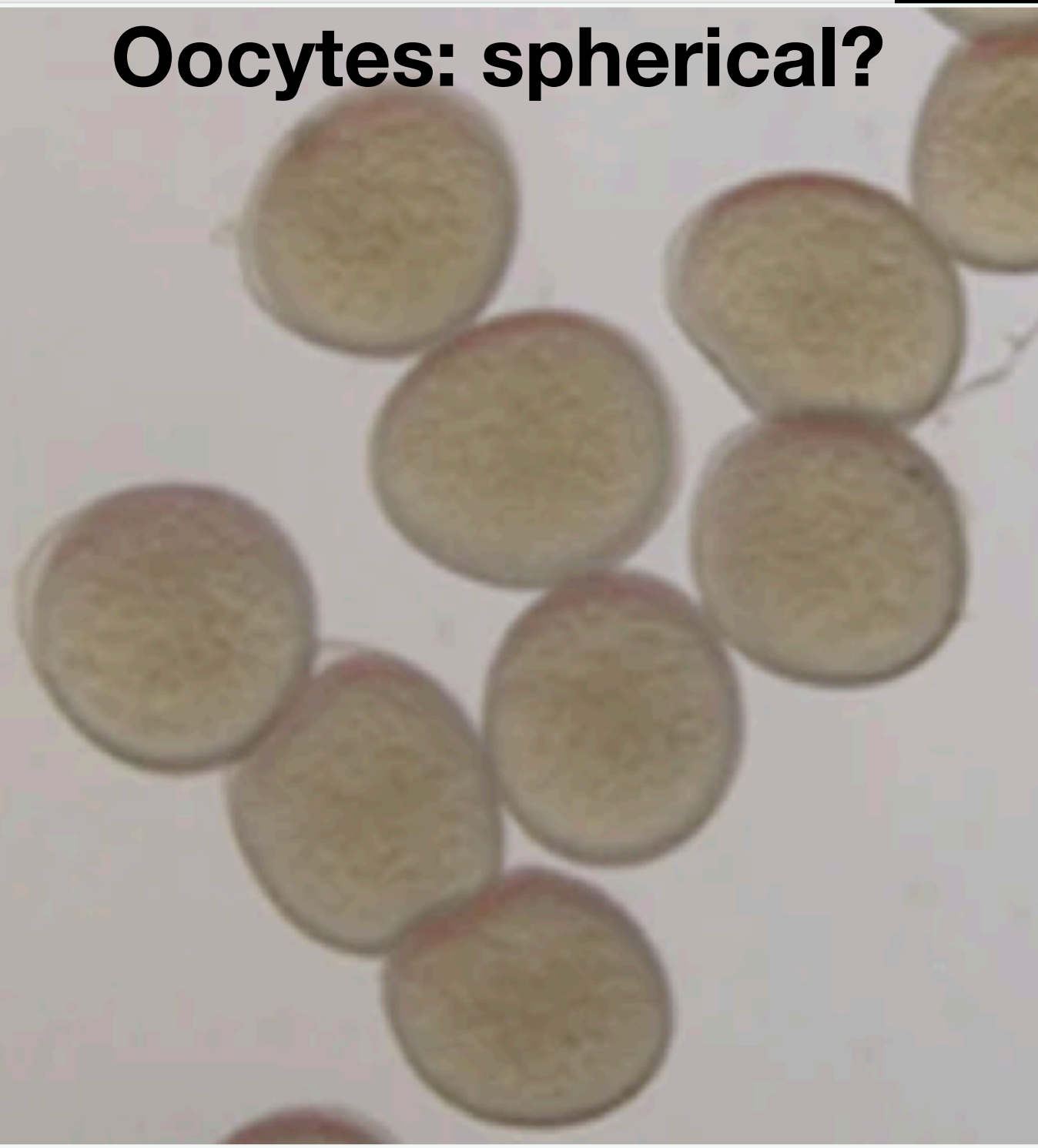
Ian Swinburne and Sean Megason. Dept. of Systems Biology, Harvard Medical School  
Licensed under Creative Commons by Attribution 3.0



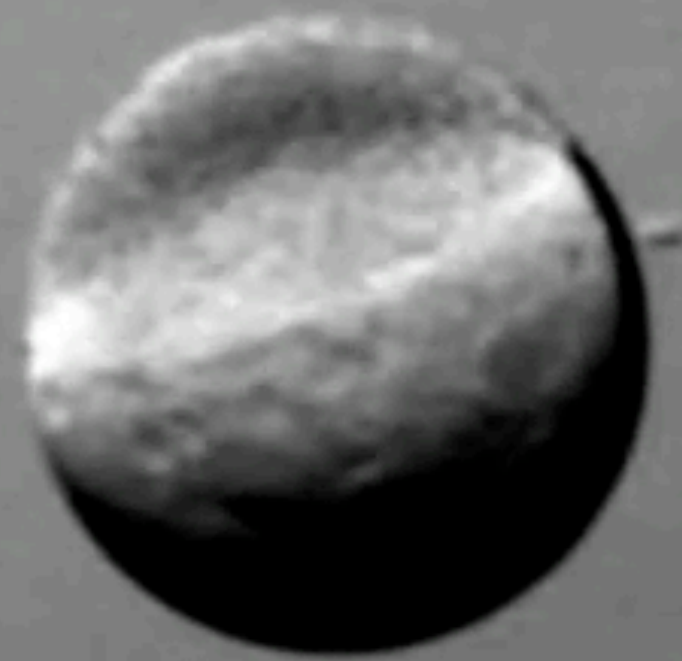
# How is symmetry broken?



Oocytes: spherical?



Zygote: radial

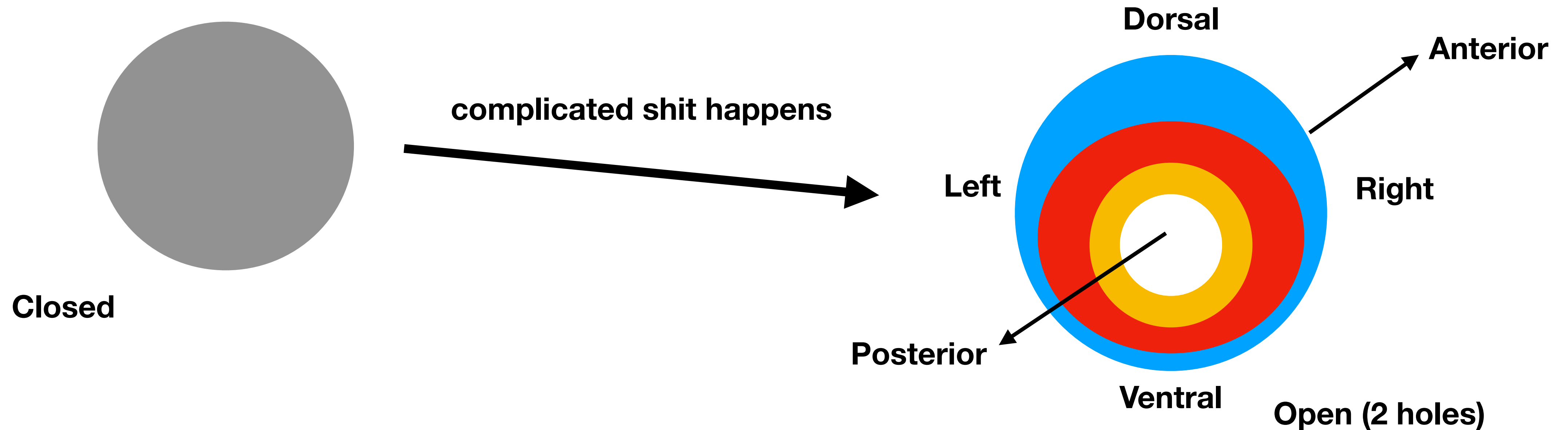


Larva (adult body plan):  
head/tail, front/back, left/right

04:24:00

1. Differentiation of three\* germ layers;
2. Gastrulation moves cells *inside* embryo;
3. Germ layers are patterned along 3 axes

**Blastoderm:** compact mass of pluripotent cells



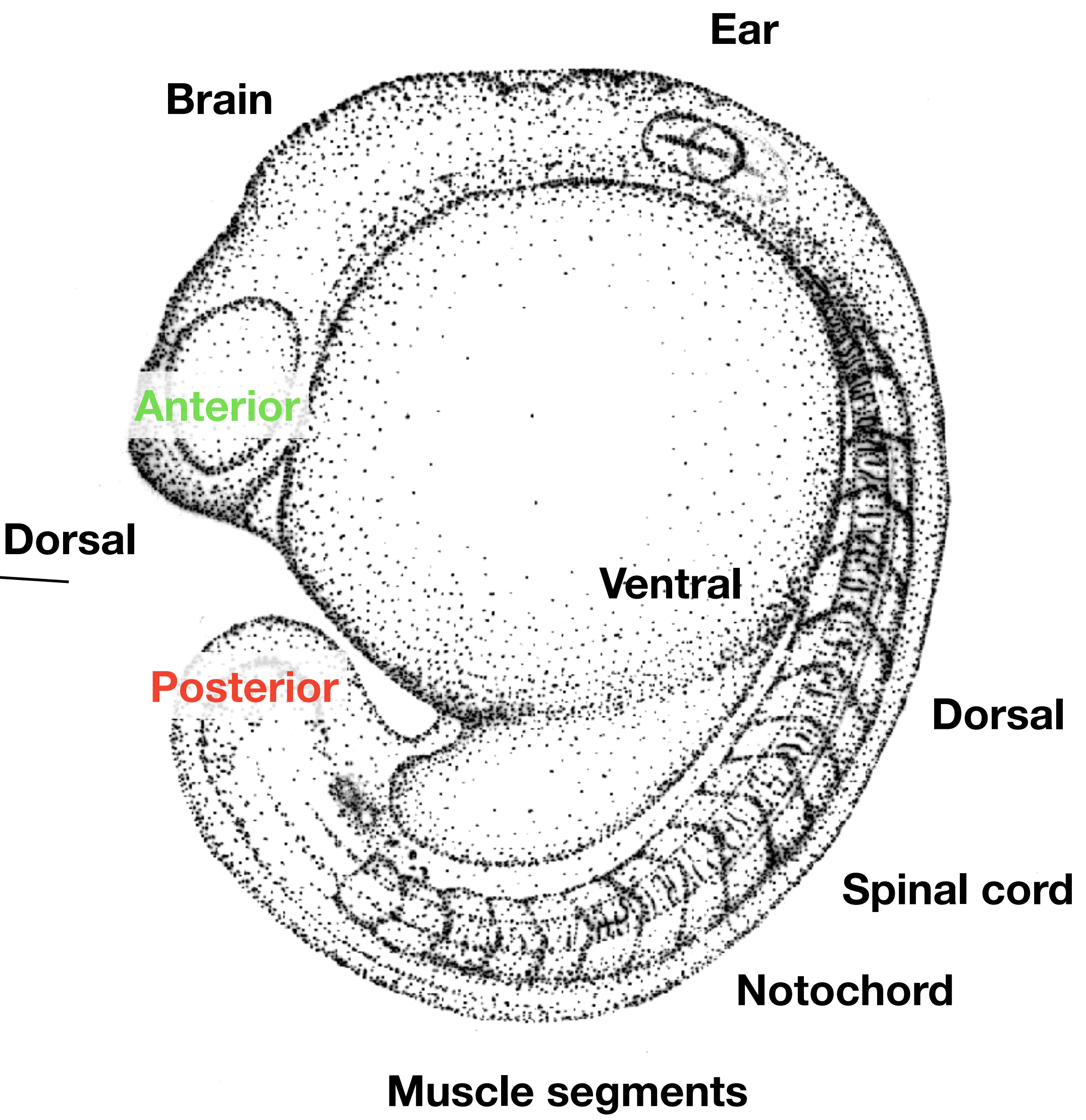
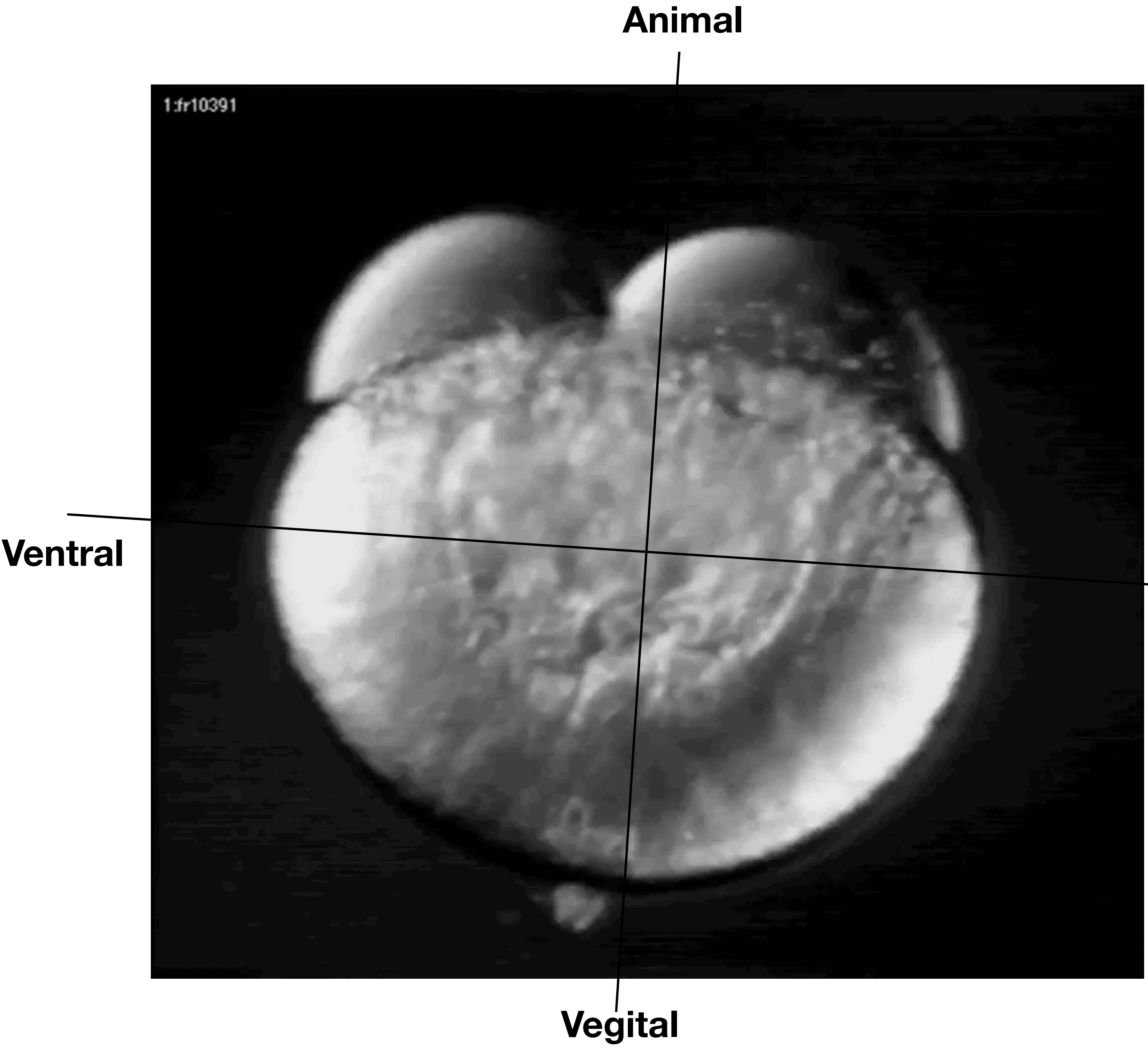
**Ectoderm:** nervous system, skin, sensory organs

**Mesoderm:** muscle, bone, kidneys, vasculature, blood

**(mes)Endoderm:** gut, liver, pancreas

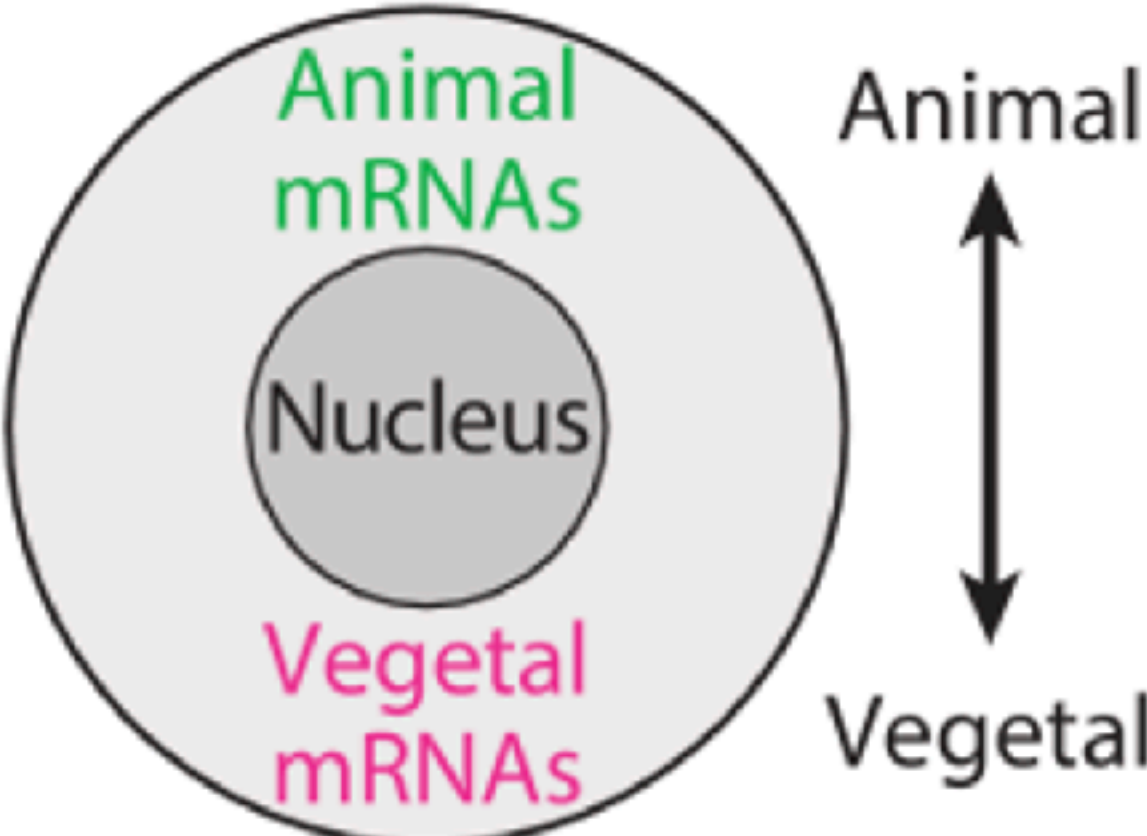
**Yolk: food!**

# Zebrafish early development: 2-cell to 18 somites, ~18 hours

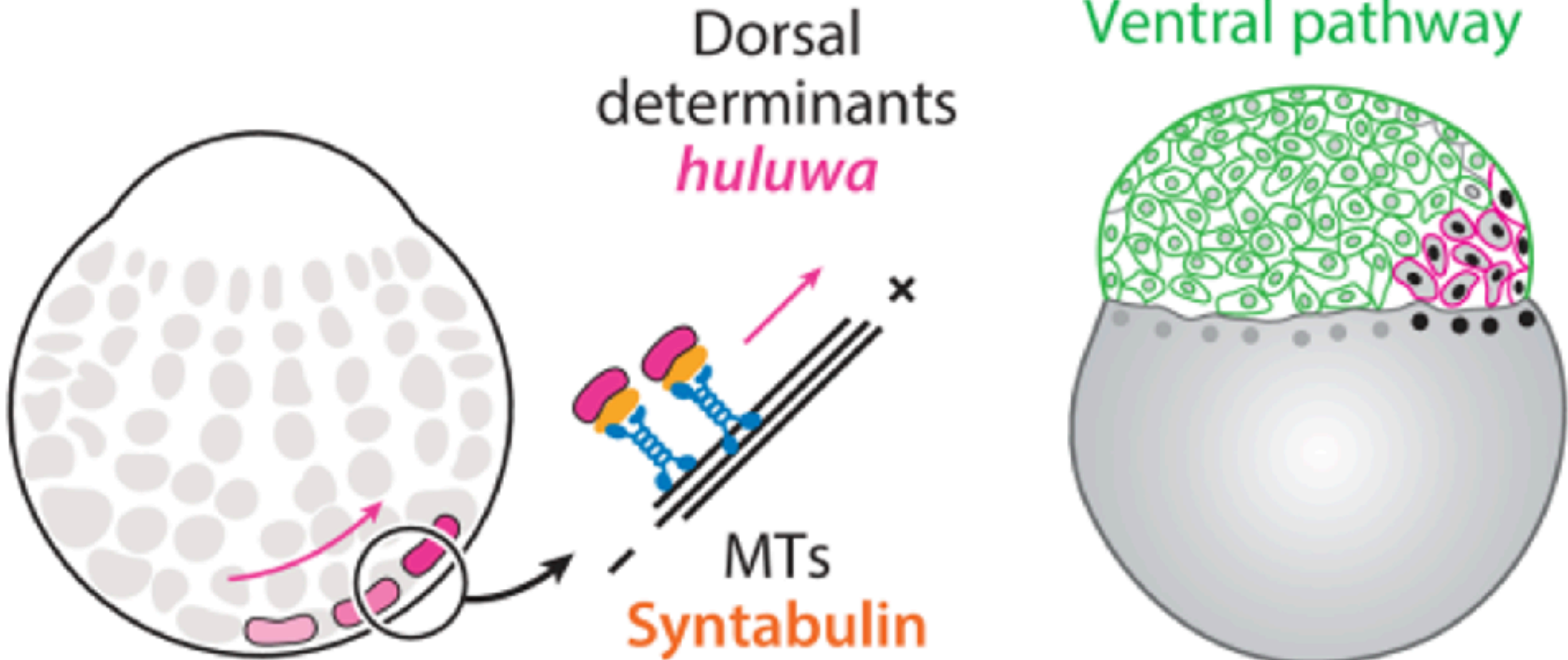


# Early symmetry-breaking

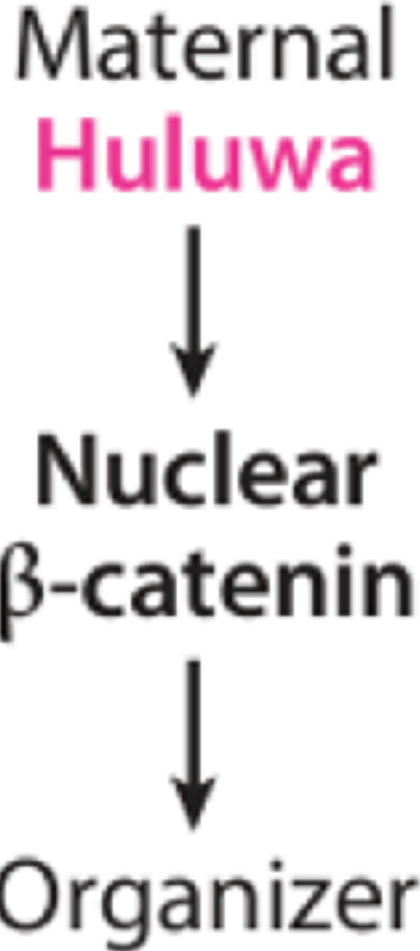
**Oocytes:  
spherical but patterned**



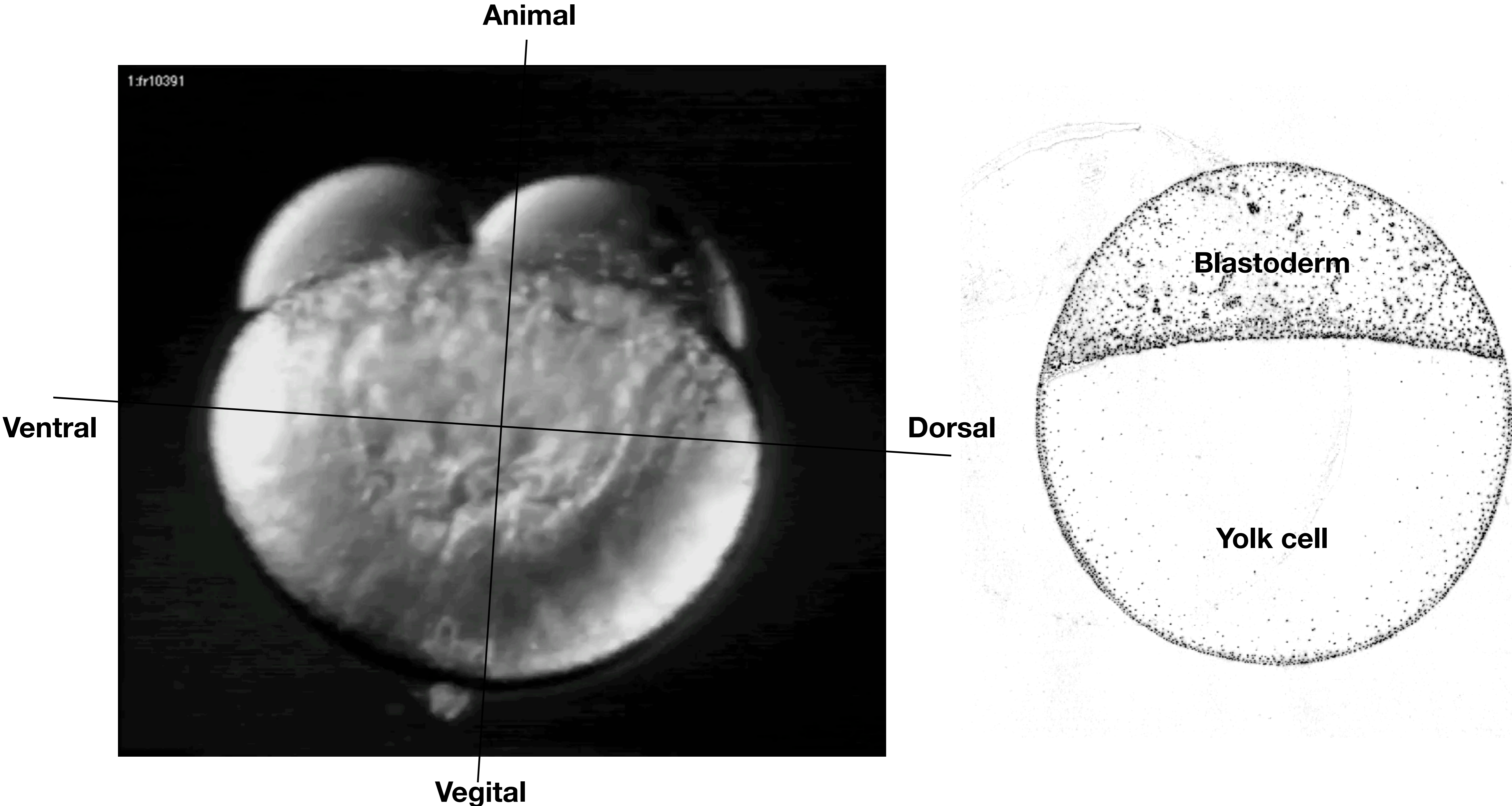
**Animal-vegetal polarity**



**Asymmetric allocation of dorsal determinants**

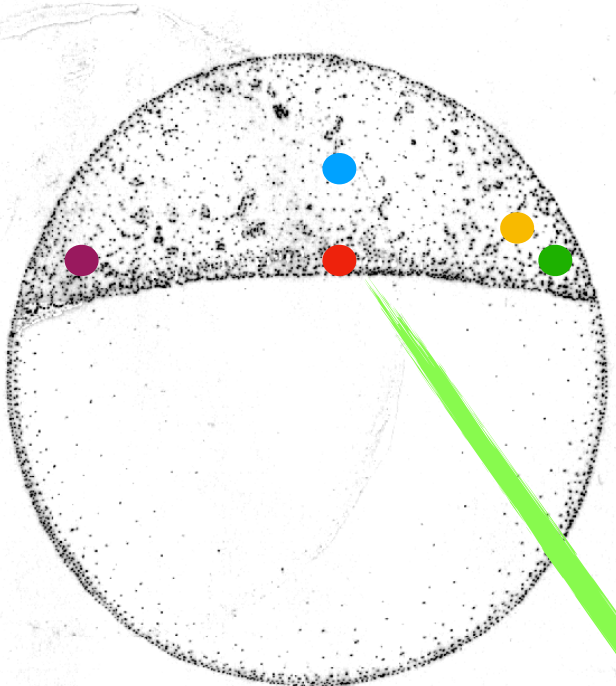


# Zebrafish early development: cleavage stage $\Rightarrow$ sphere

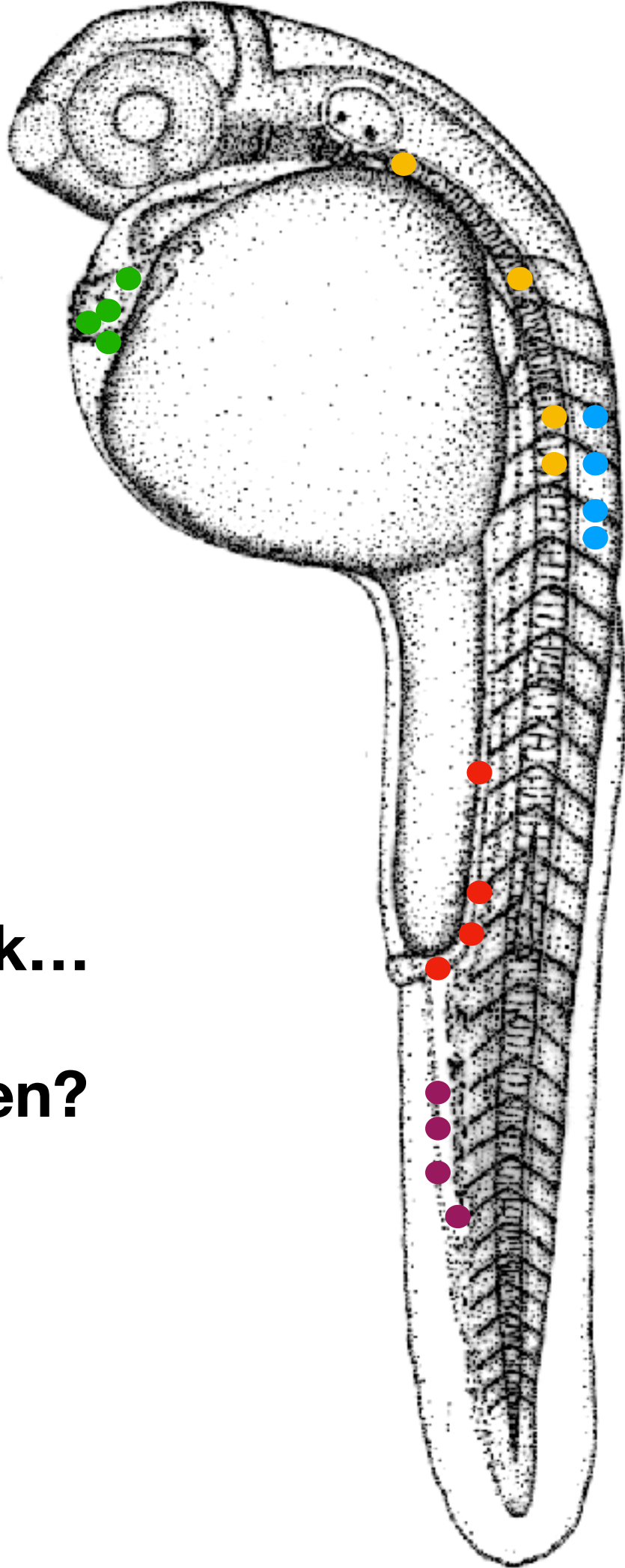


# Fate map and lineage of the early zebrafish

Sphere-stage  
Pre-gastrulation

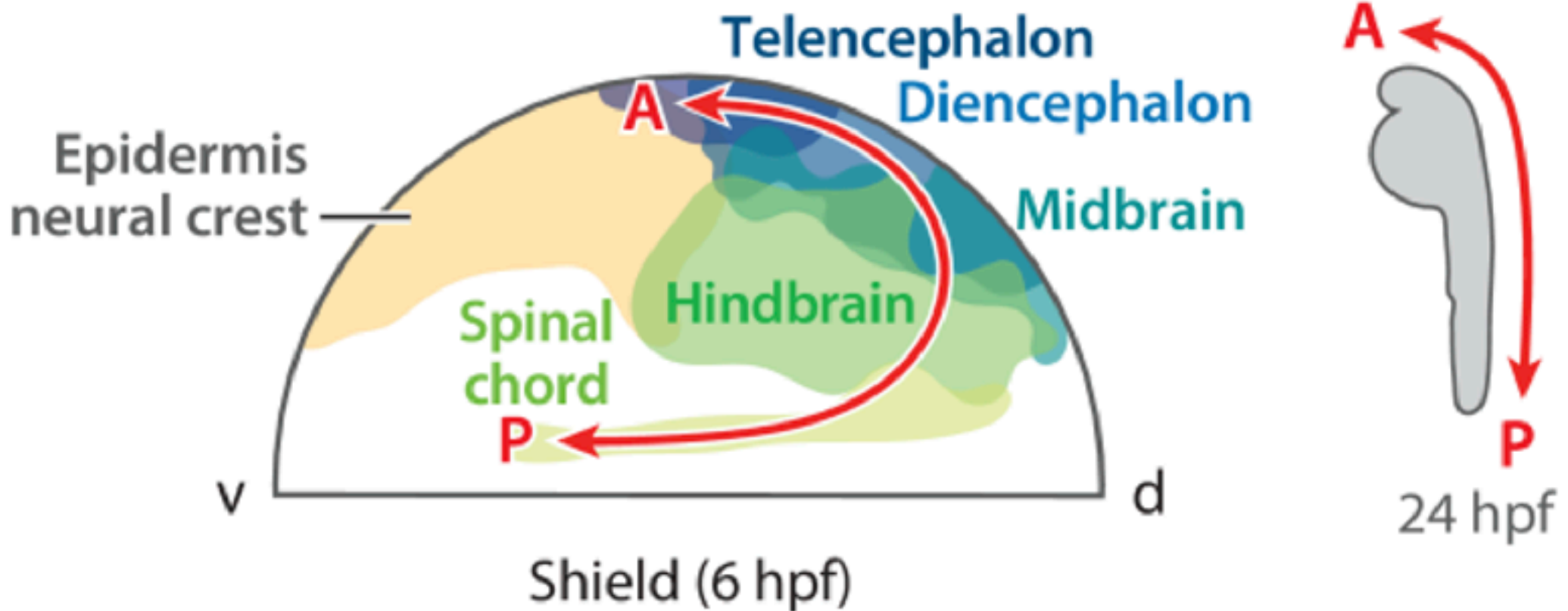
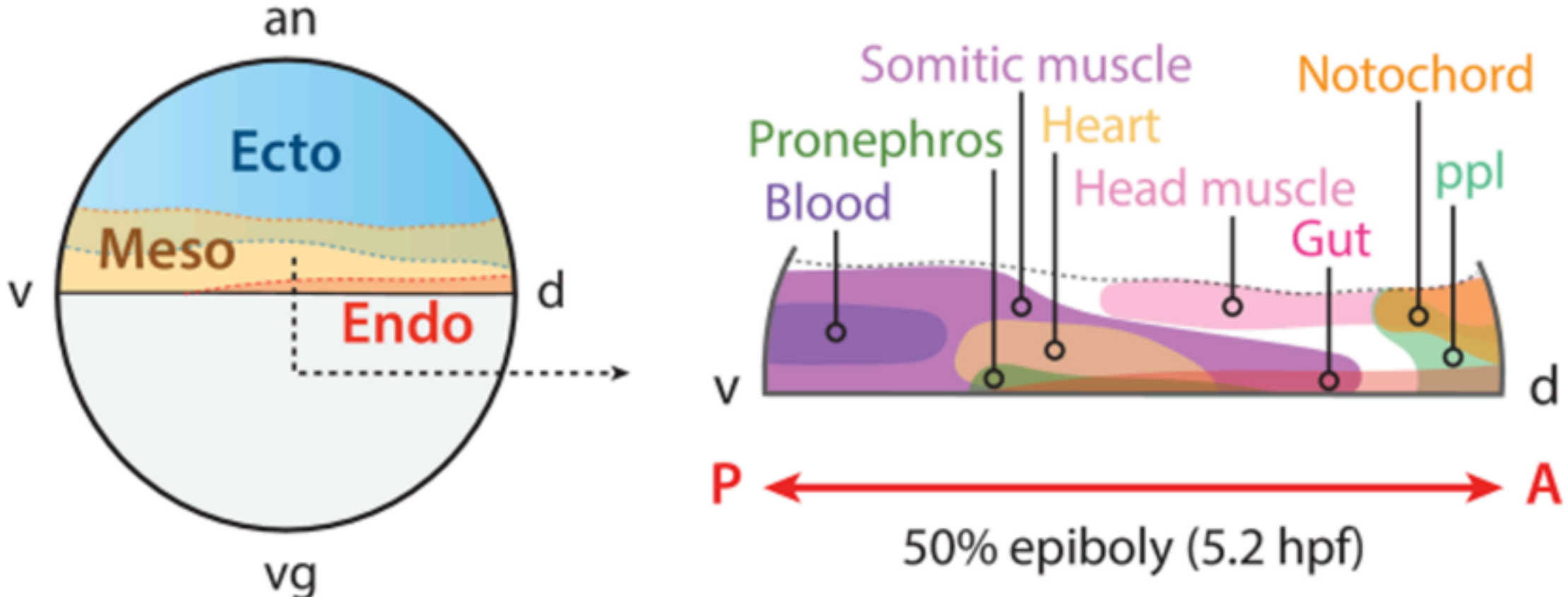


Time

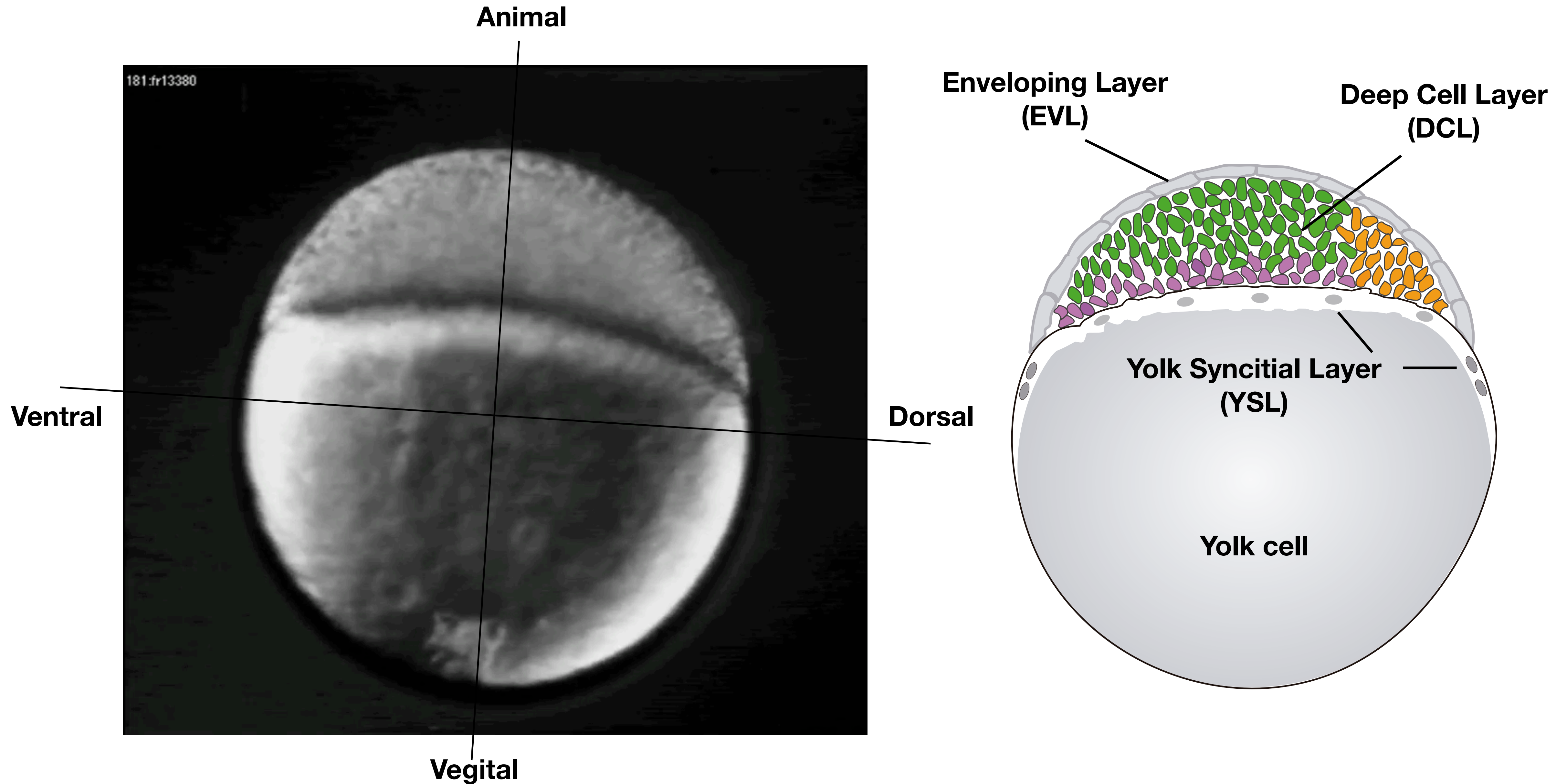


Fluorescent molecule, beads, ink...

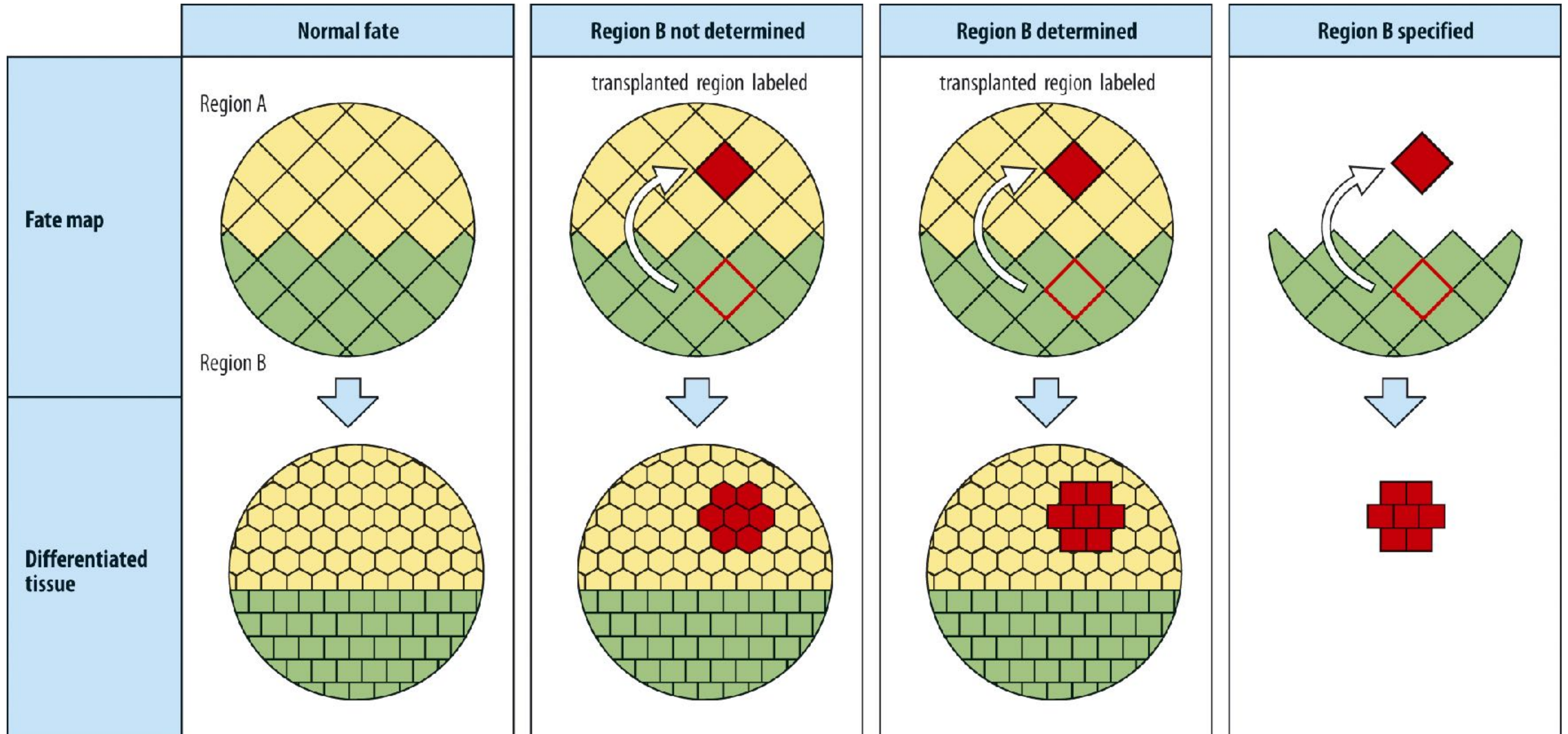
Labelled when? Grown until when?



# Zebrafish blastoderm well-patterned before gastrulation

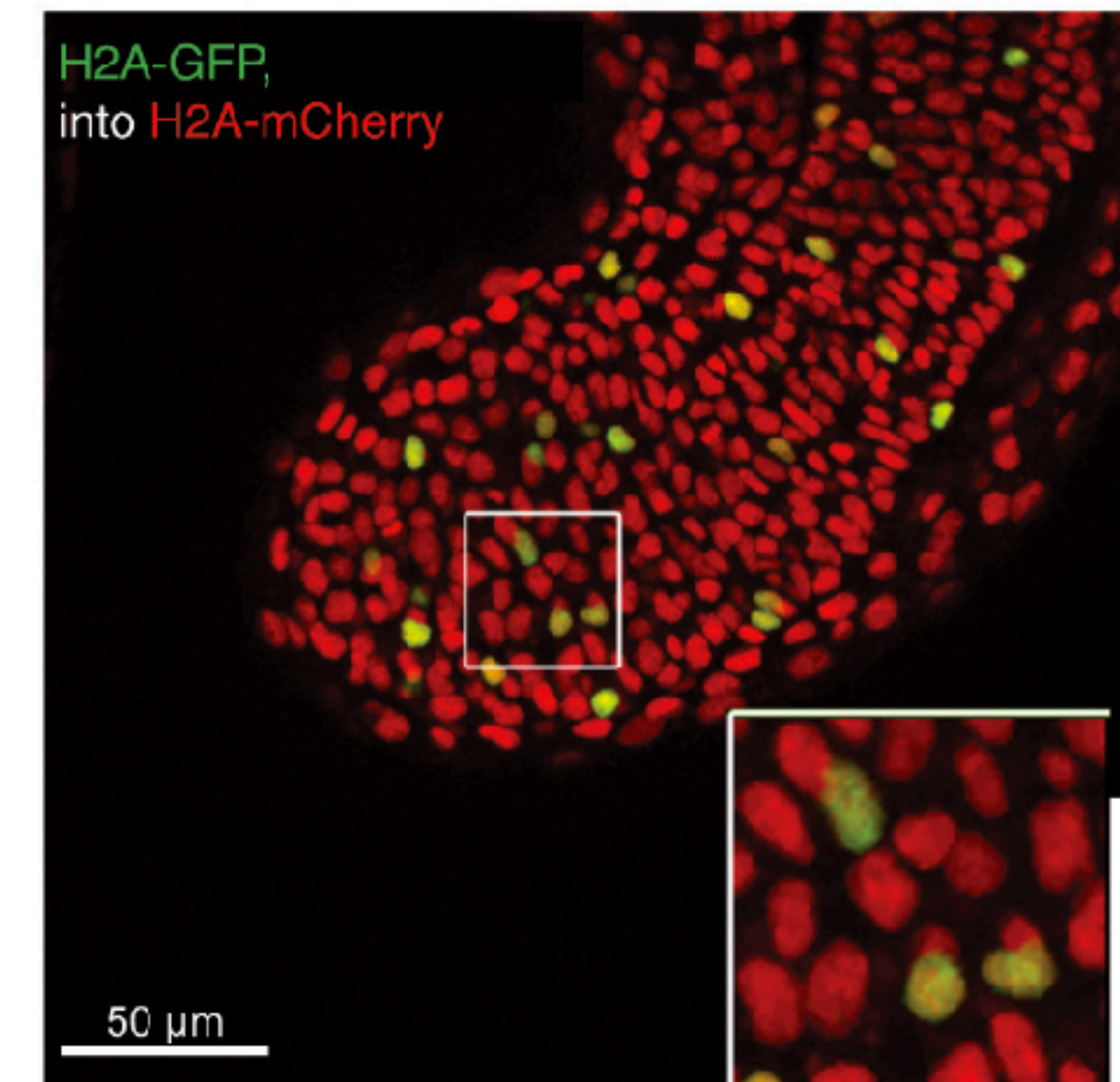
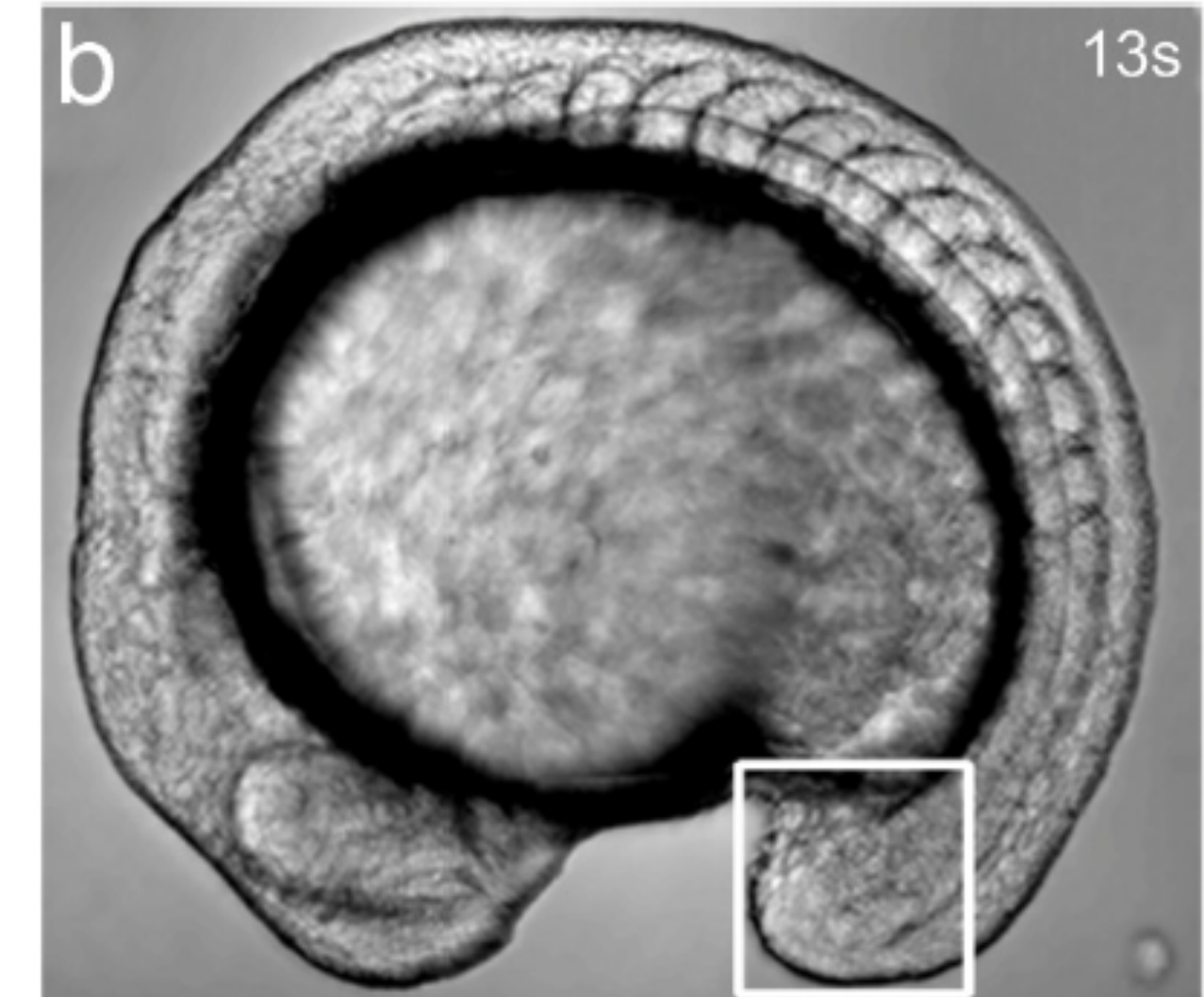
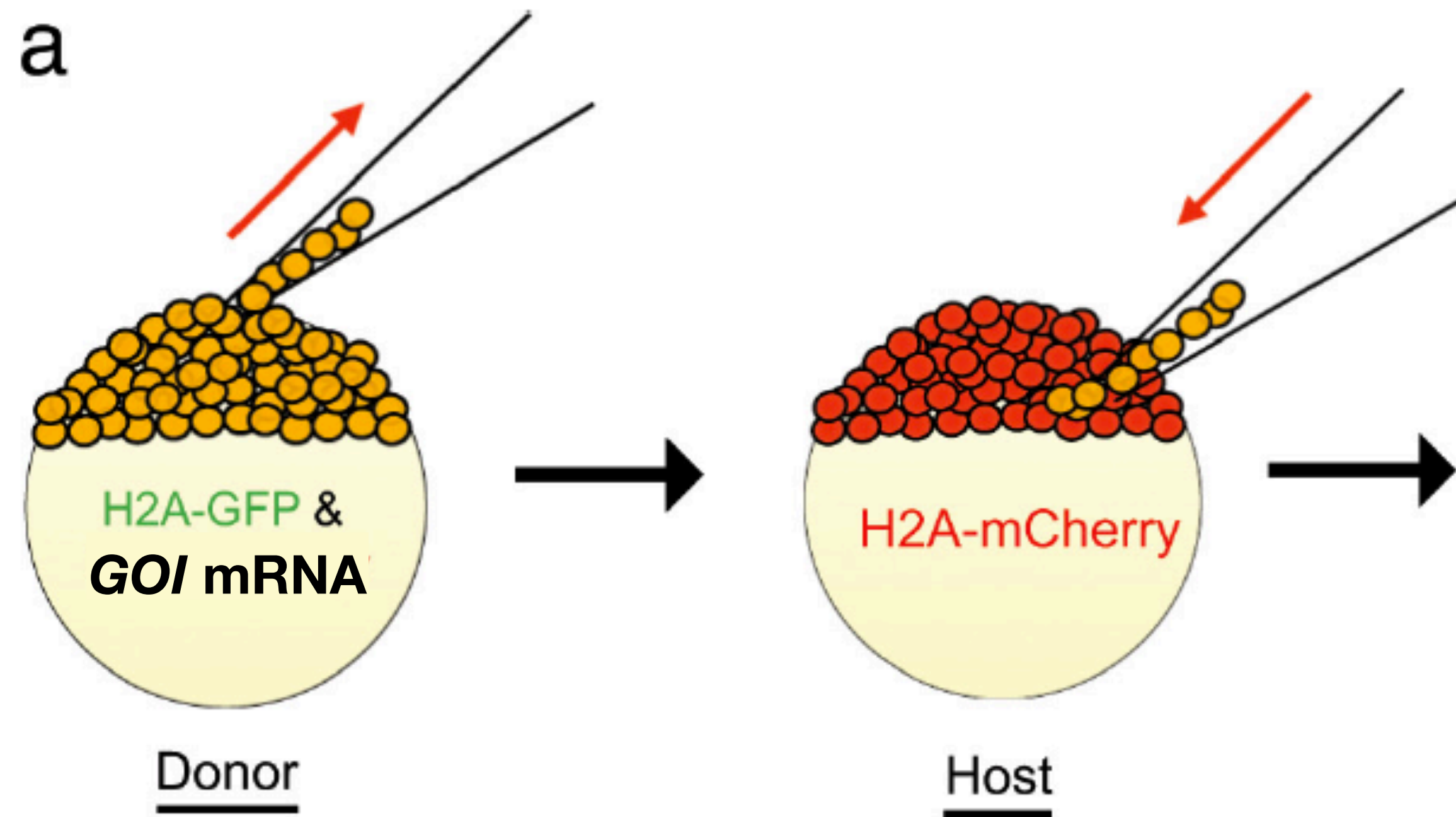


# Fate maps, specification and determination



Green/yellow: geographical location; Red: labelled cells; Shape: cell type

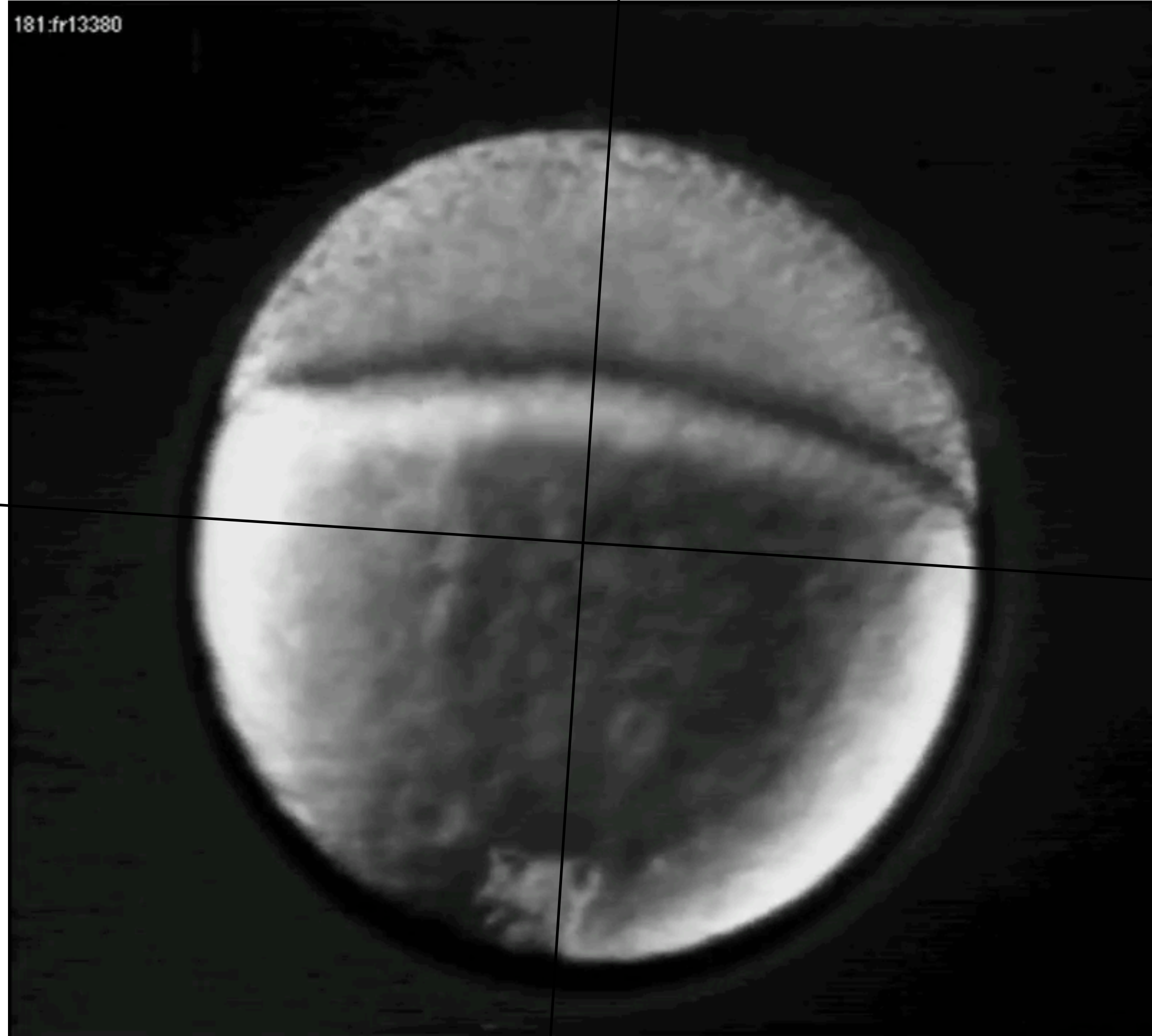
# Transplantation of cells at sphere stage



# Zebrafish early development: sphere $\Rightarrow$ shield

Animal

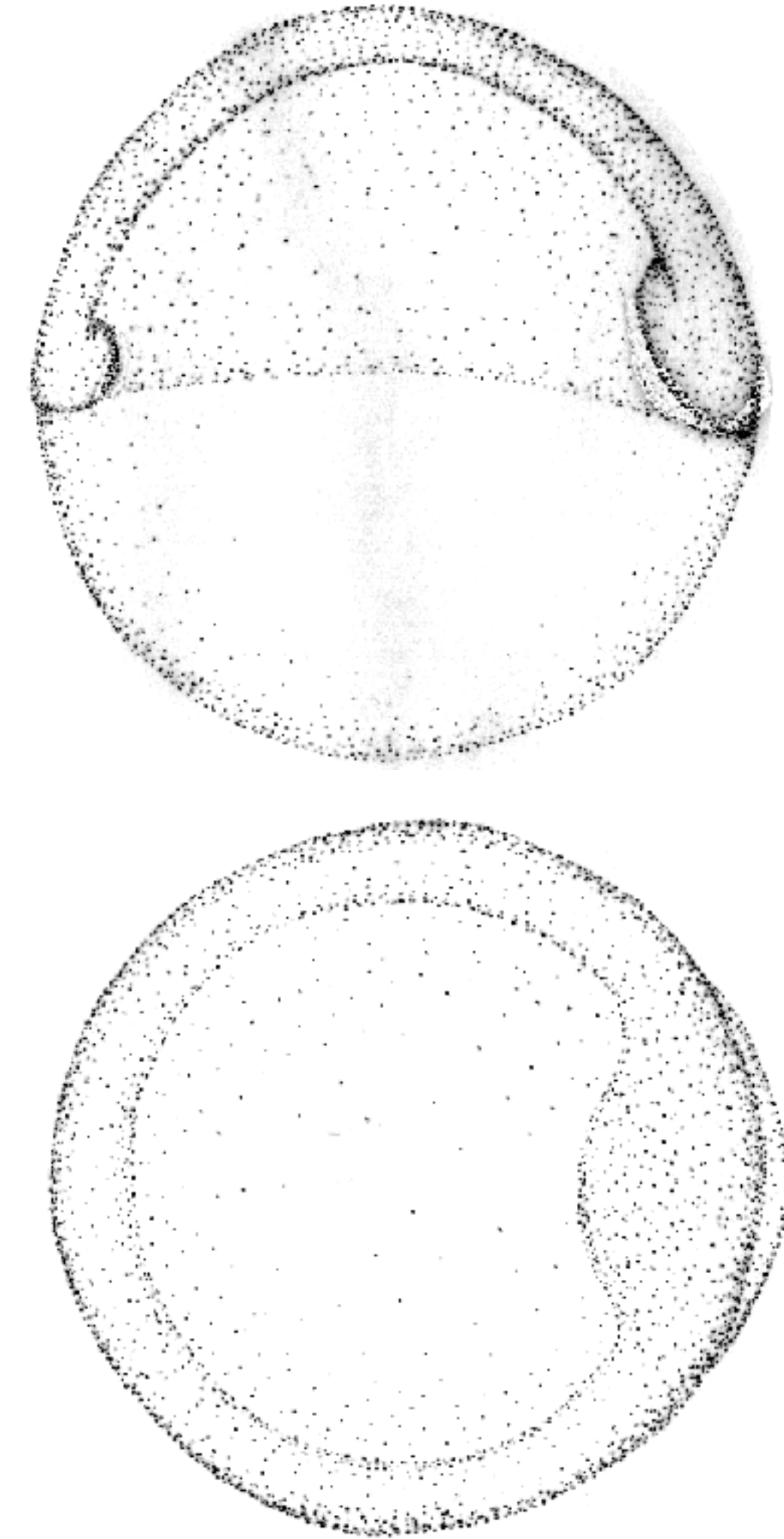
181.fr13380



Ventral

Dorsal

Vegital

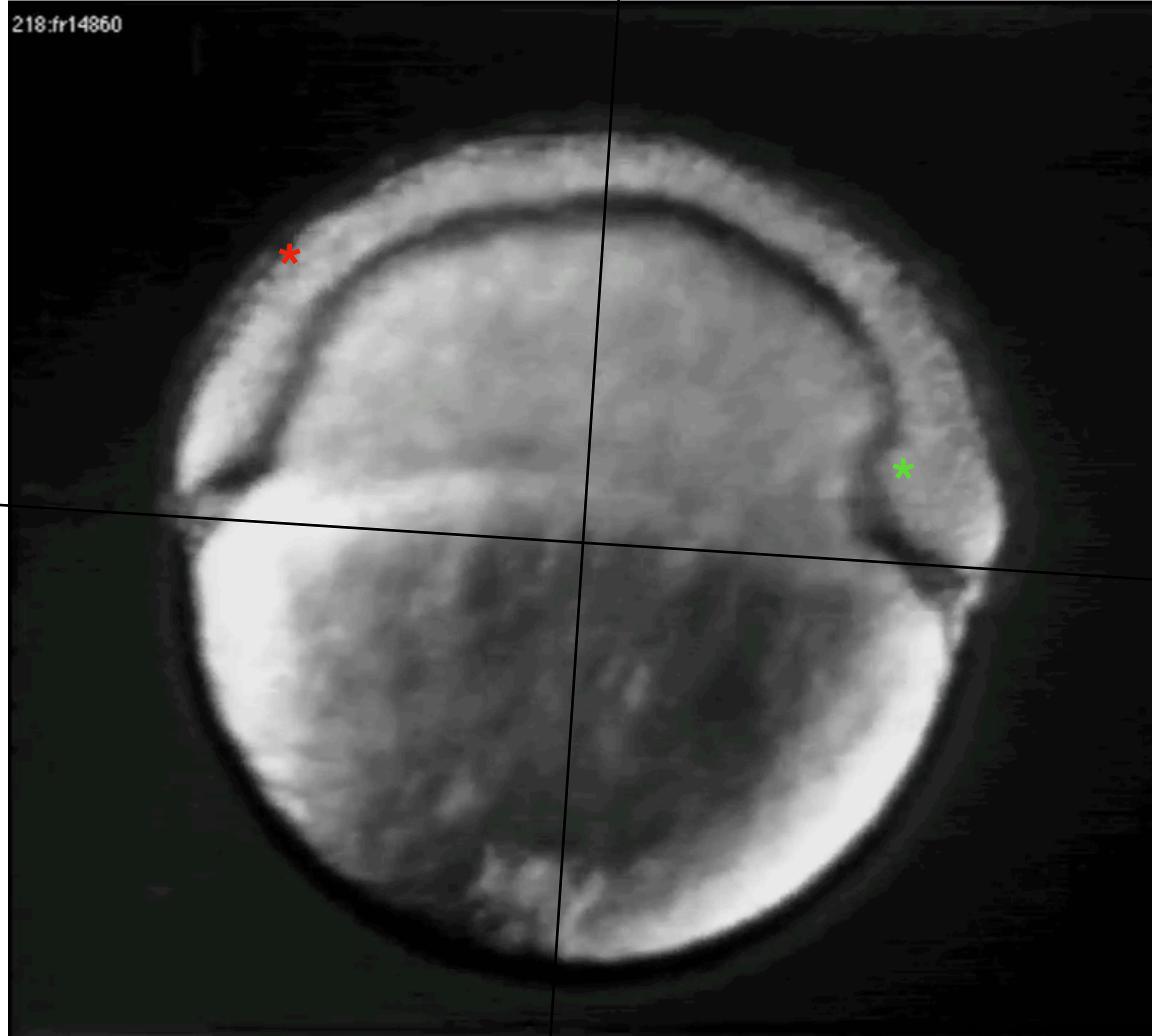


**First morphological asymmetry,  
onset of gastrulation**

# The dorsal mesoderm moves up to the anterior

Animal

218:fr14860

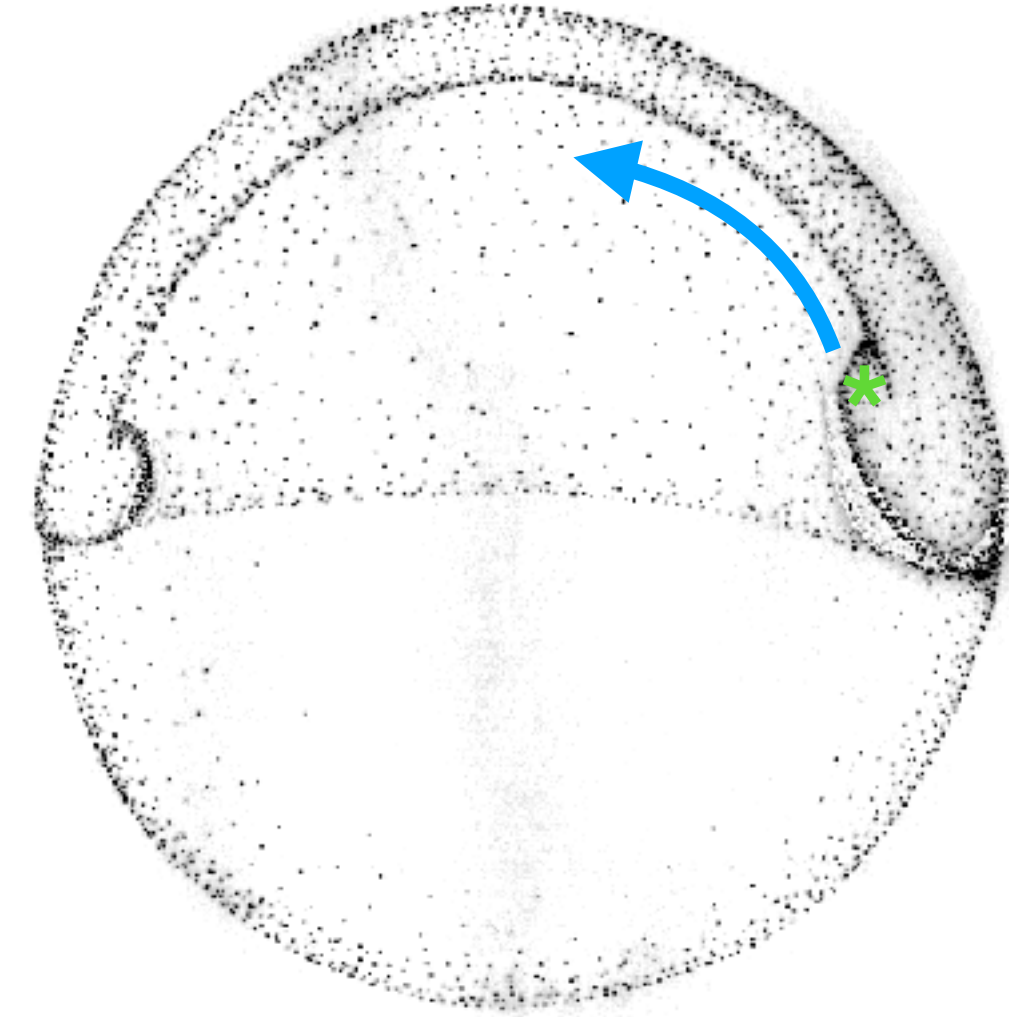


Ventral

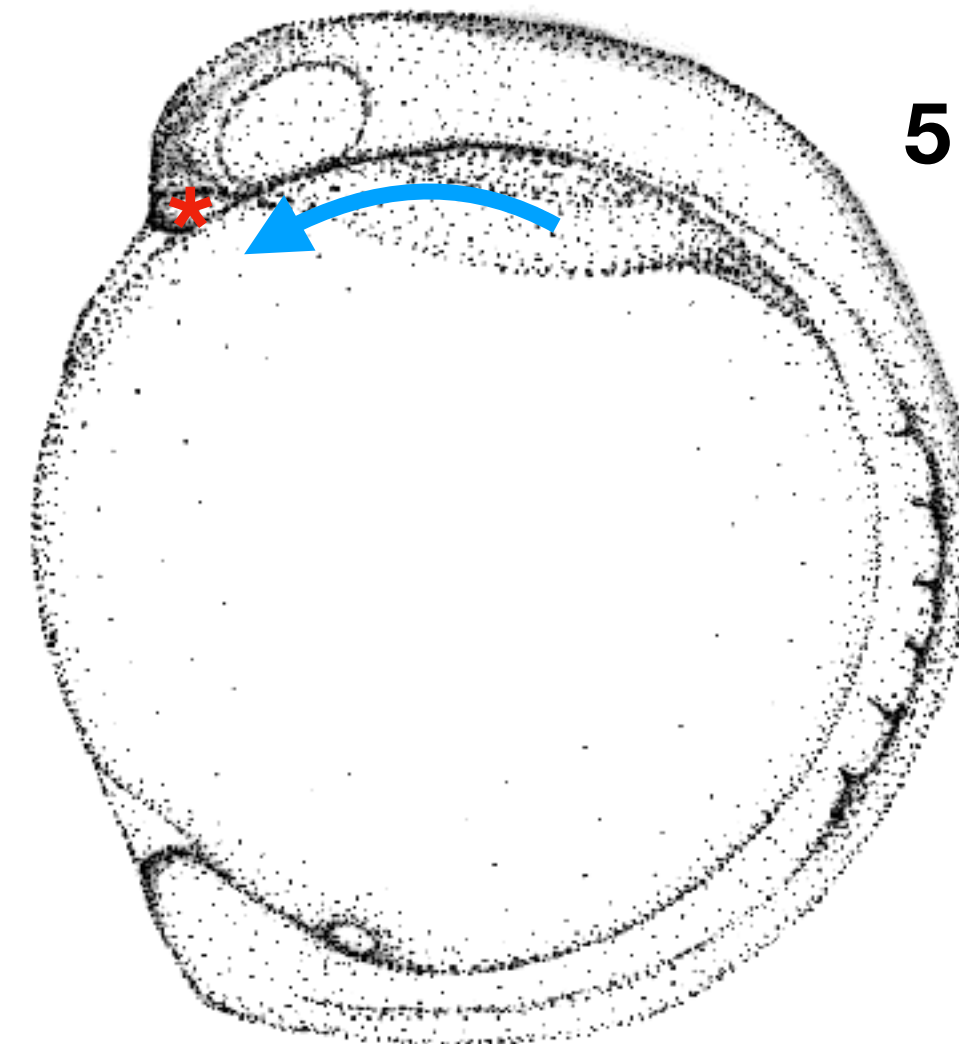
Dorsal

Vegital

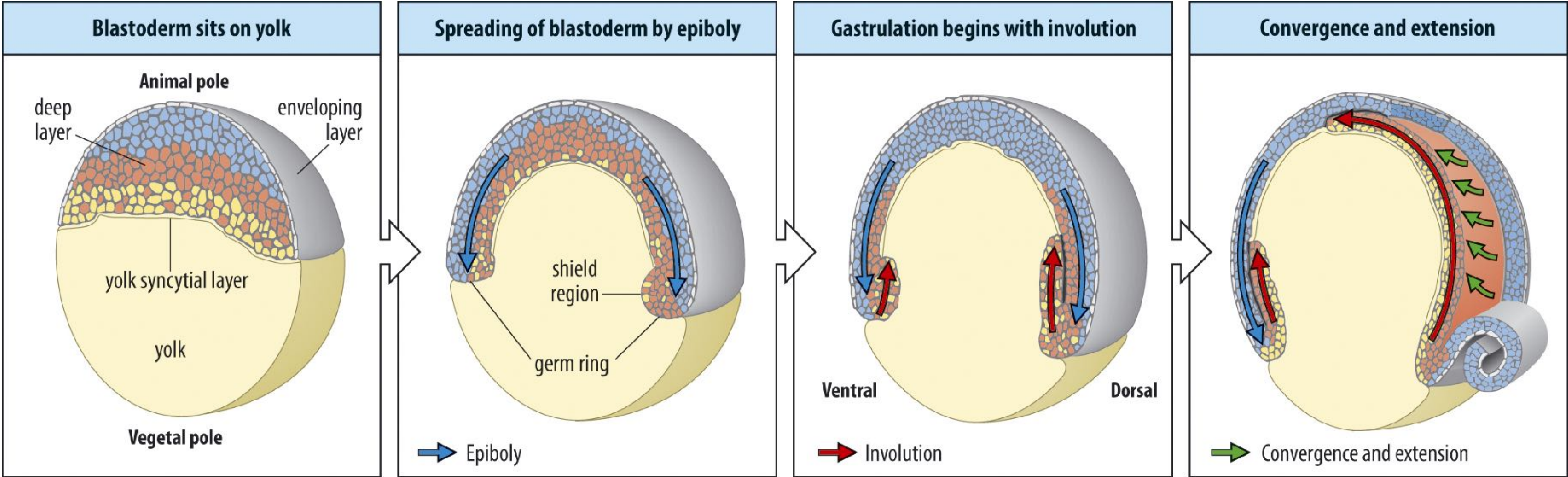
sheild stage



5 somite stage

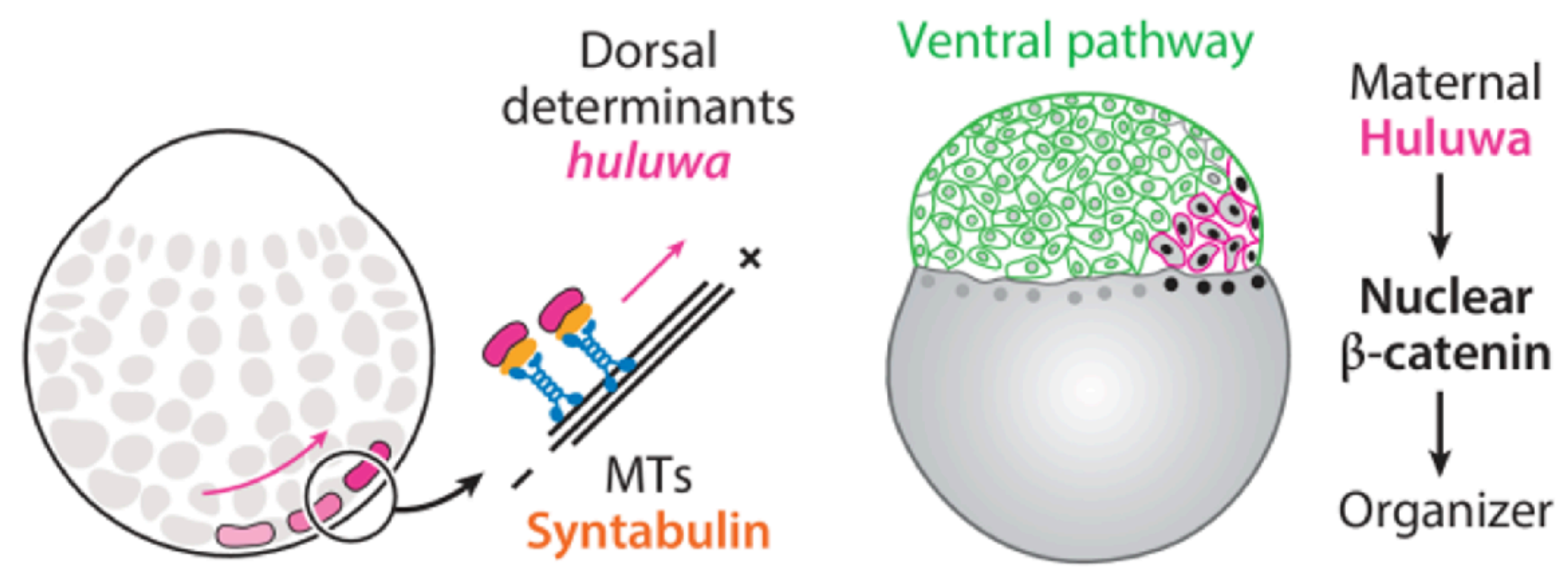


# What starts out as Dorsal cells/tissue moves to the Anterior



DV and AP axes are tightly coupled

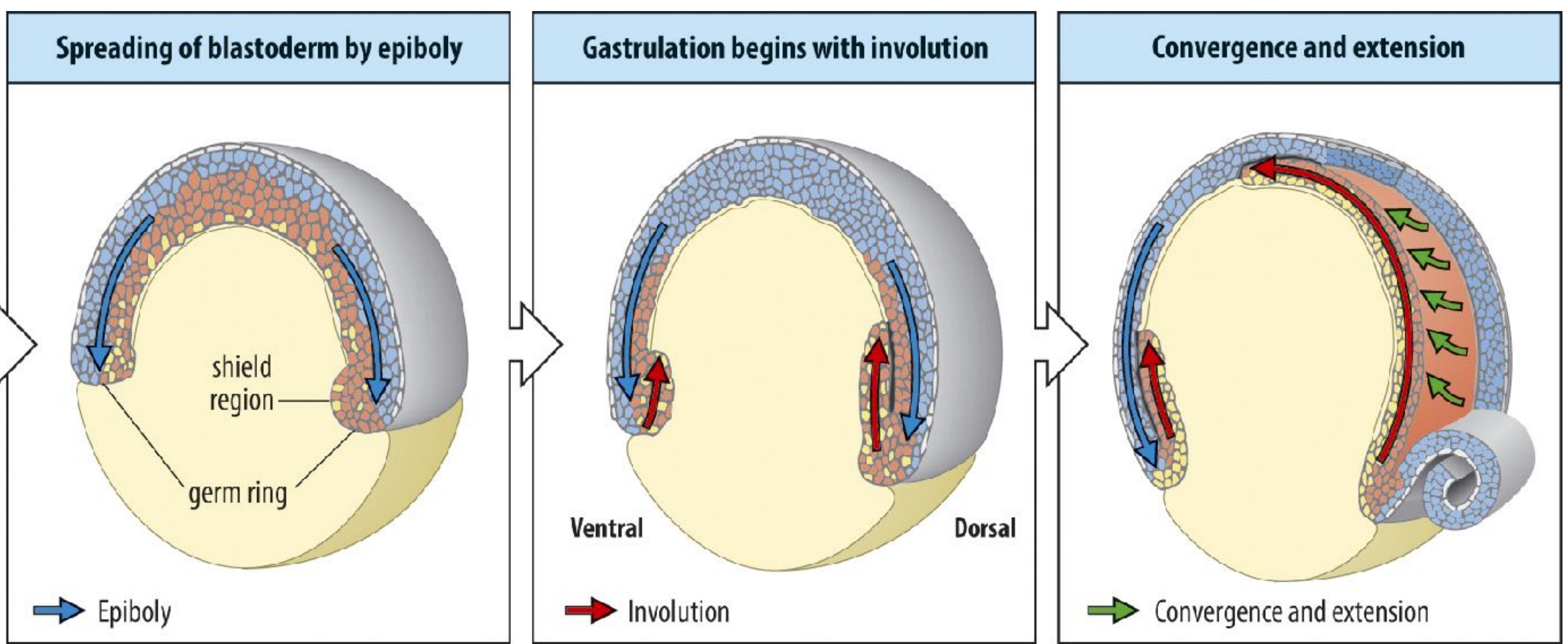
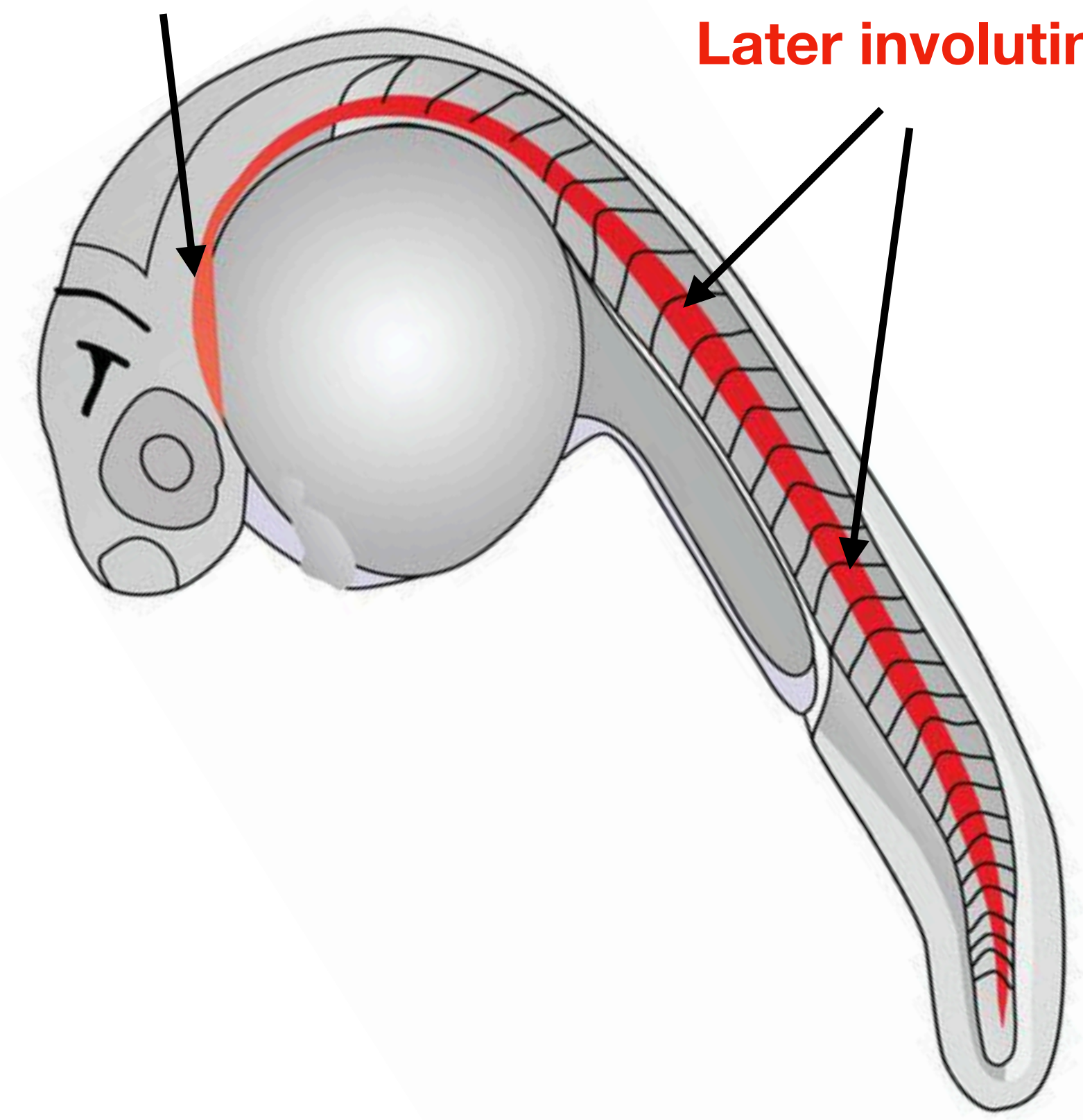
# Dorsal maps to Anterior! Shield makes notochord!



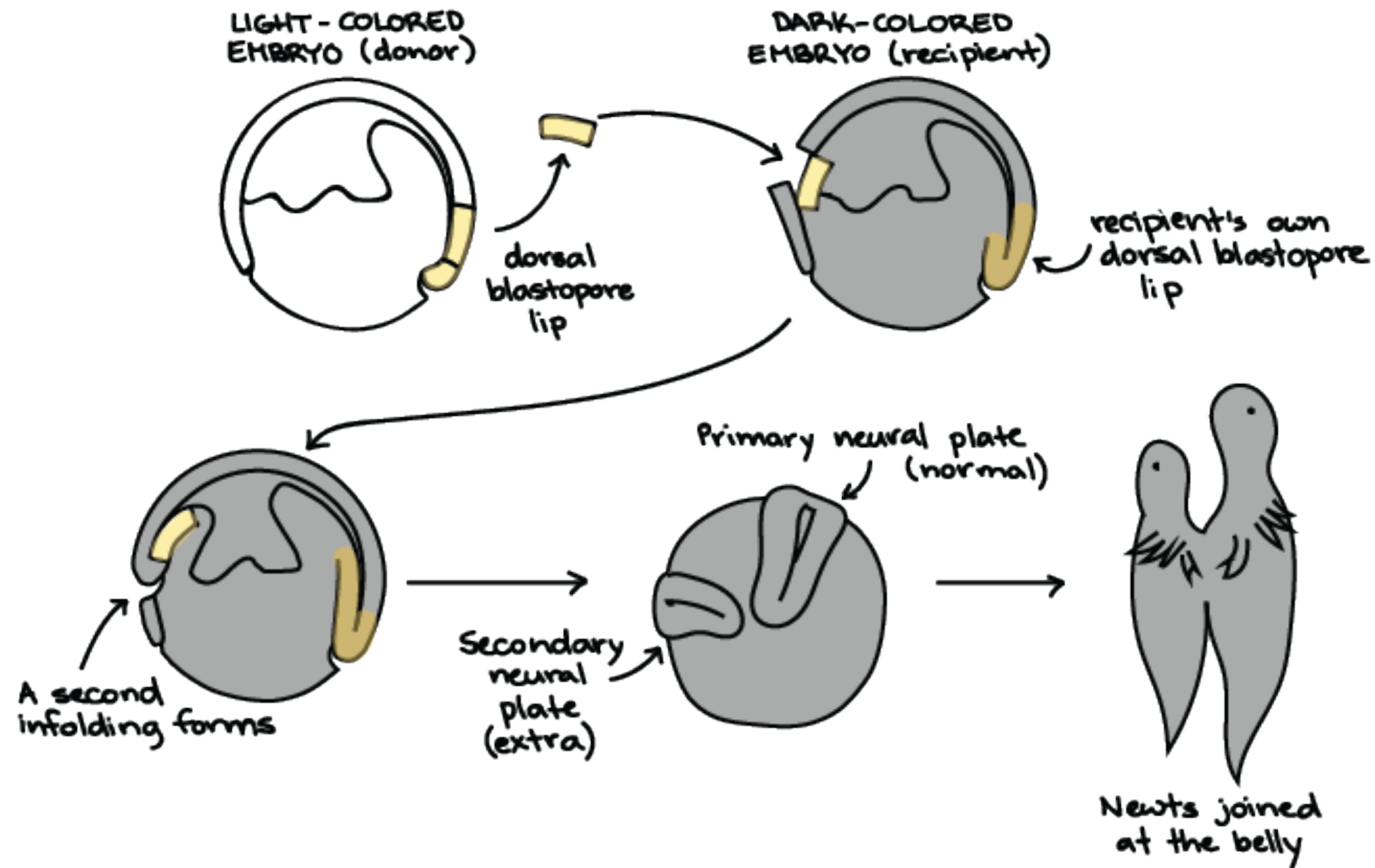
Asymmetric allocation of dorsal determinants

Early involuting cells

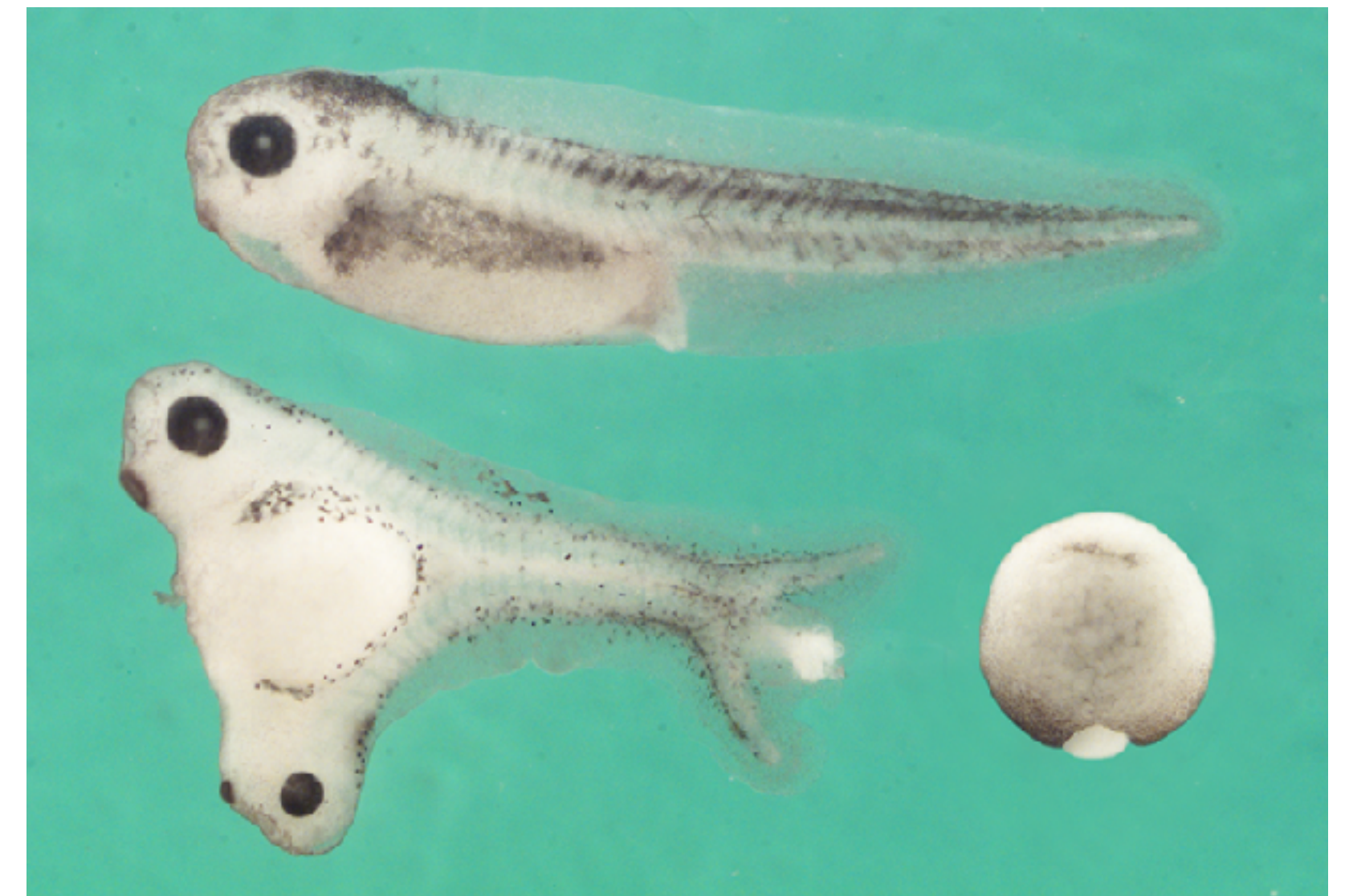
Later involuting cells



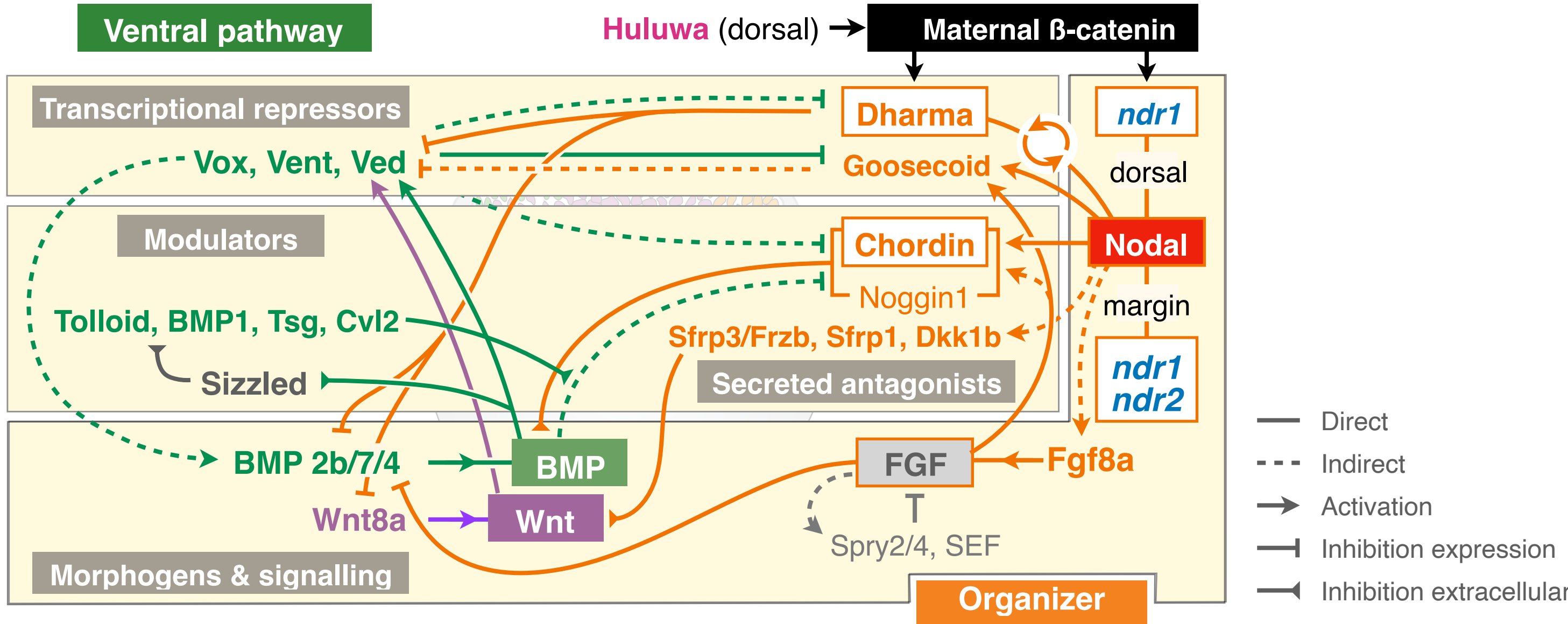
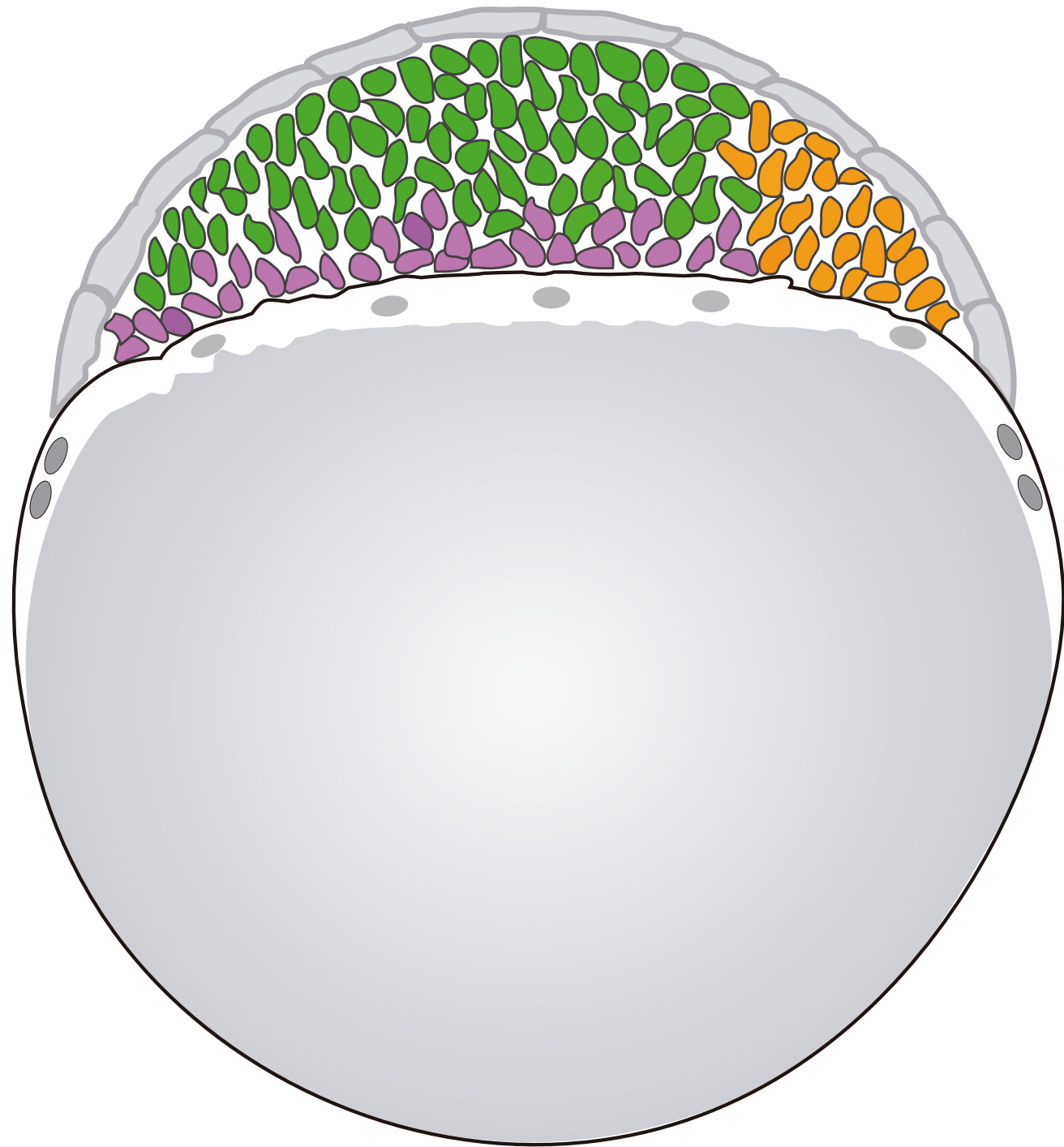
# **Embryonic Induction:** Spemann and Mangold, 1924



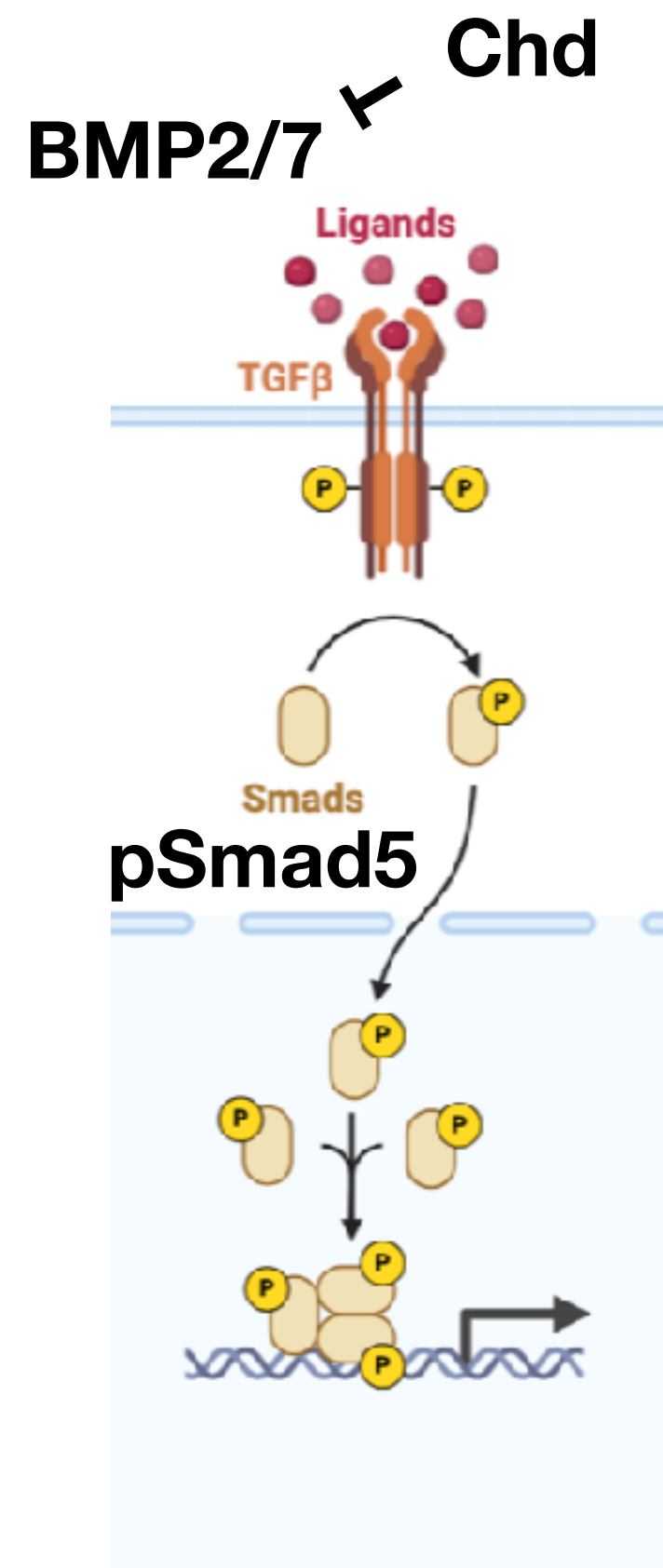
A signal from the light-coloured embryo instructed the dark-coloured embryo's cells to change their behaviour: "organizer" of a new blastopore



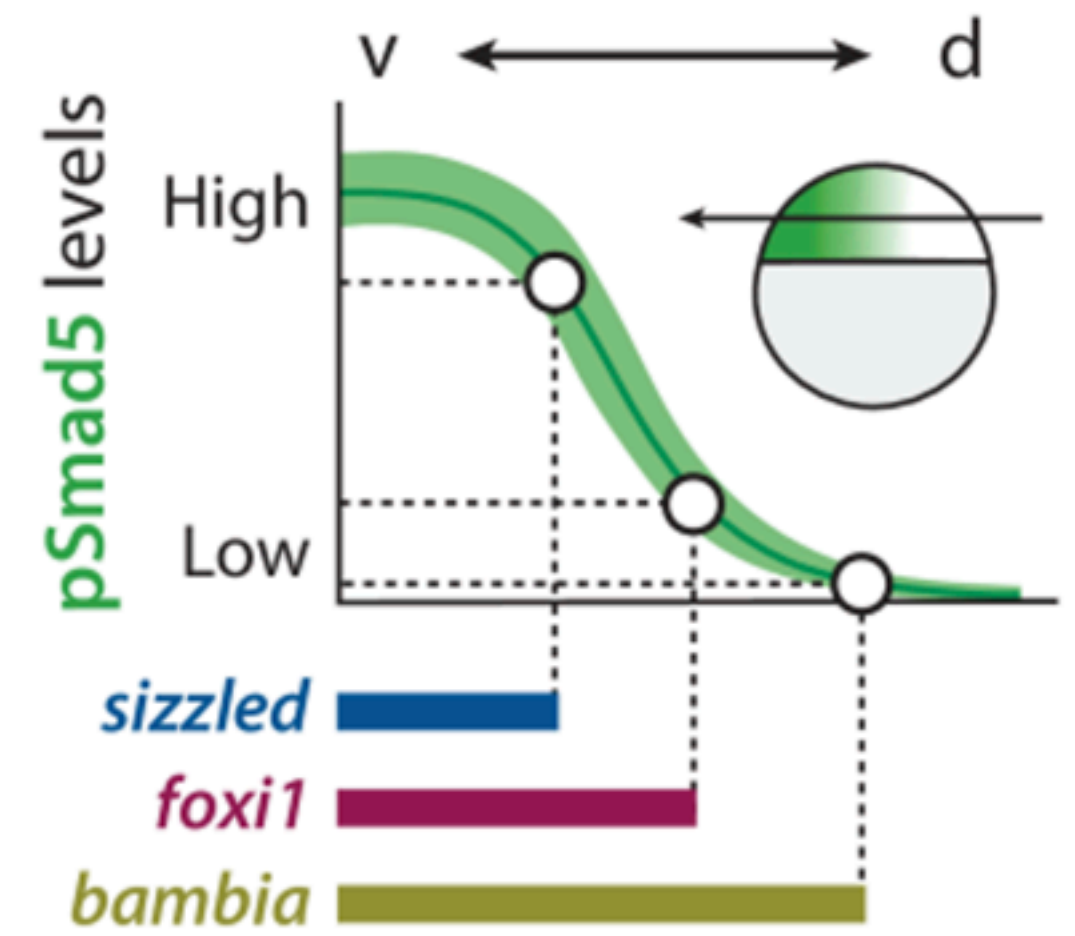
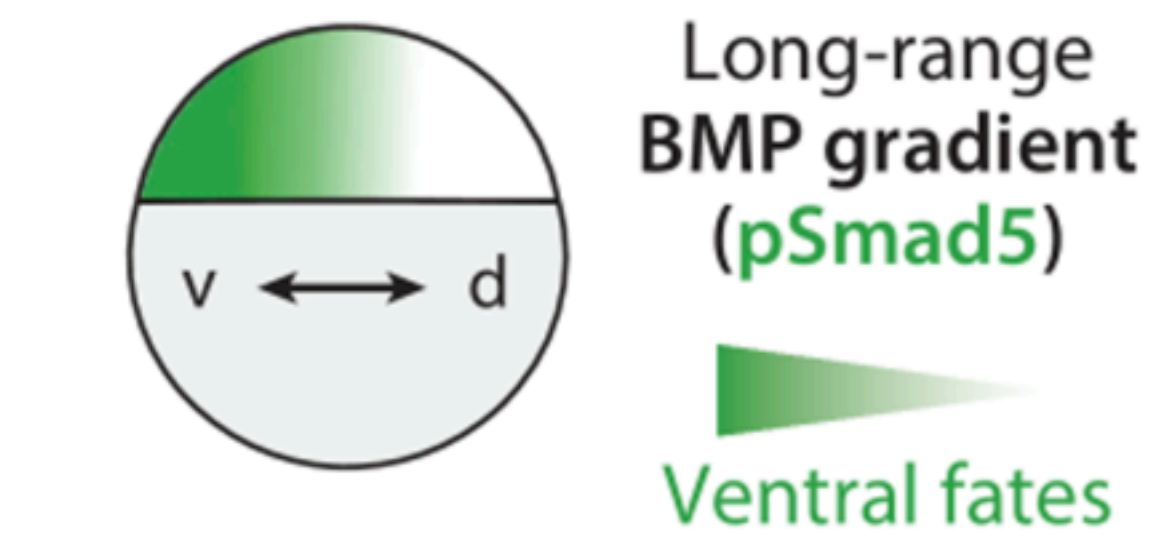
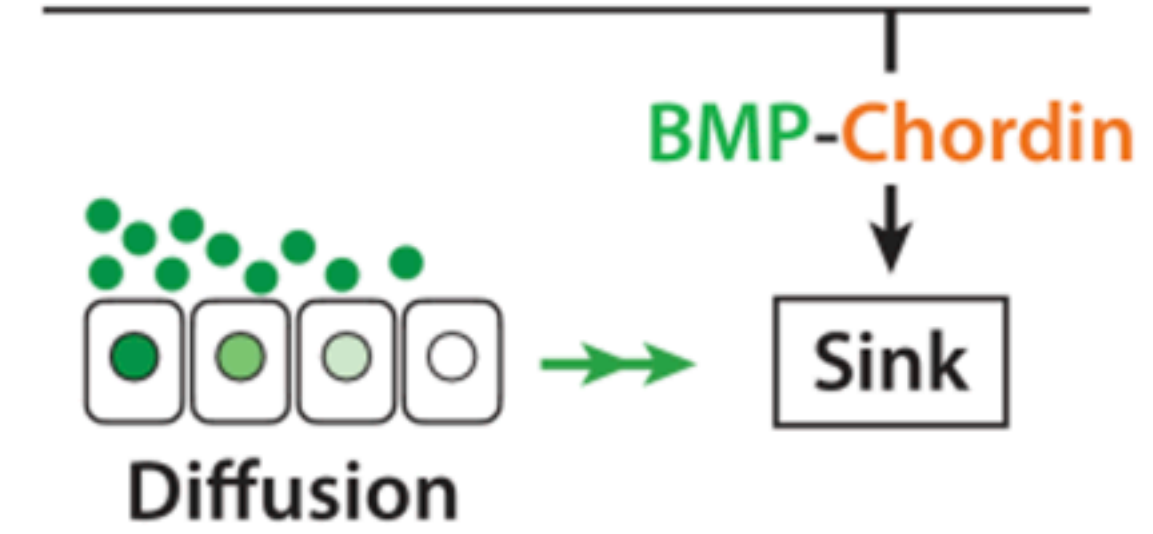
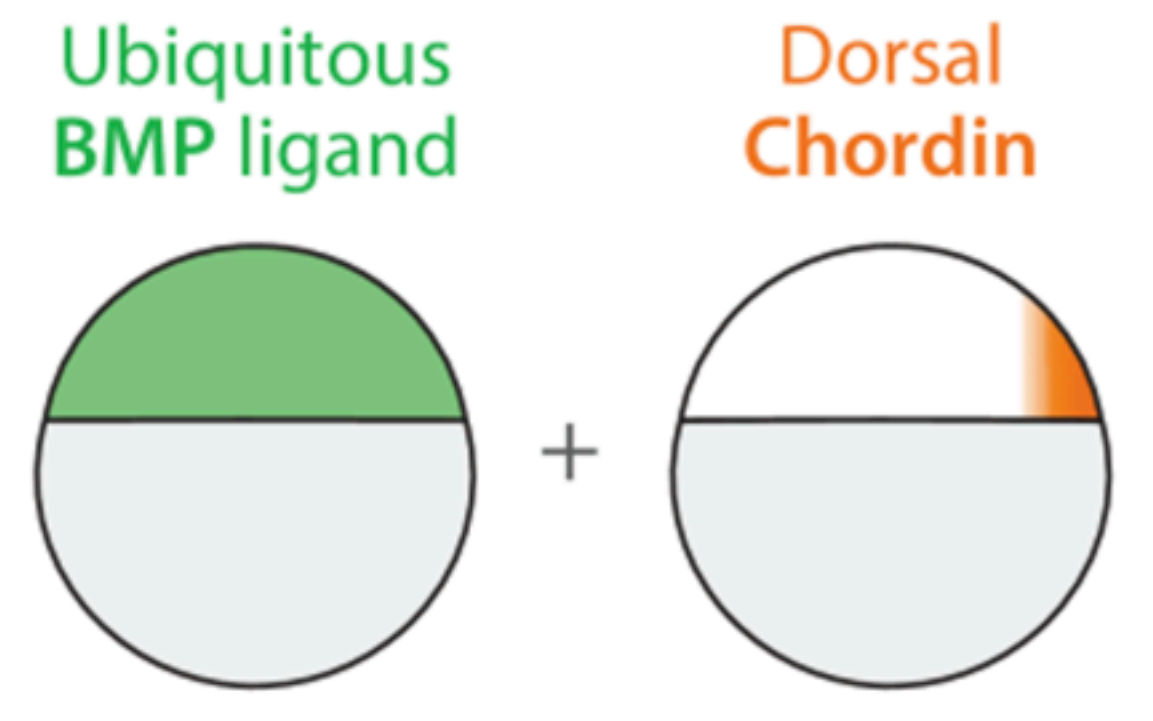
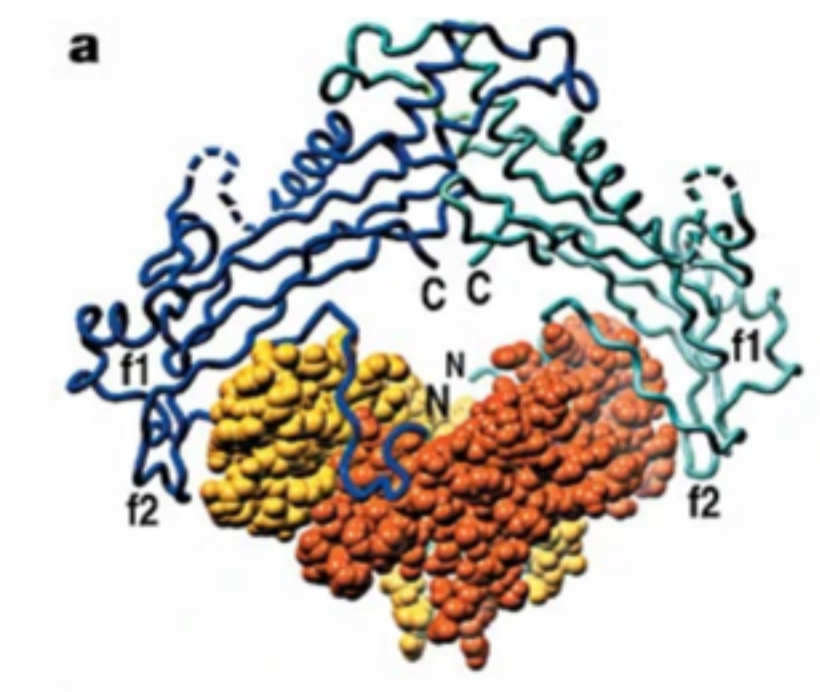
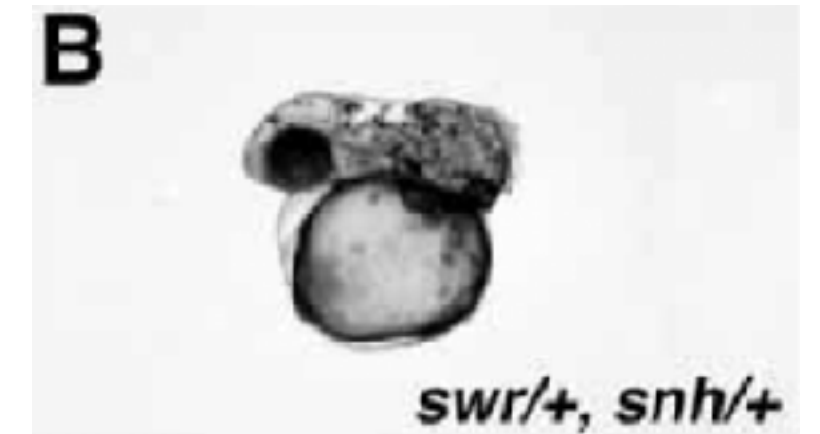
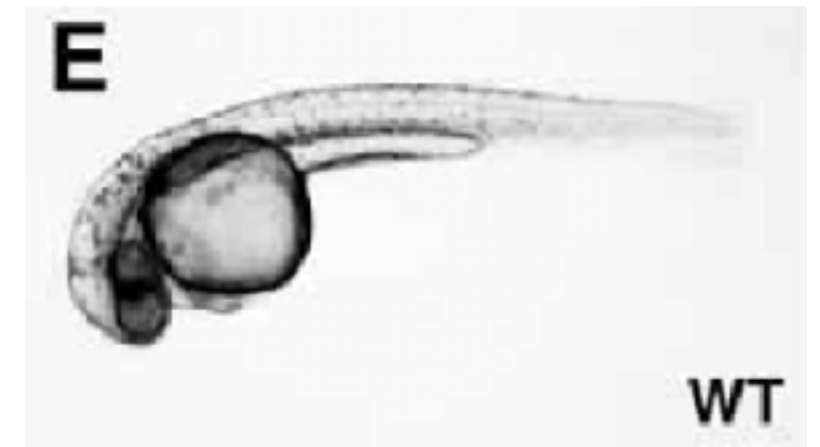
# Organizer signaling in the zebrafish



# Organizer signaling: dorsal Chordin blocks BMPs



**TGFβ receptors**  
 Activate Smad transcription factors in the cytosol by phosphorylation

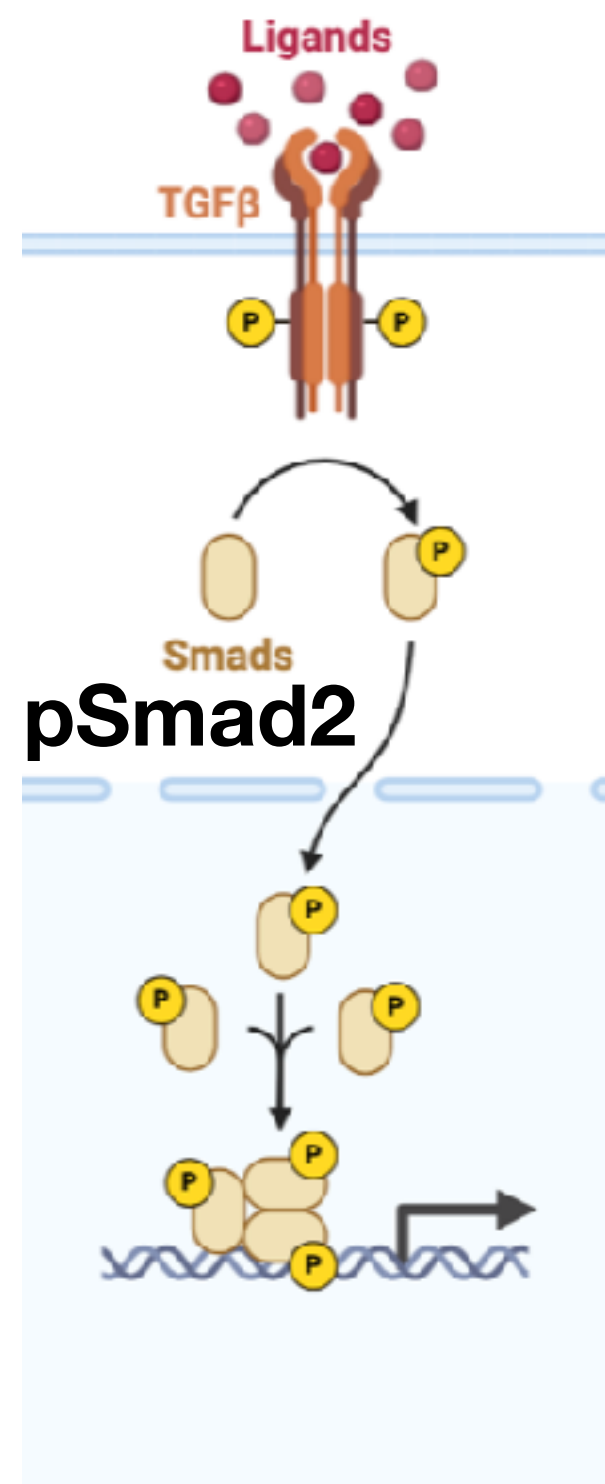


Threshold-based gradient interpretation (DV pattern)

**Required for A-P patterning**

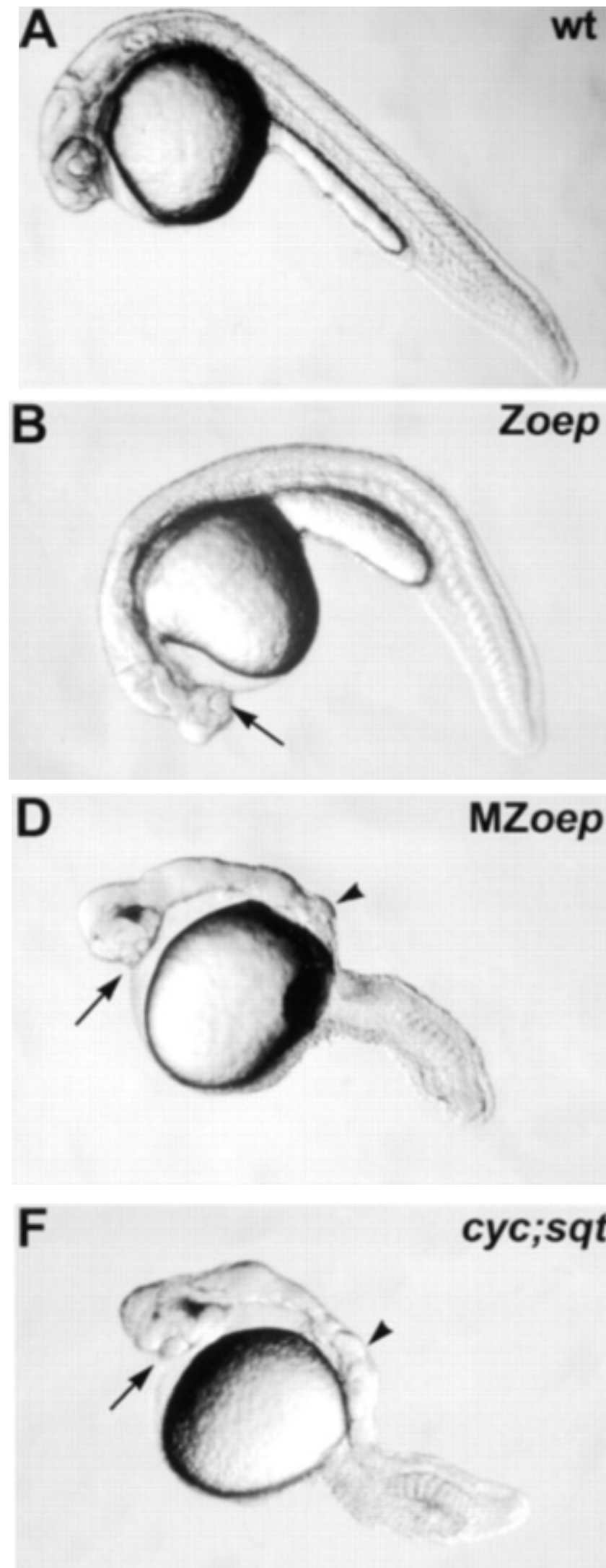
# Organizer signaling in the zebrafish: Nodal gradients

## Nodal-related 1/2

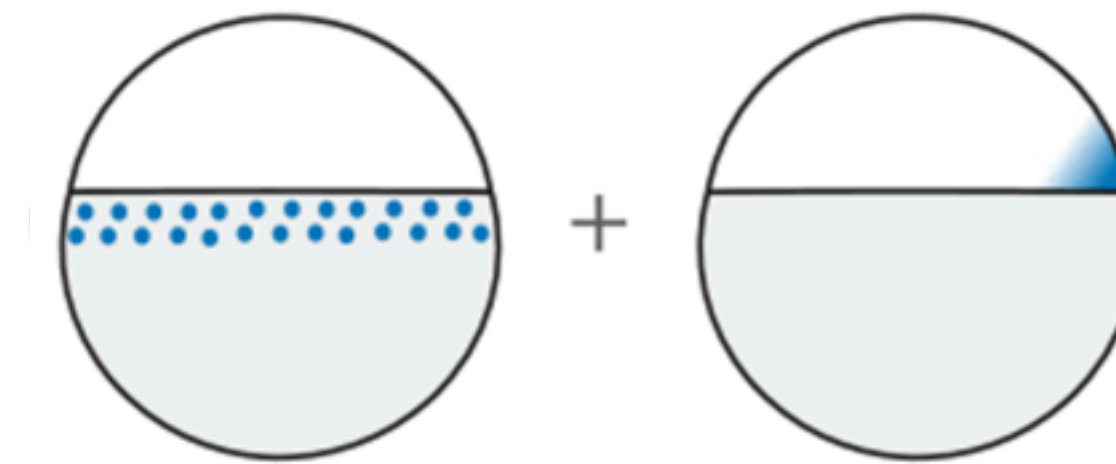


**TGFβ  
receptors**

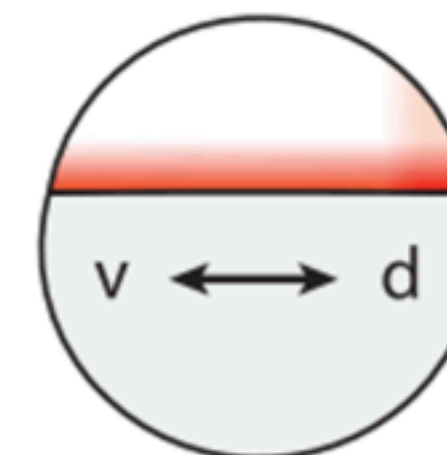
Activate Smad  
transcription factors in the  
cytosol by phosphorylation



YSL  
Ndr1/2 ligand      Dorsal  
Ndr1 ligand



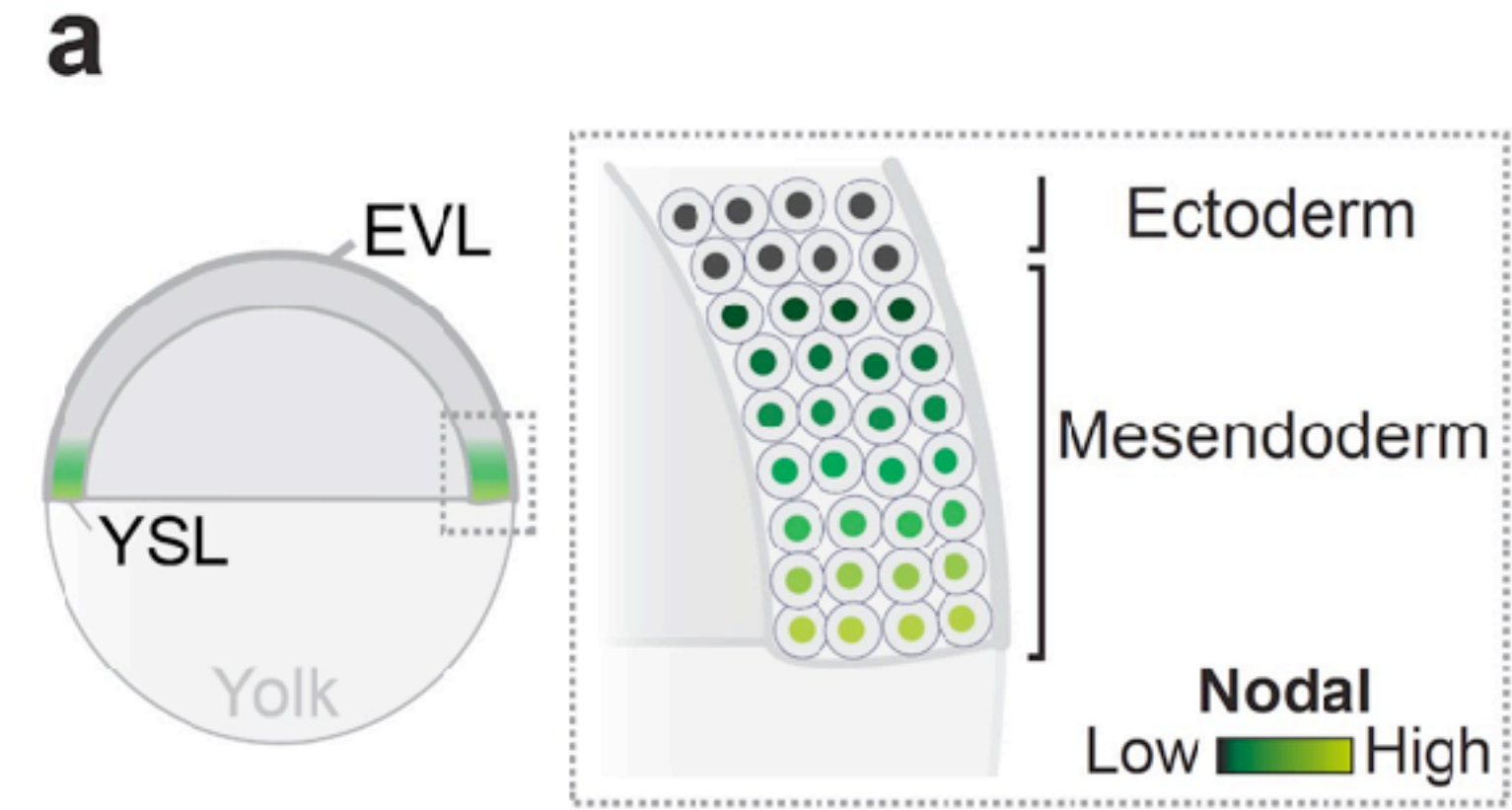
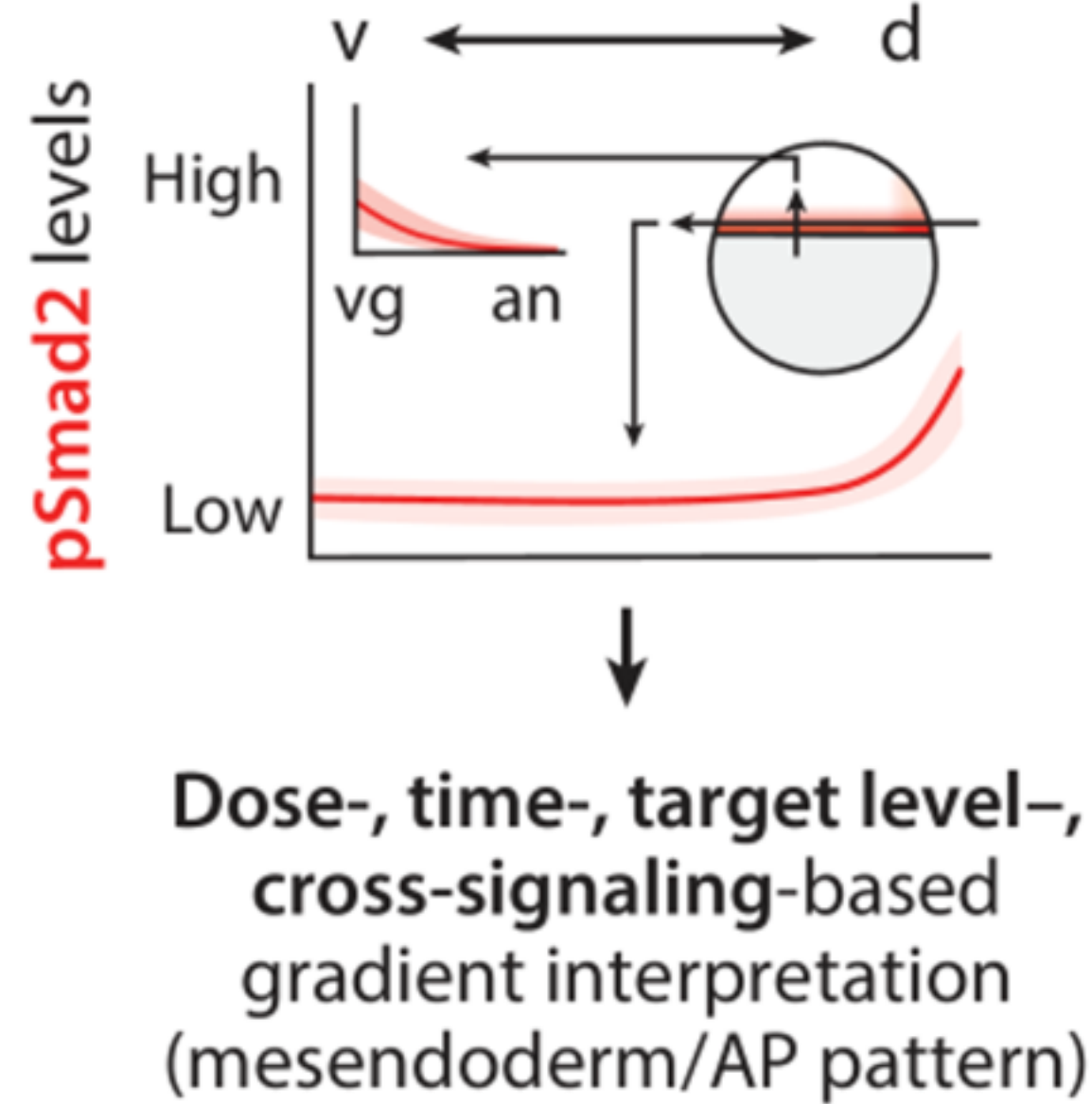
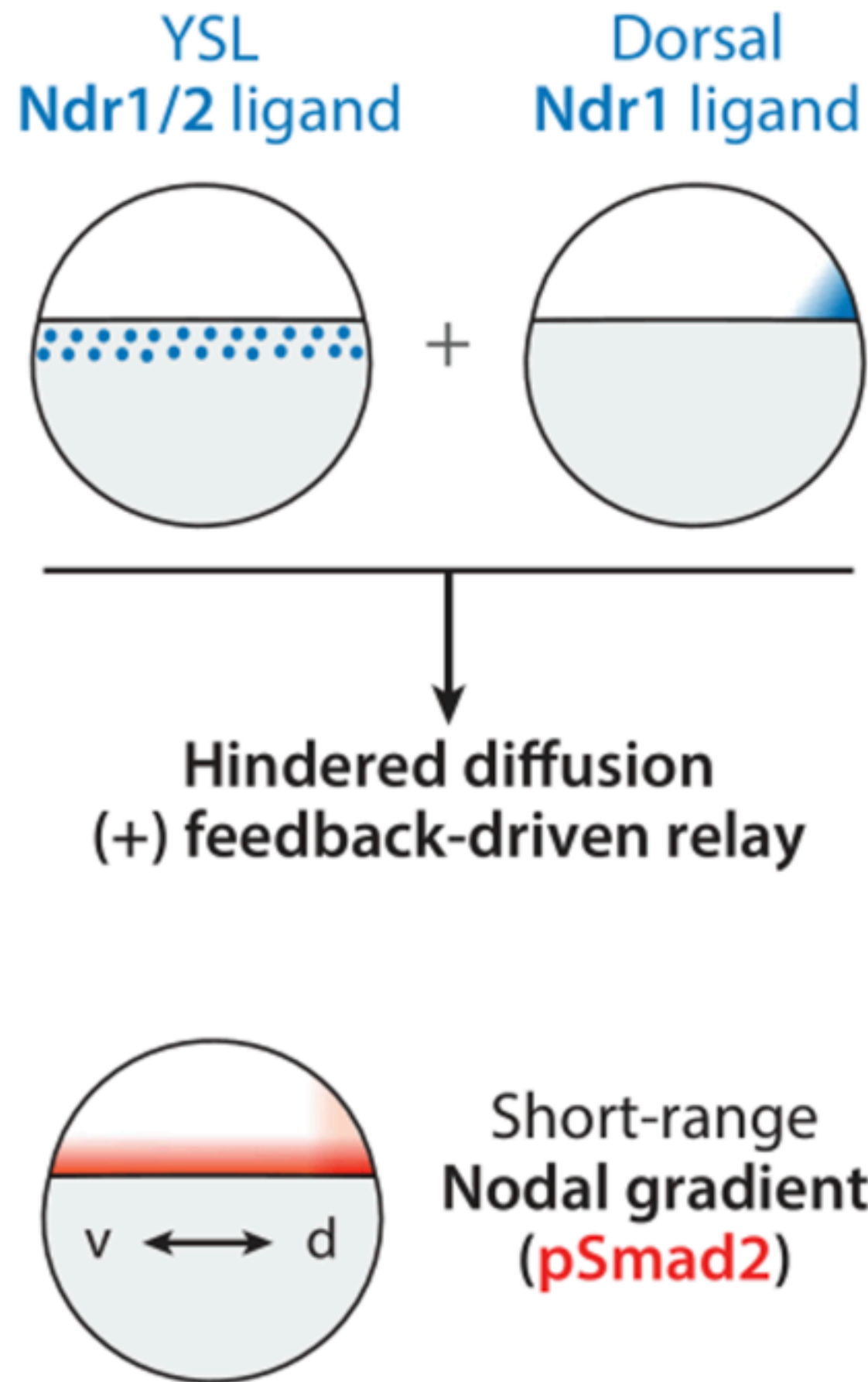
Hindered diffusion  
(+) feedback-driven relay



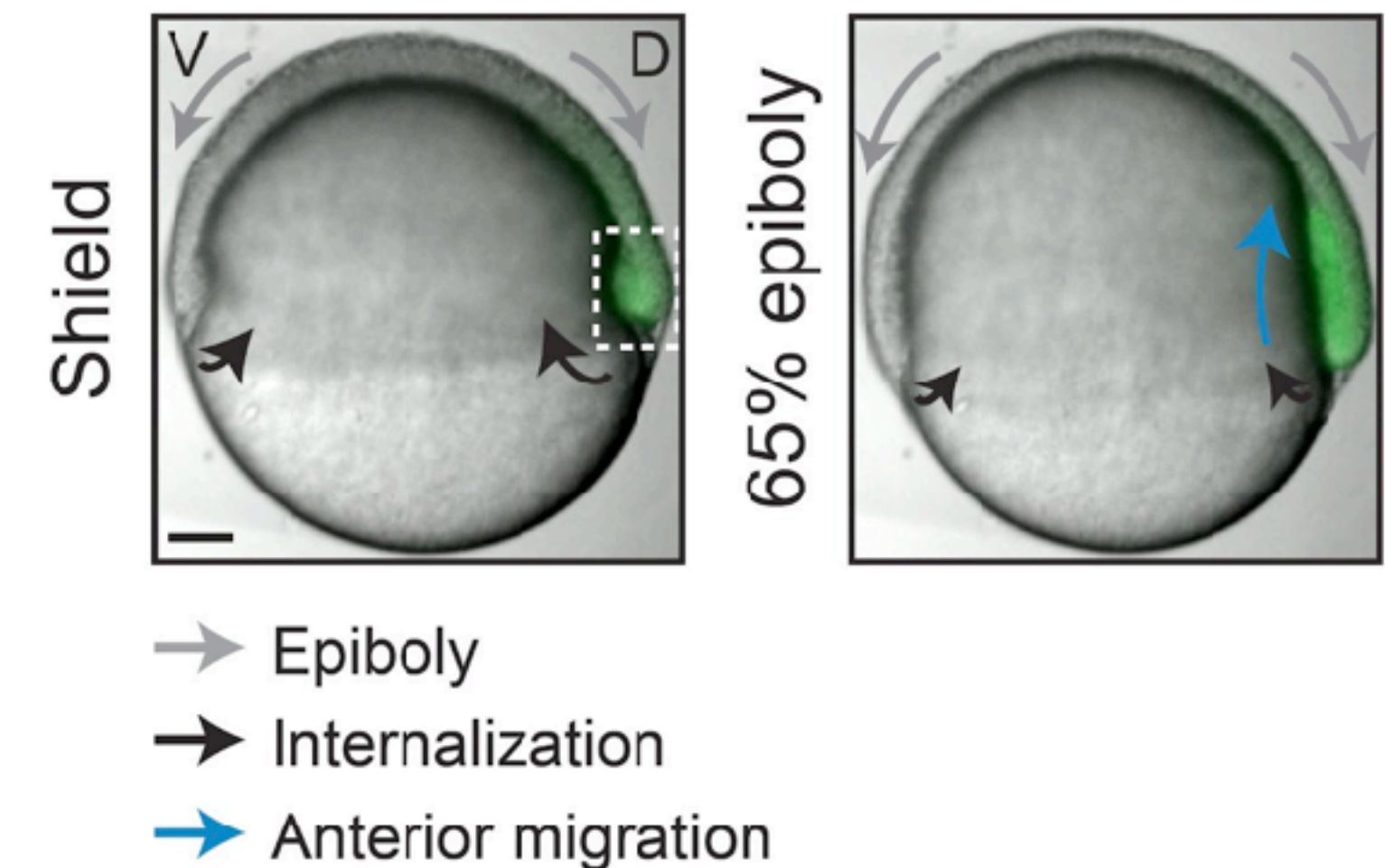
Short-range  
Nodal gradient  
(pSmad2)

**Nodals required for  
Mesoderm and Endoderm,  
& Chordin induction**

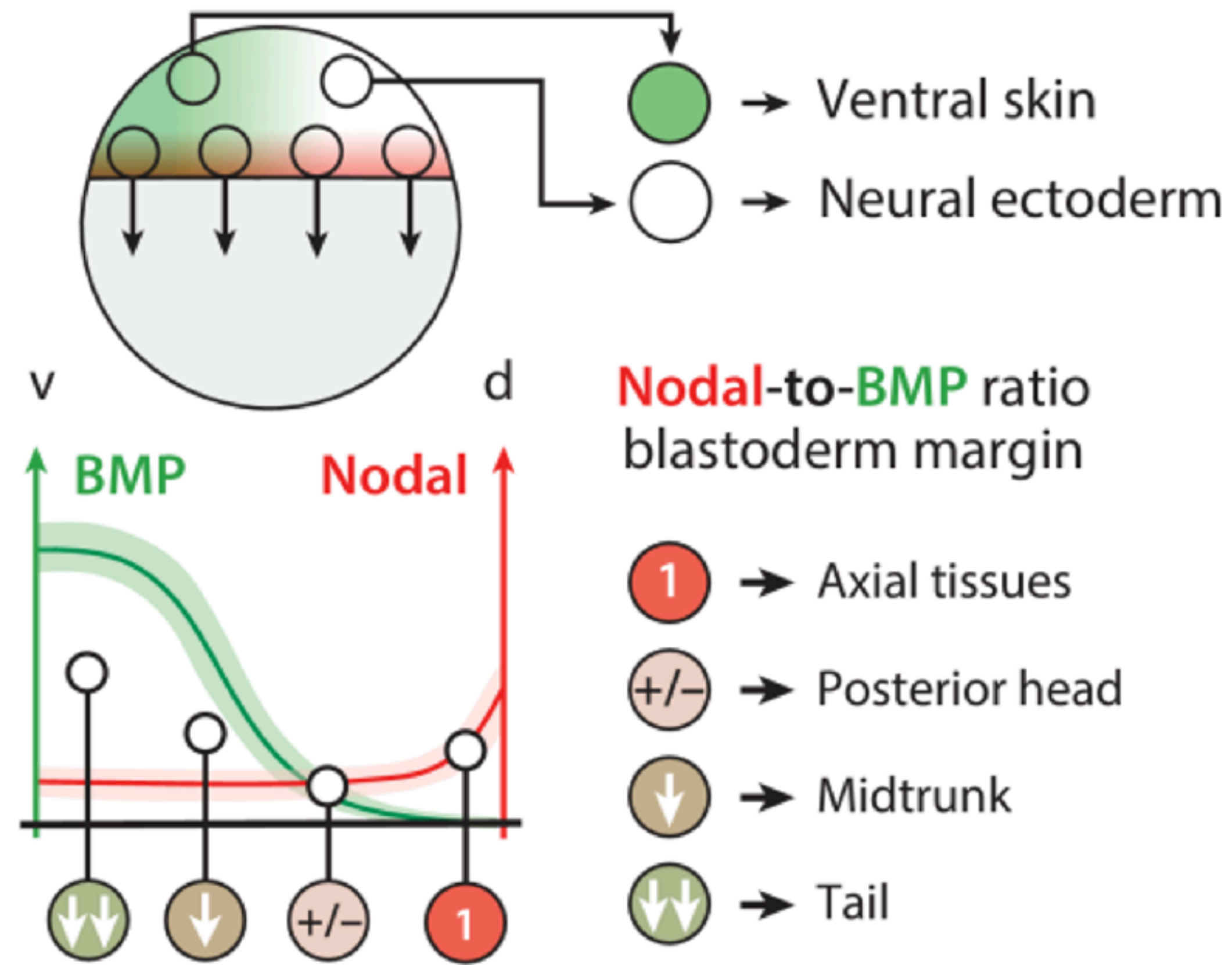
# Organizer signaling in the zebrafish: Nodal gradients



a' BF *Tg(gsc::EGFP-CAAX)*

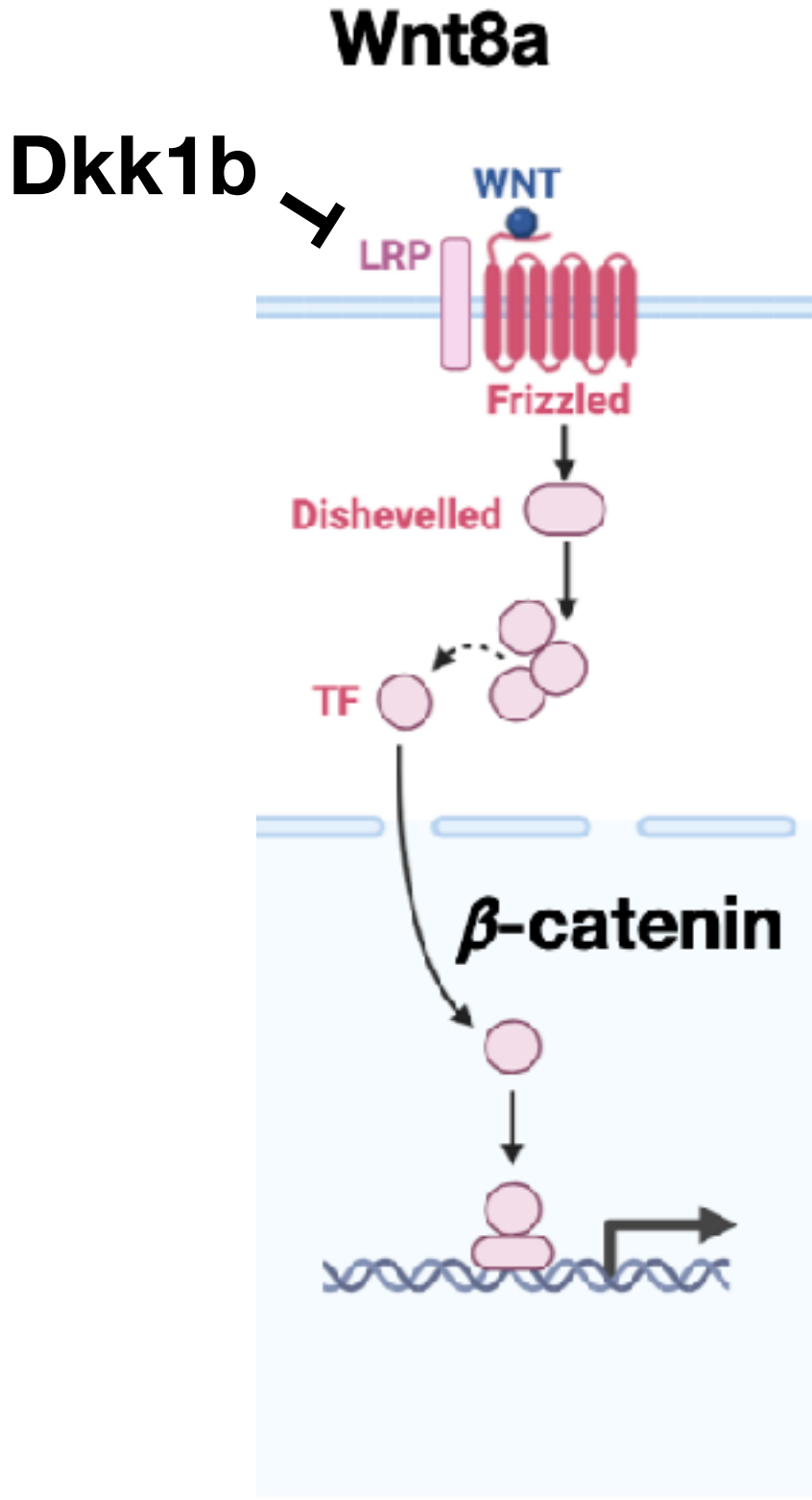


# Nodal / BMP ratio sets Anterior-Posterior position



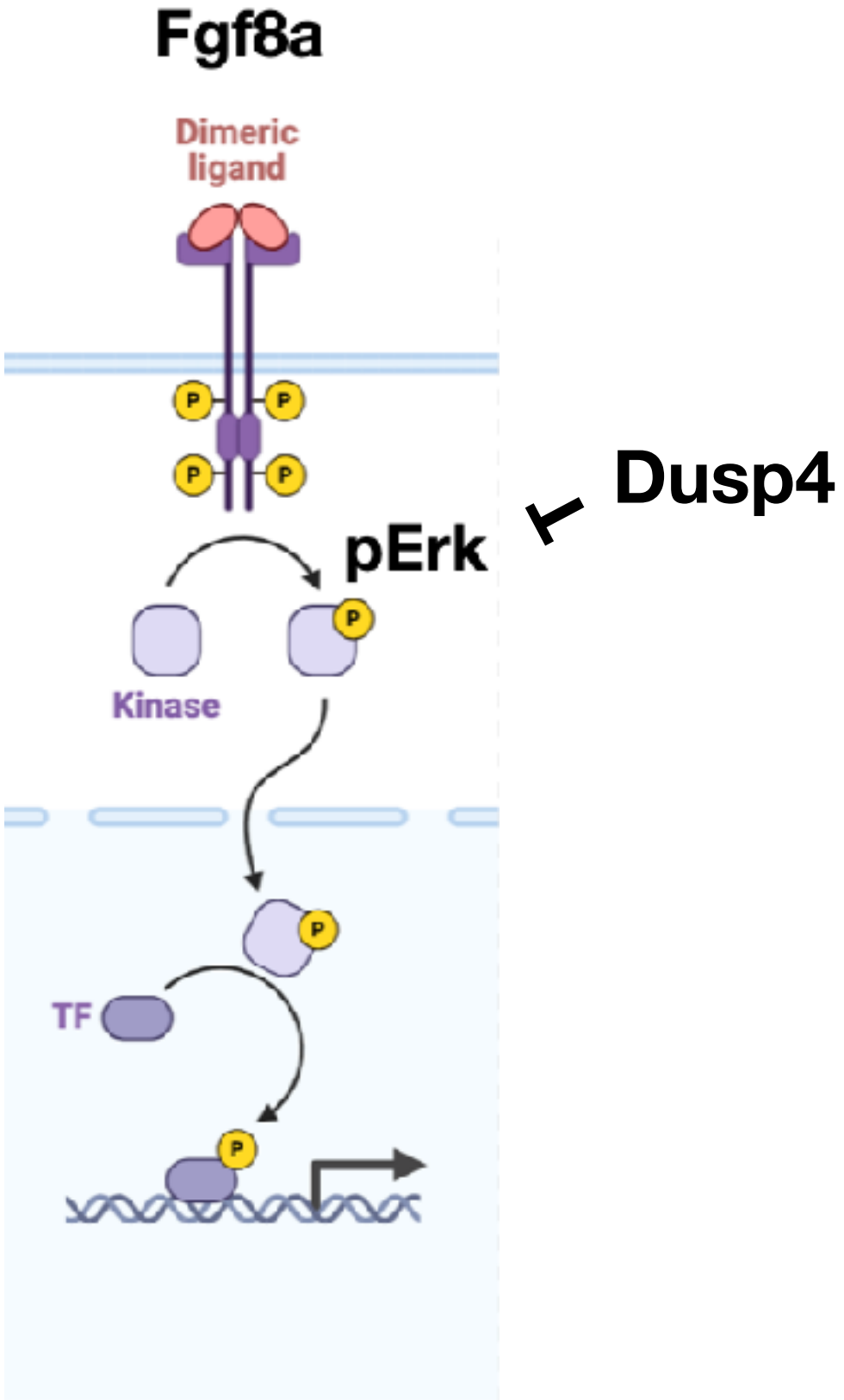
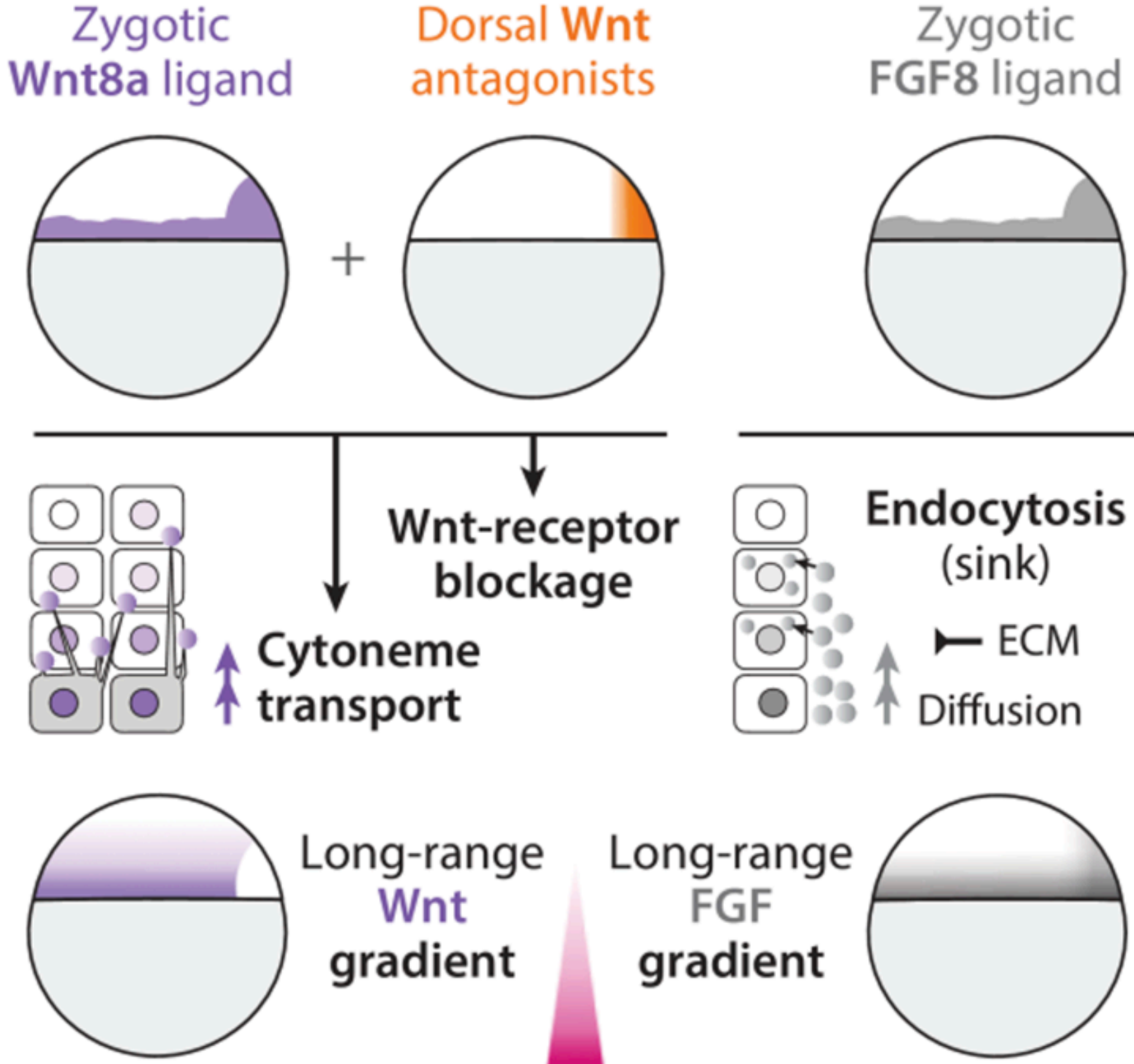
We didn't cover this in class

# Wnt8a and FGF8 pattern posterior body fates



**Wnt receptors**

Release an activated transcription factor from a multiprotein complex in the cytosol

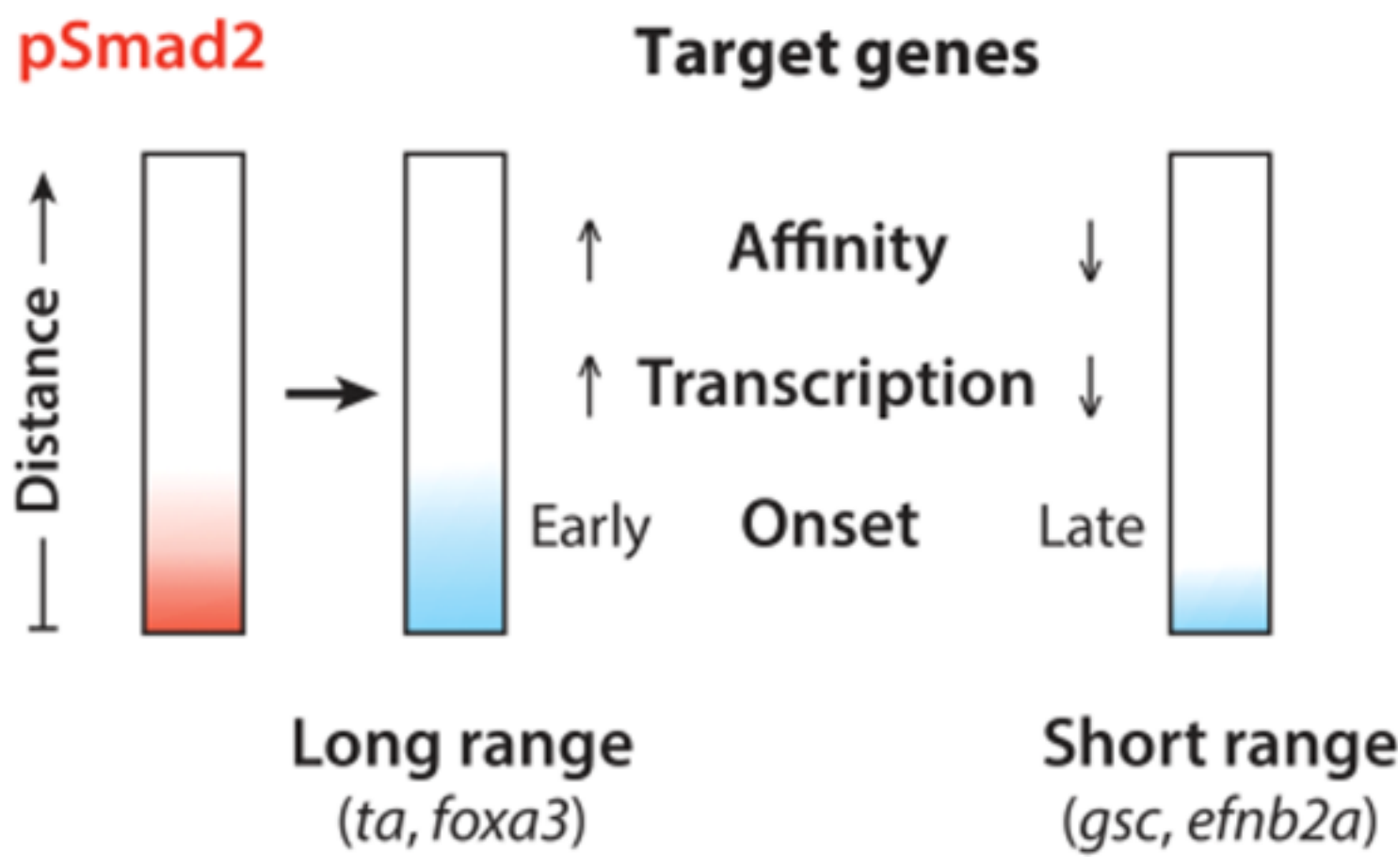
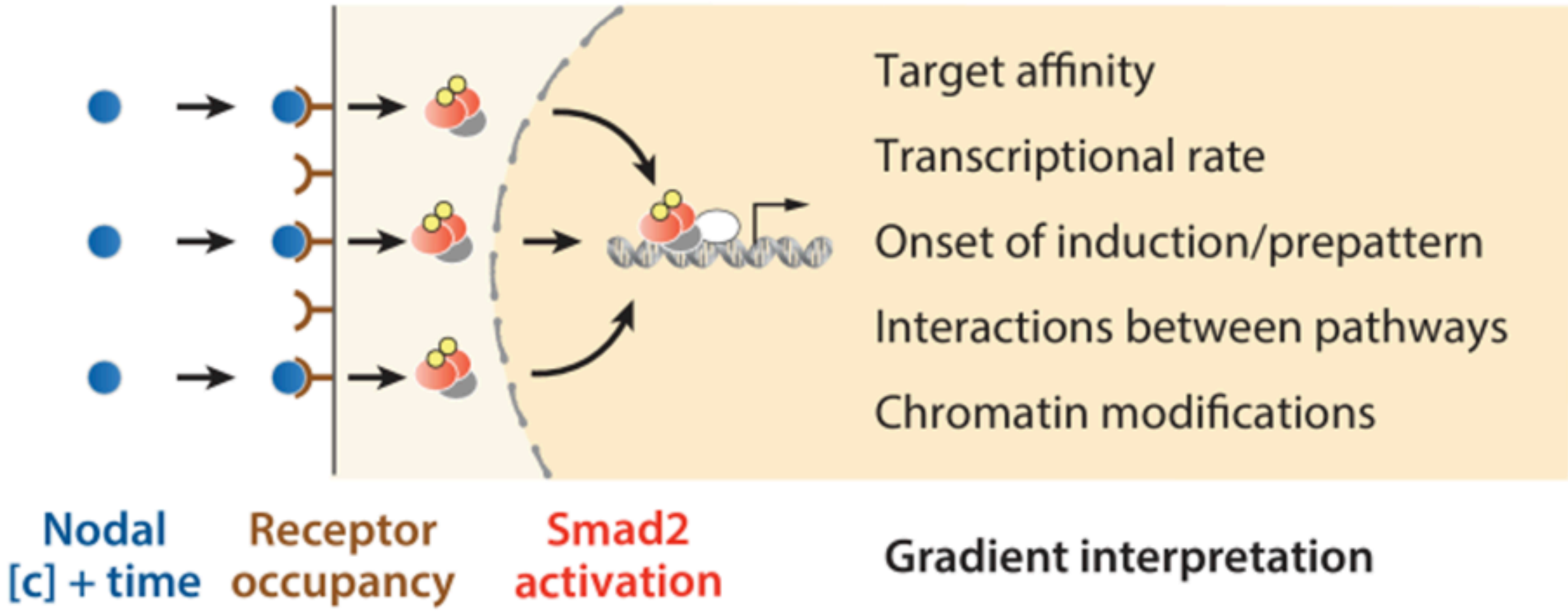


**Receptor tyrosine kinases**

Activate cytosolic kinases that translocate to the nucleus and activate transcription factors by phosphorylation

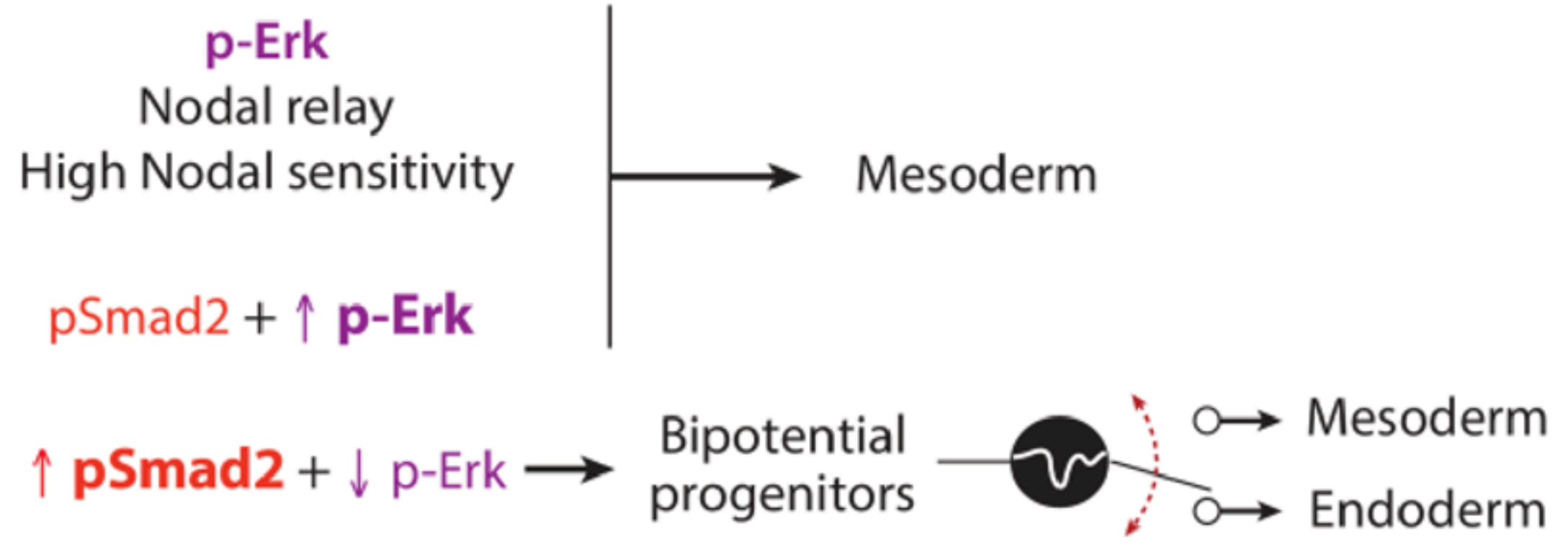
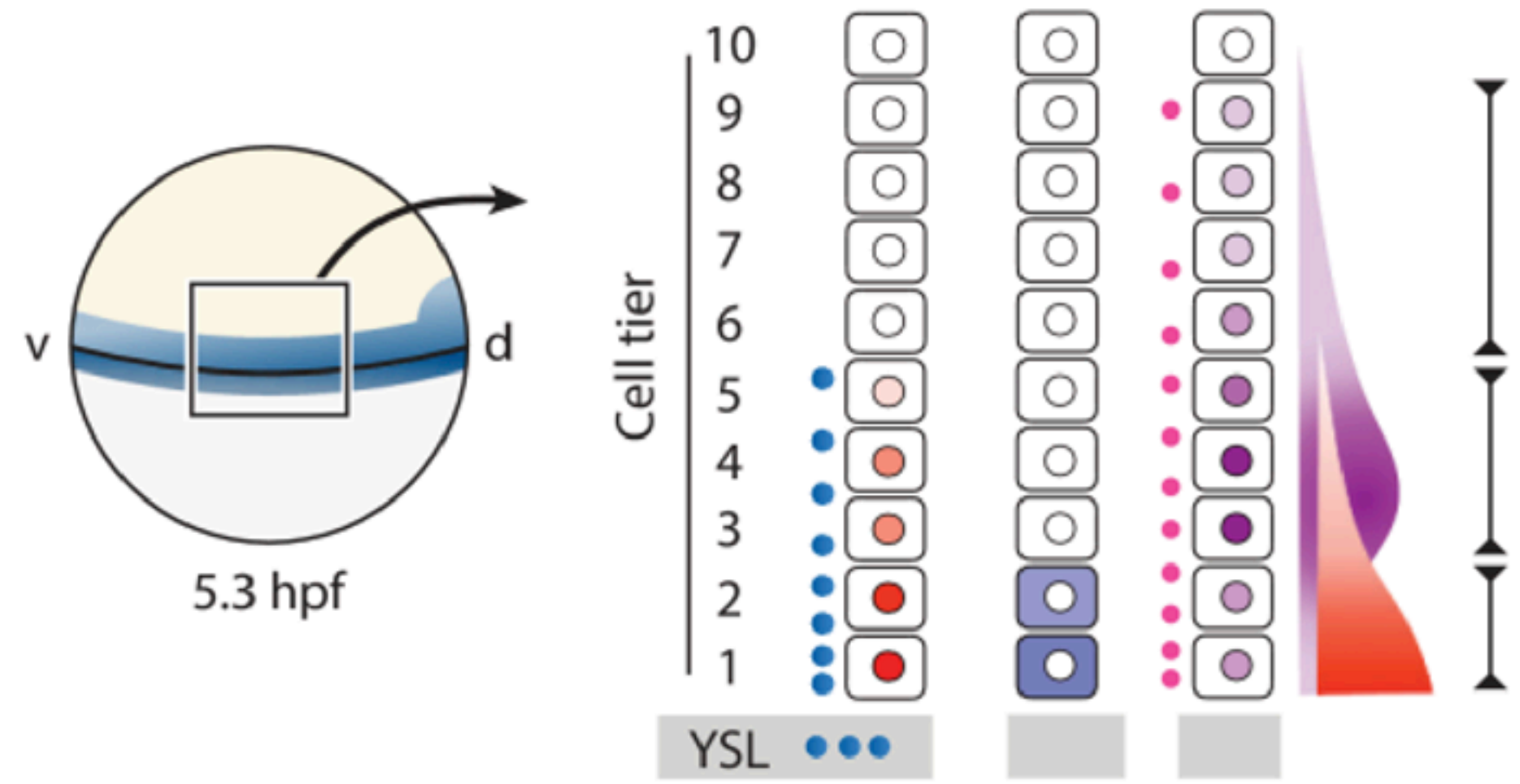
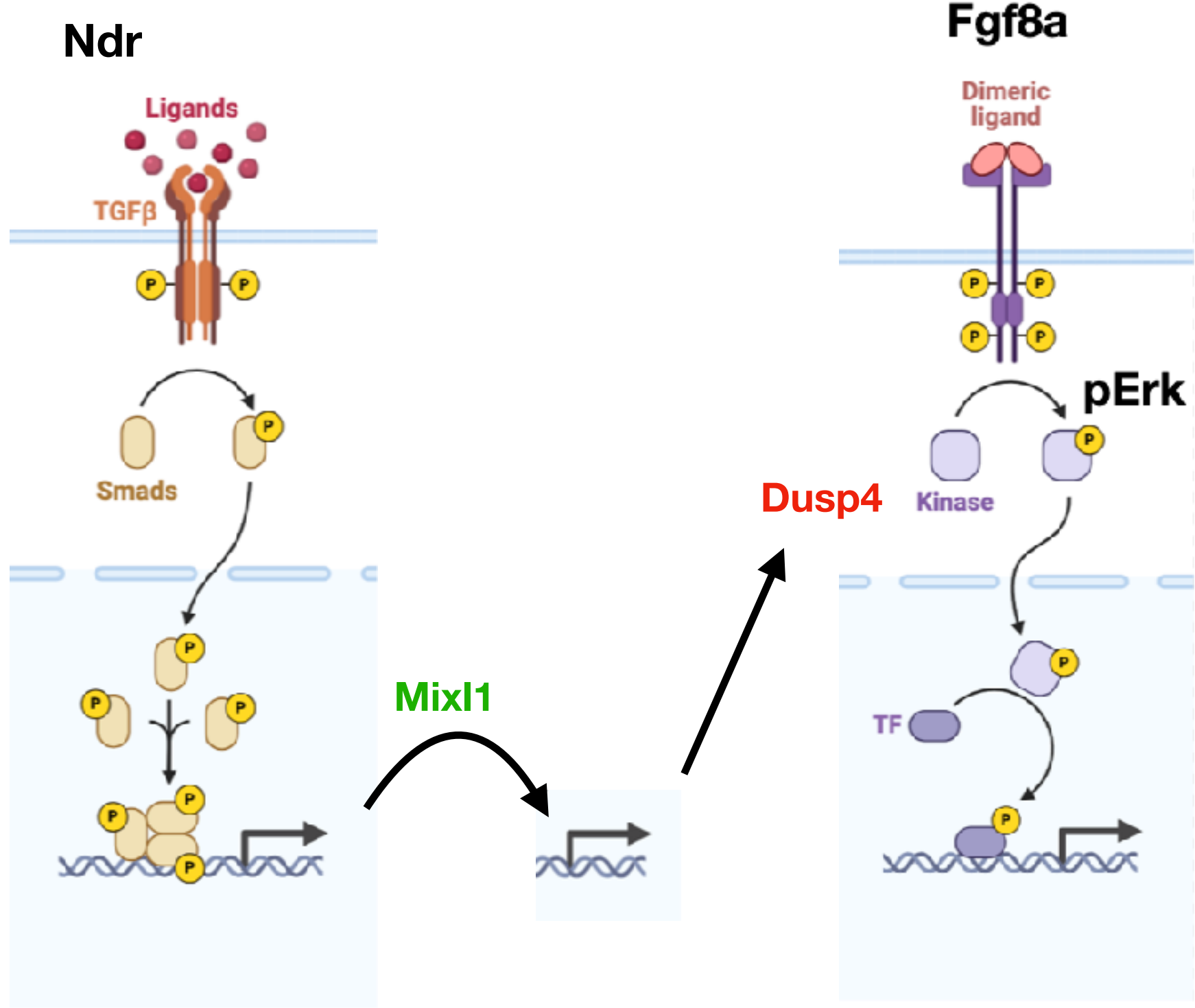
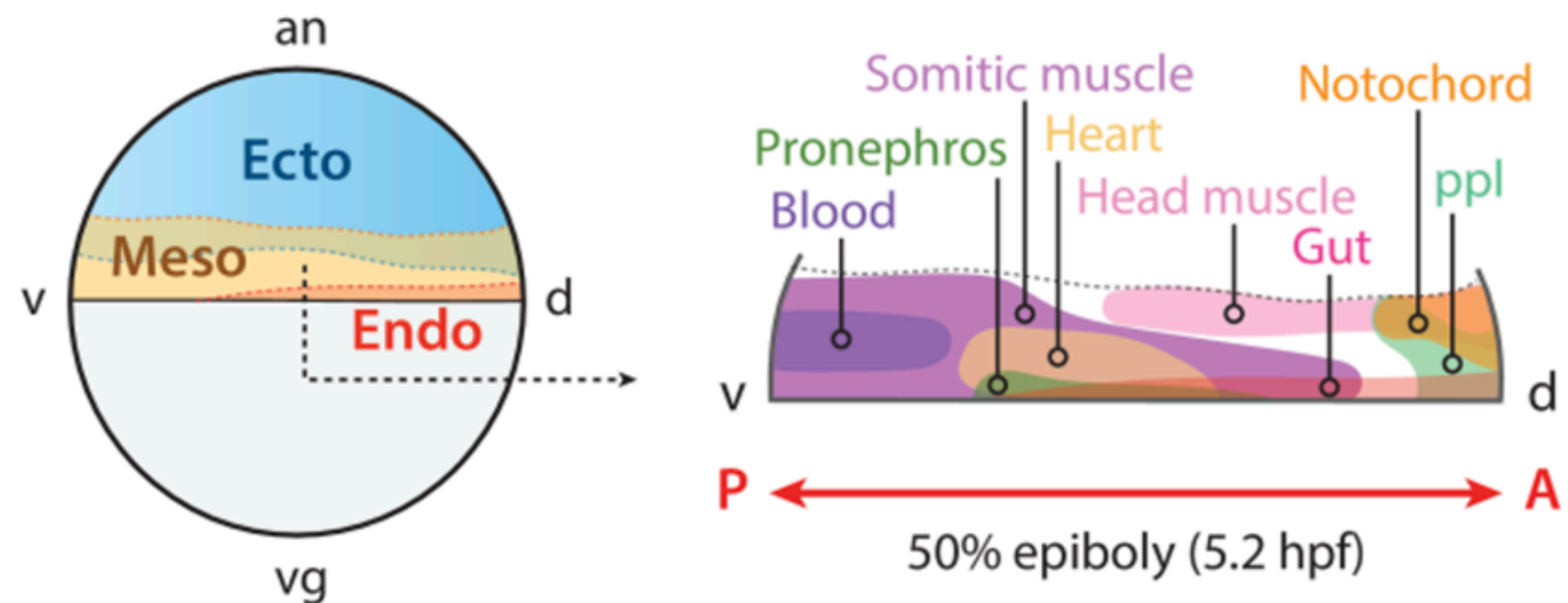
We didn't cover this in class

# Nodal gradient interpretation - short versus long range

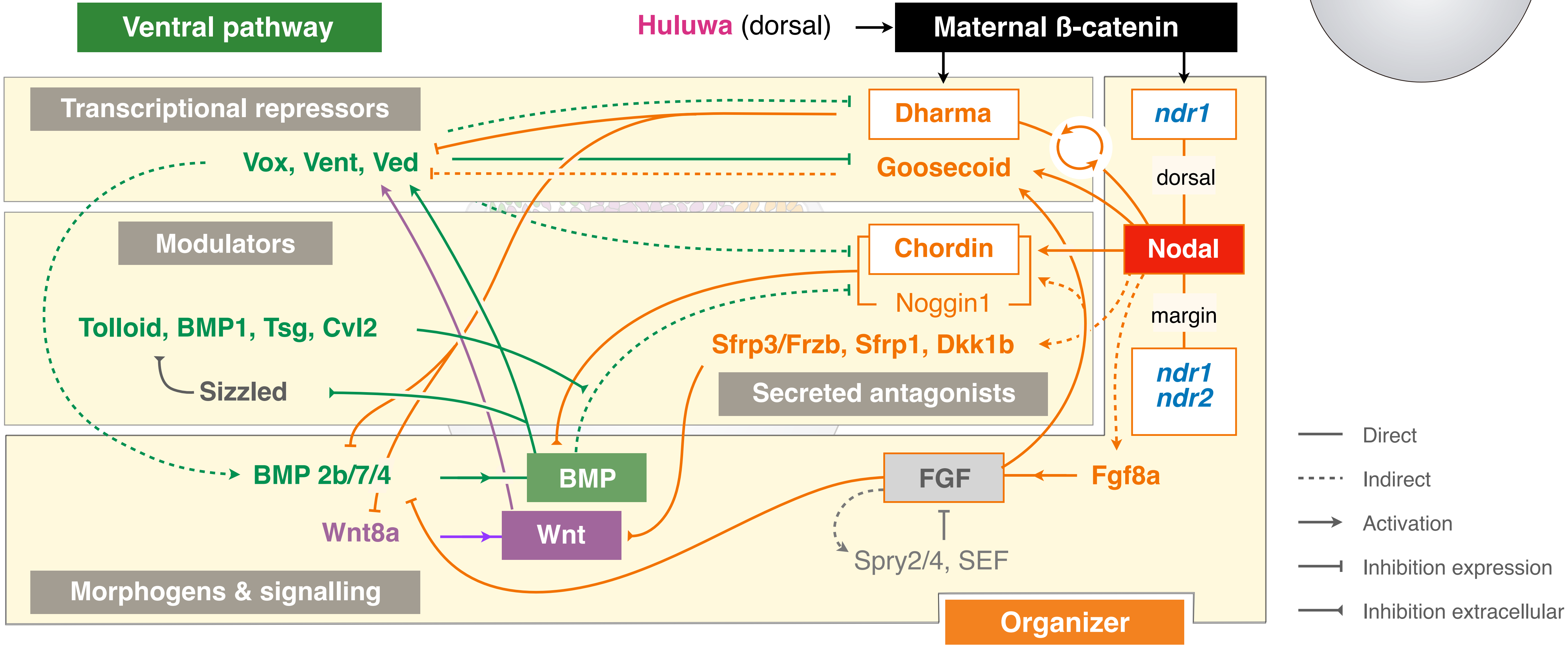
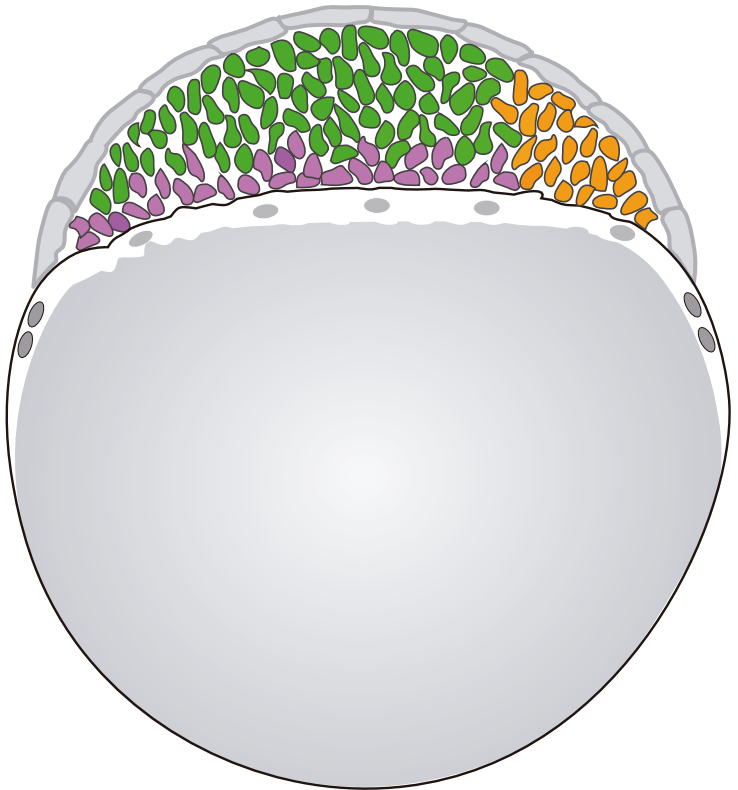


We didn't cover this in class

# Nodal and Fgf gradient interpretation - mesoderm versus endoderm



# Organizer signaling in the zebrafish

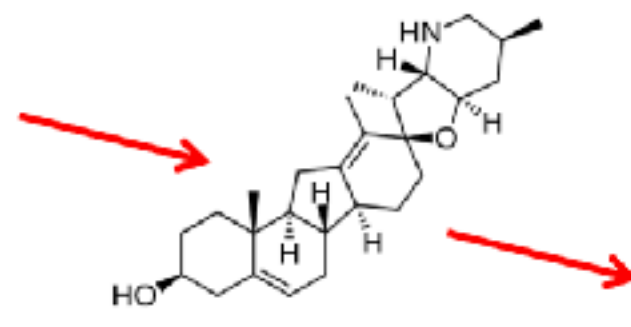


# Why does loss of Hh cause cyclopia?

## ● Blocking Hh signaling in mammals



corn lily



cyclopamine

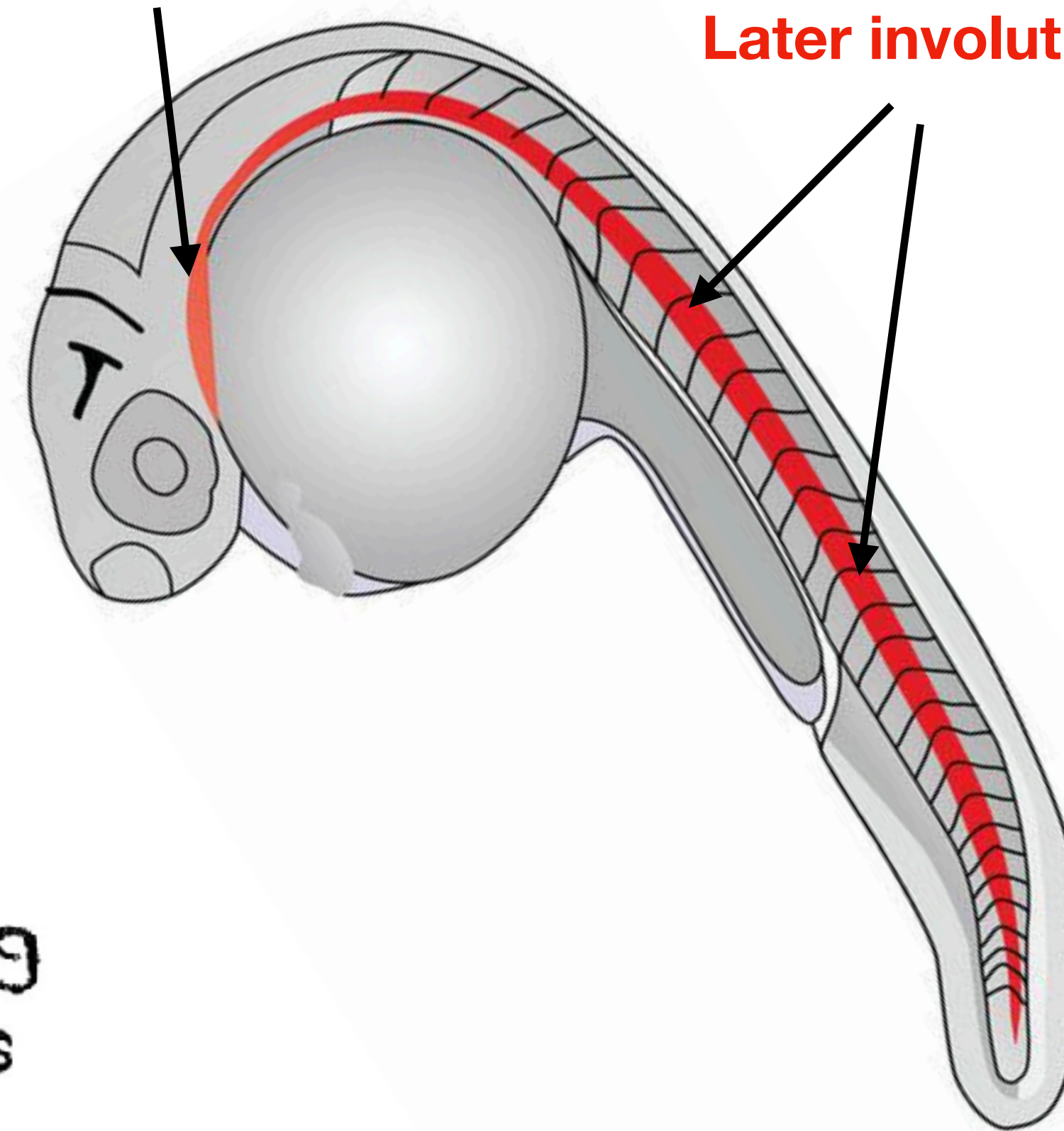


cyclopia (Idaho lamb)

Works also in other species

Early involuting cells

Later involuting cells



**Are these gradients sufficiently precise to account for tissue distributions?**

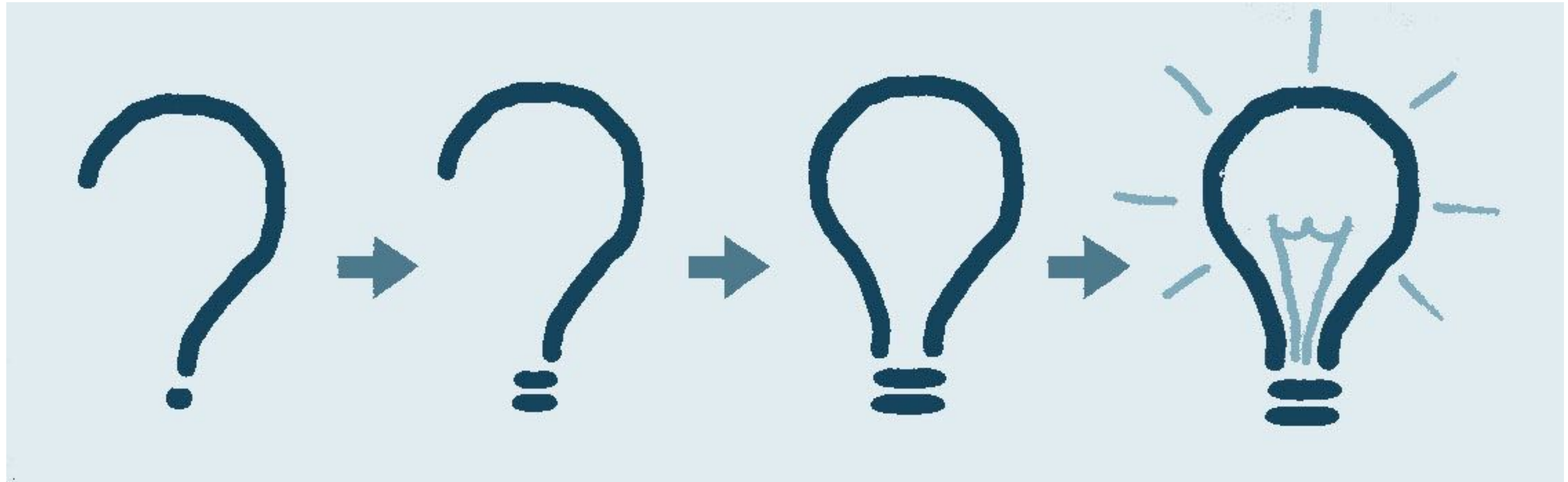
**What mechanisms are involved?**

**How do the gradients scale to regulate smaller or bigger embryos?**

**Is there a tail organizer?**



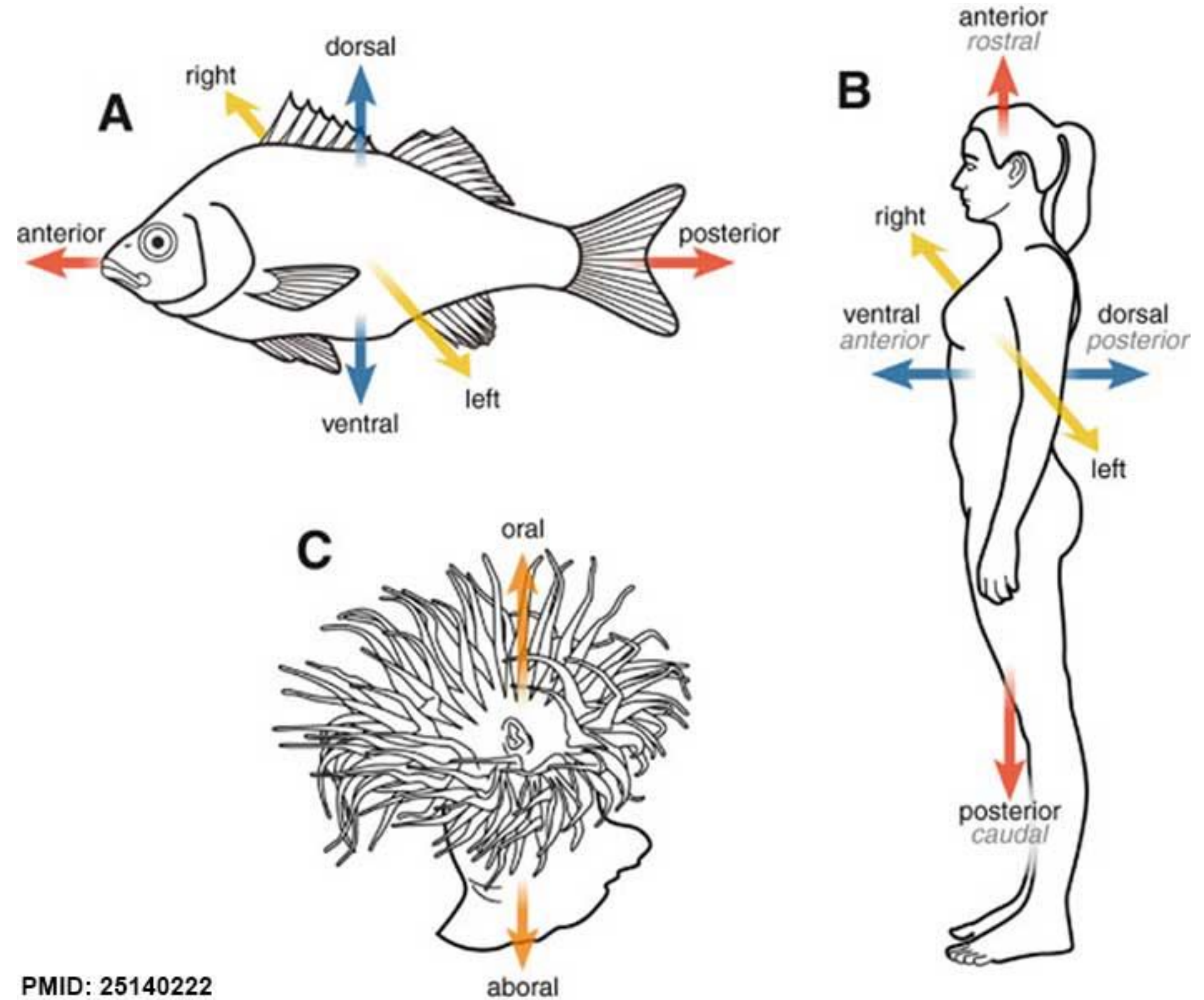
# Questions?



**Take a break**



# Big questions

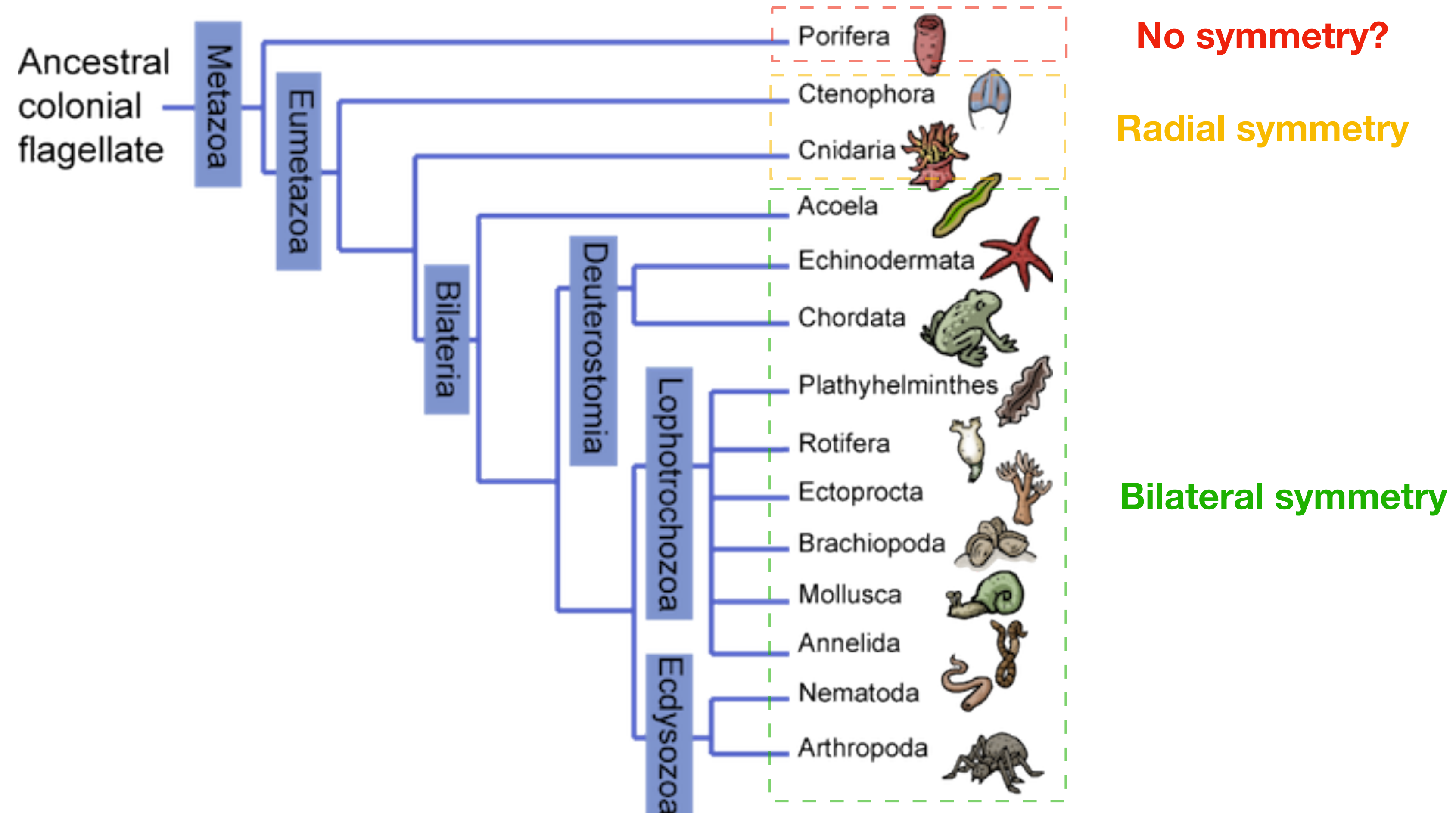


PMID: 25140222

# Today's menu - 2 courses

- *Front and Back*
- *Nobel experiment - embryonic induction*
- *Specification and commitment*
- *Gastrulation and patterning of germ layers*
- *Morphogen gradients (again)*
- *Left and Right*
- Situs inversus
- Cilia in the node - directed flow
- Planar polarity
- Nodal and calcium signals to the lateral plate

# Body plans across the animal kingdom



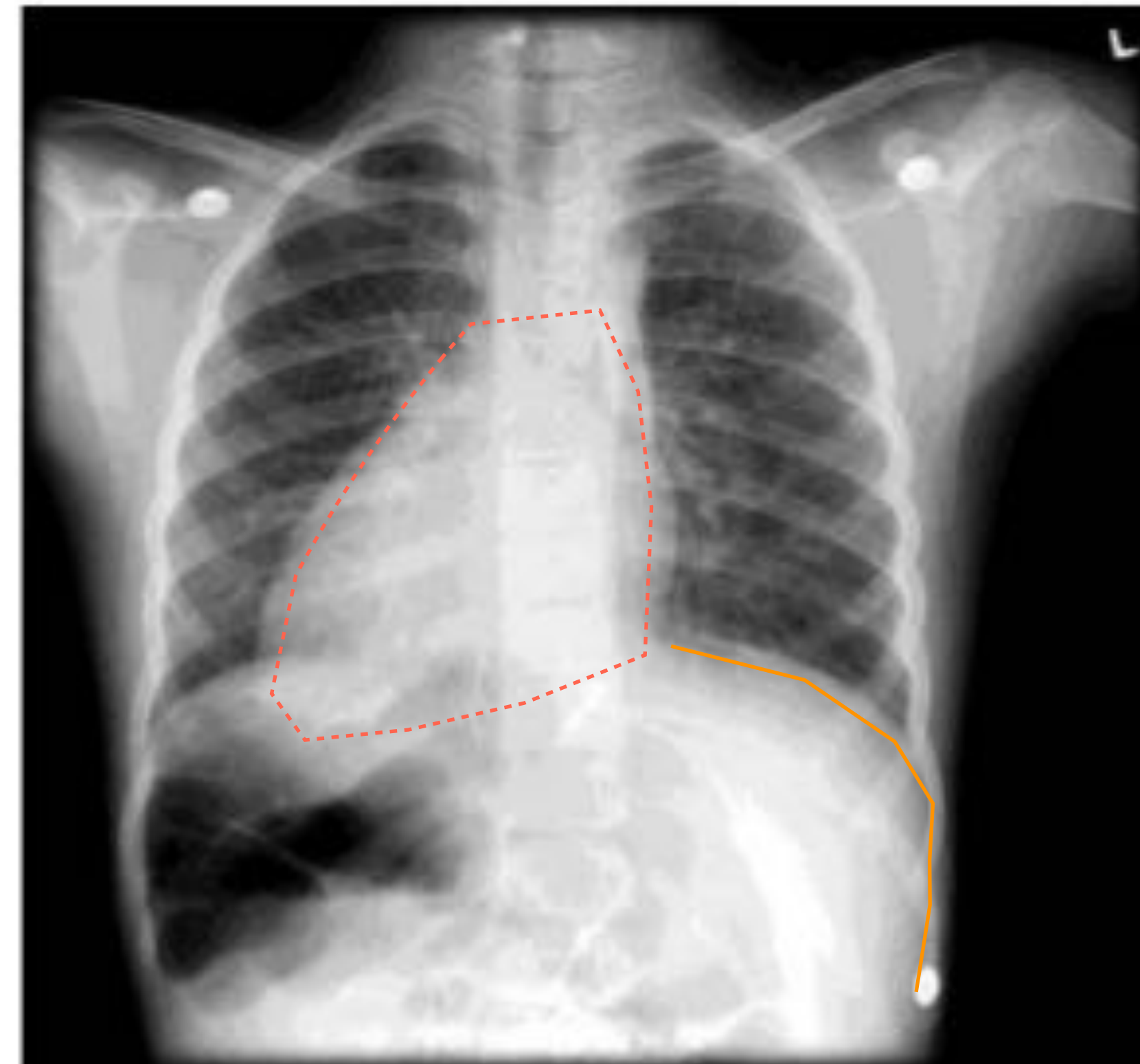
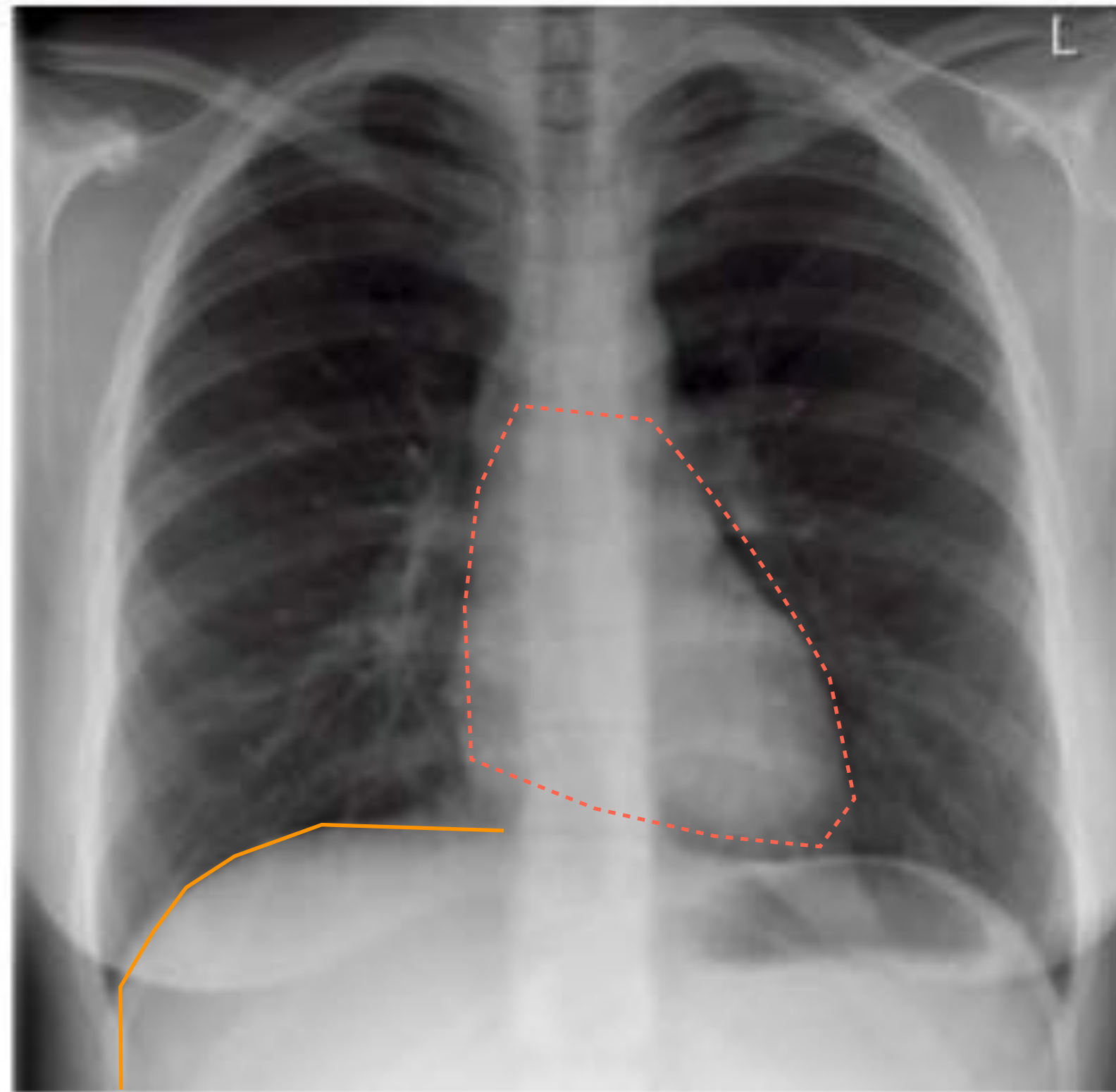
# Individual puzzle



# Medical Quiz: Spot the difference?

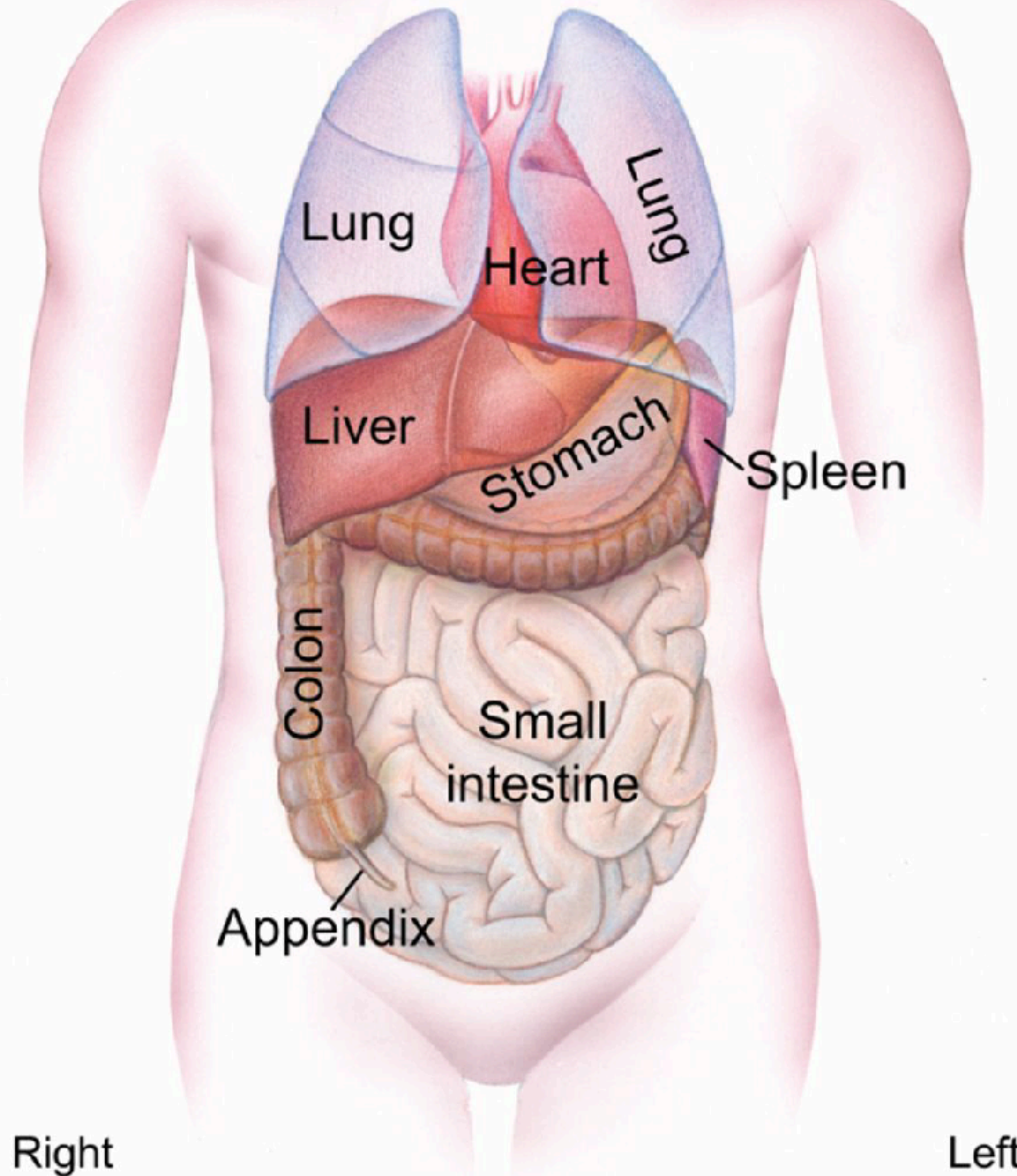


# Spot the difference?

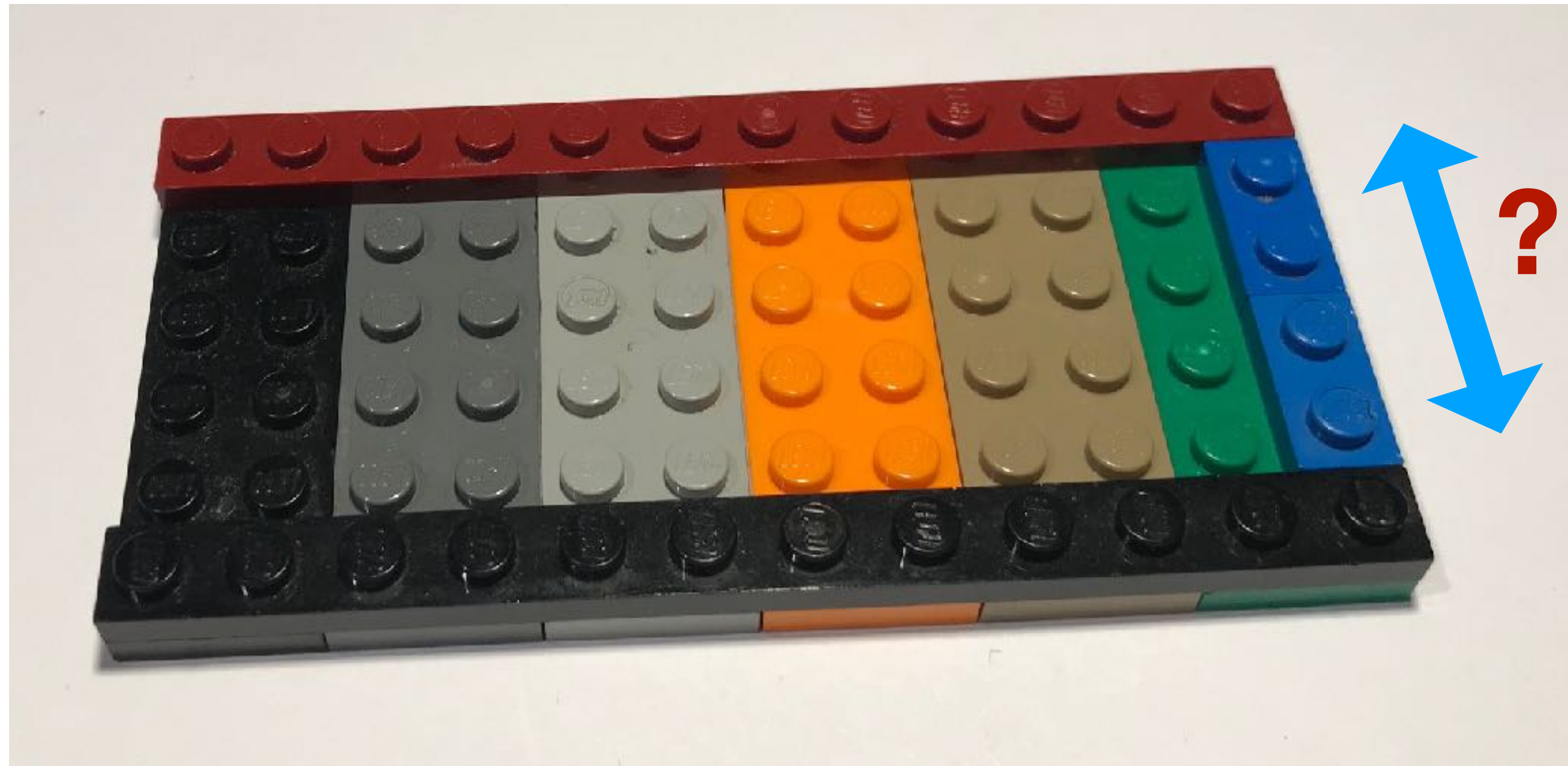


**Situs inversus**

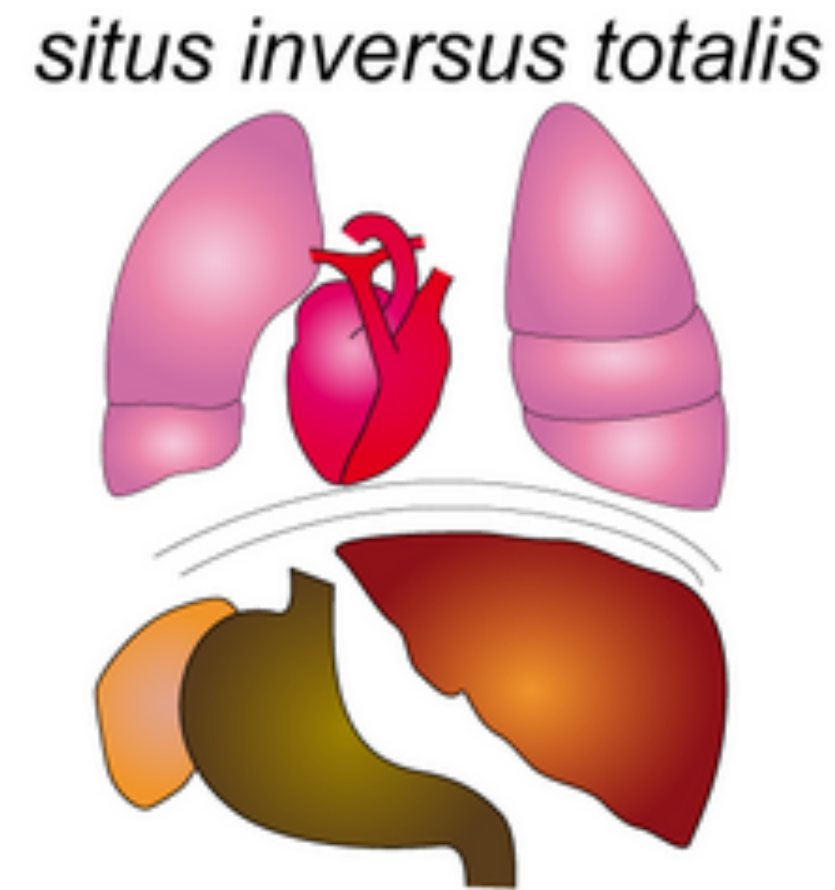
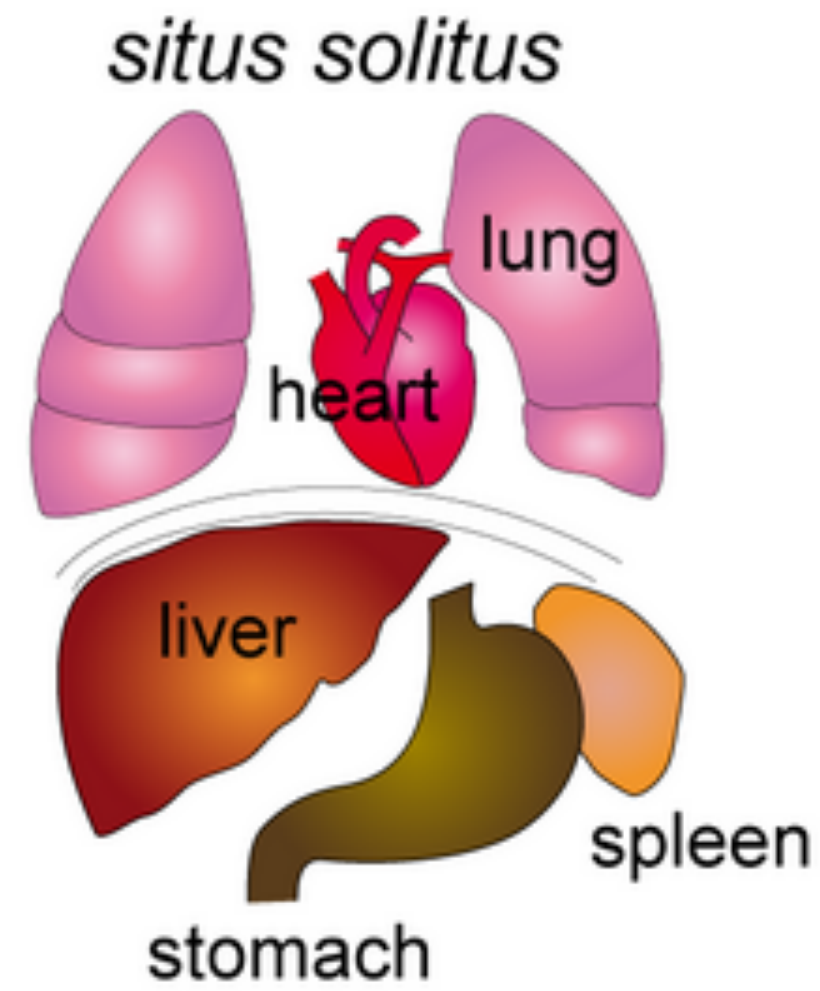
# Internal organs are asymmetric



# Left-Right axis

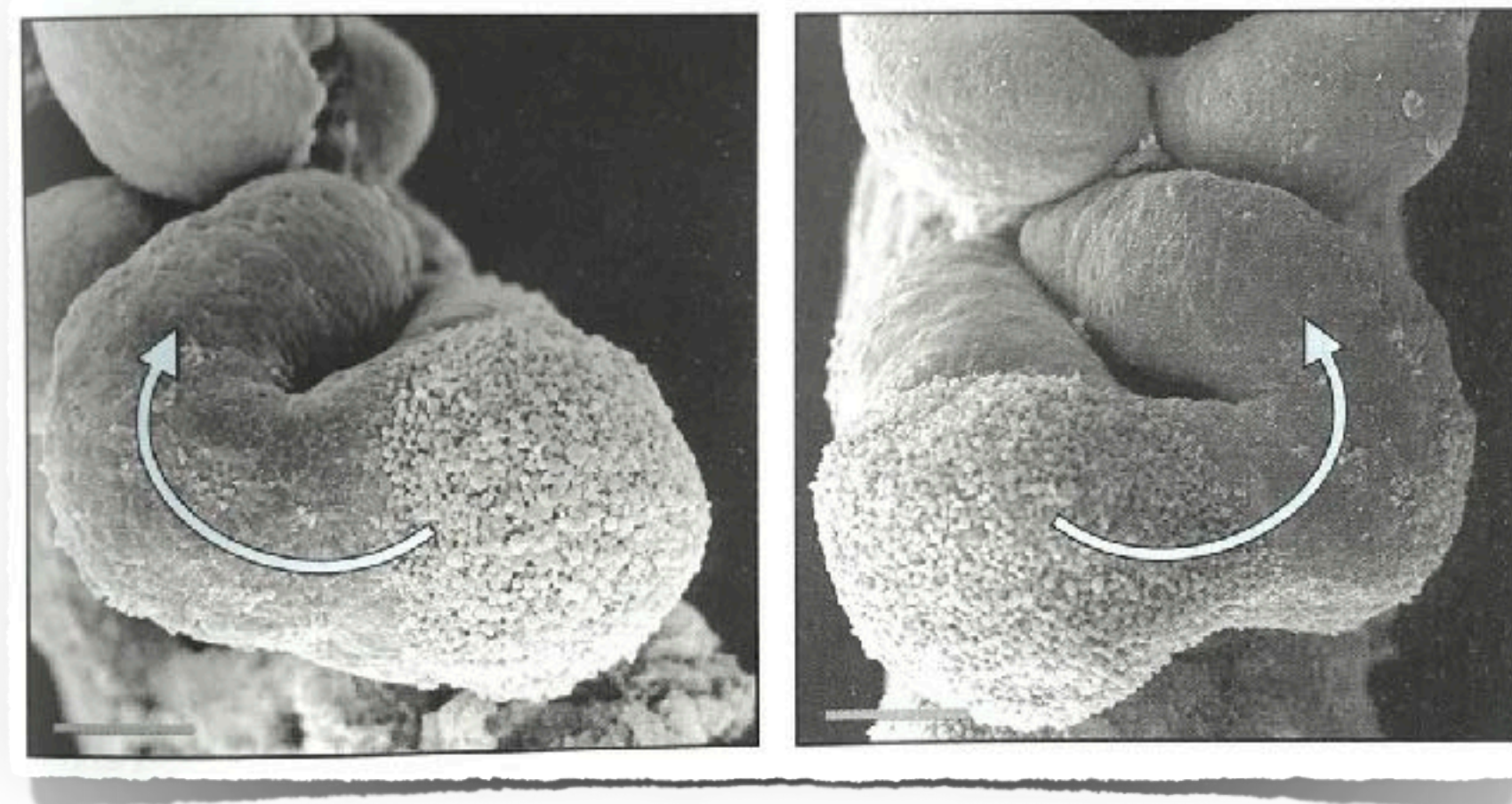
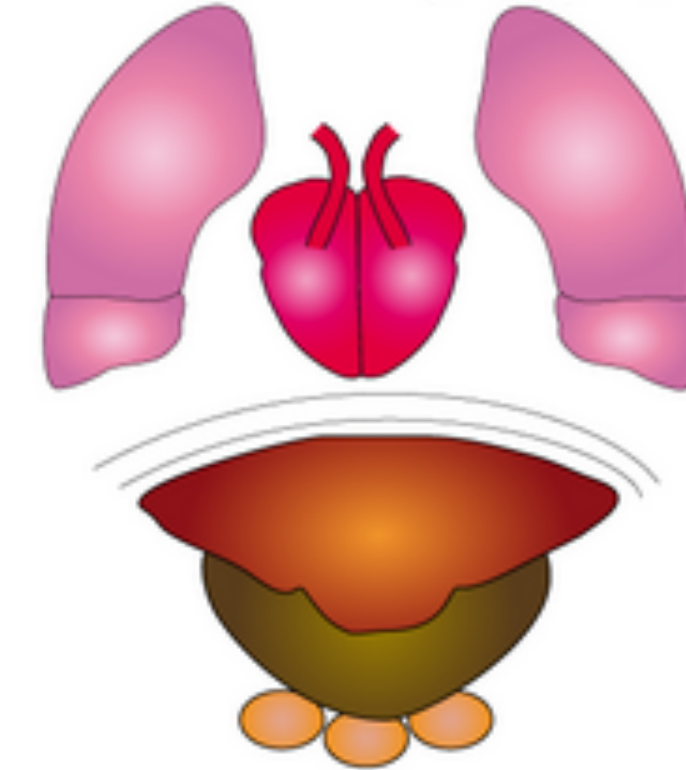


# Variant internal symmetries in mammals



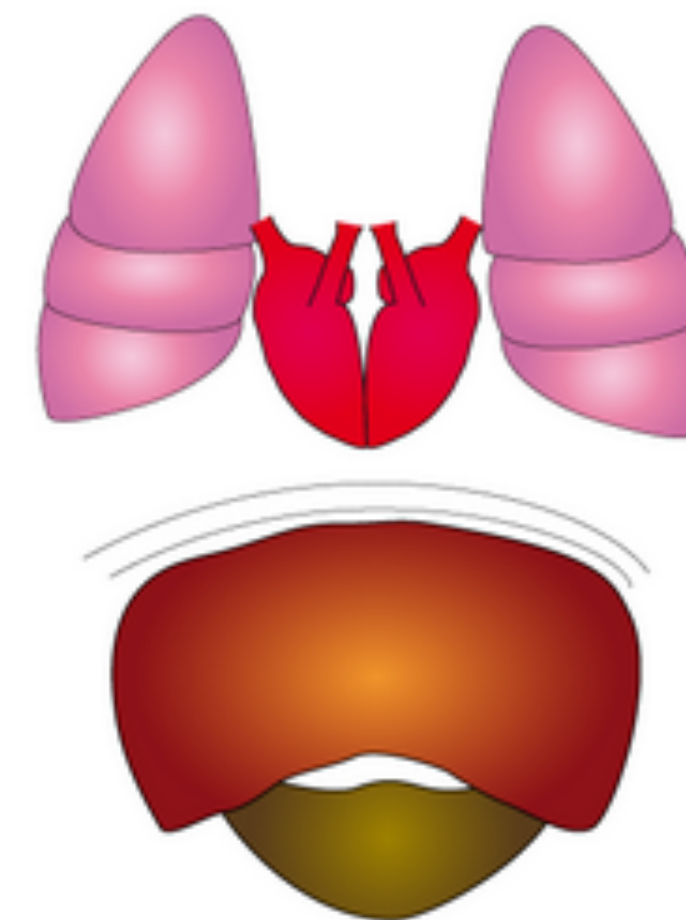
## Heterotaxy

*left isomerism (polysplenia)*

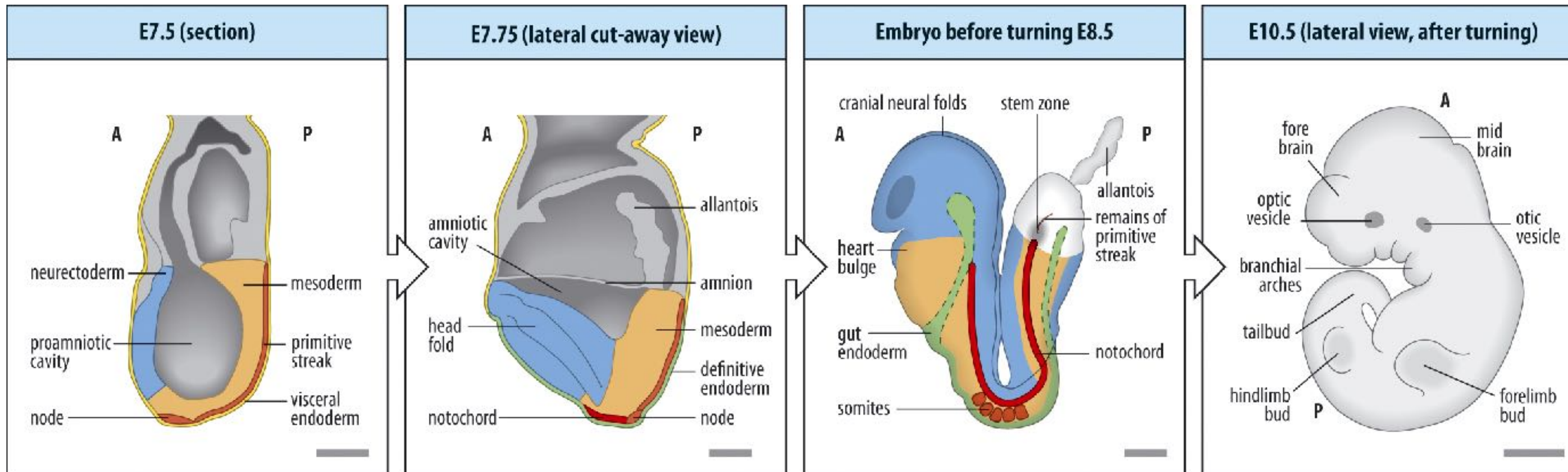
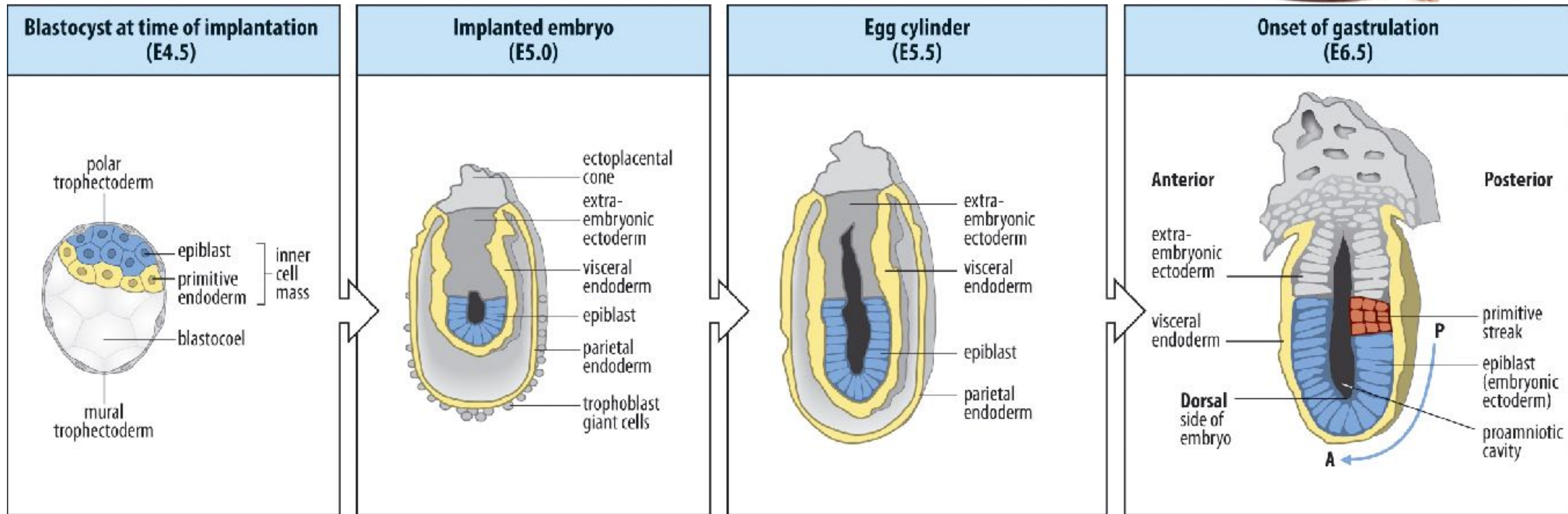


**Mouse *inversus viscerum (iv)* mutant:  
heart looping left-right 50%**

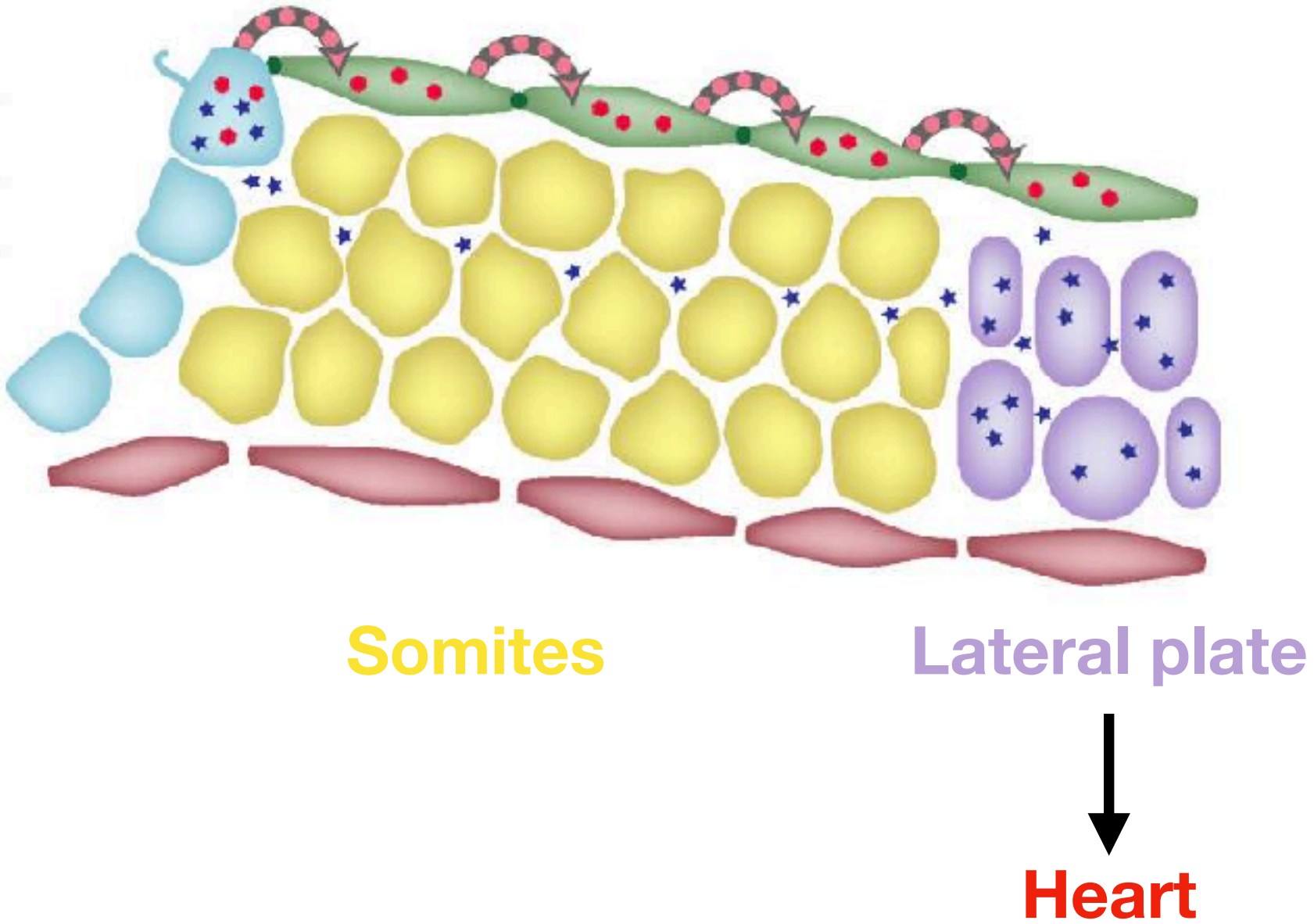
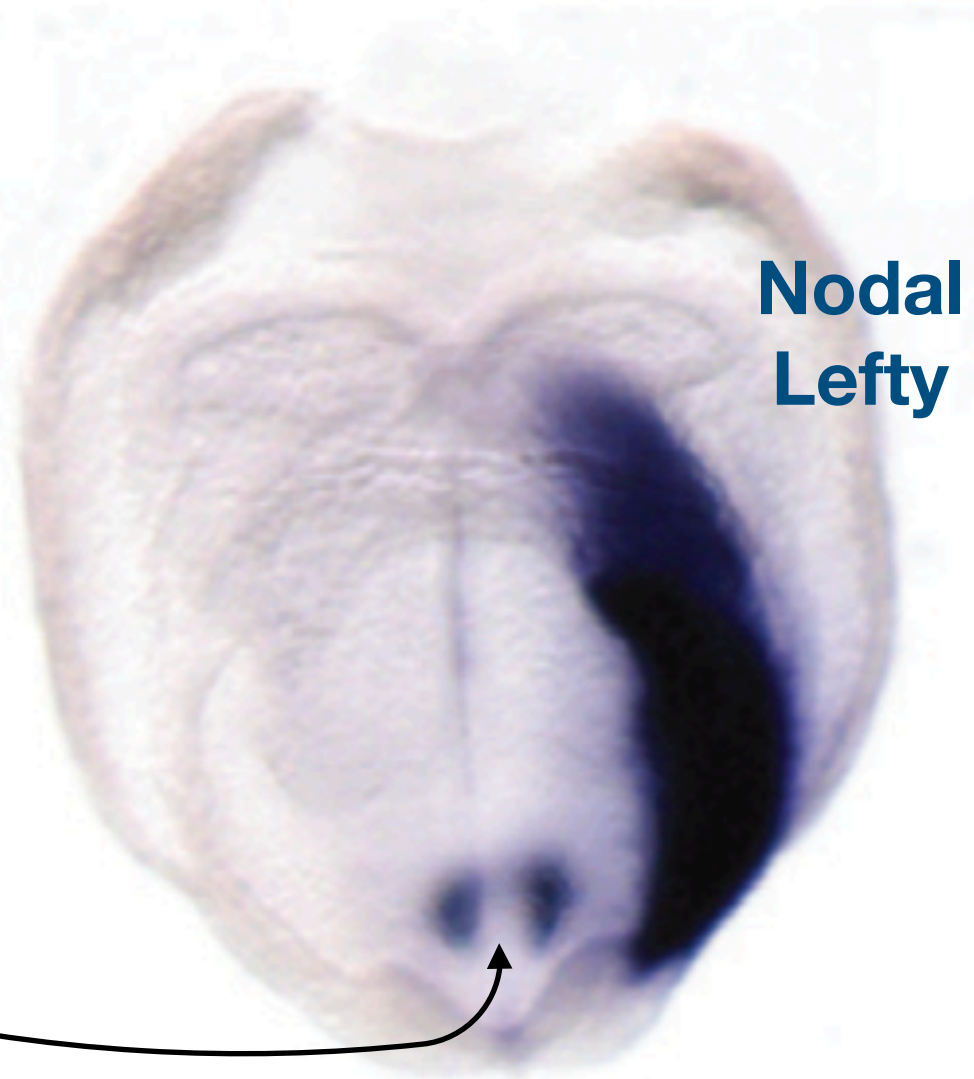
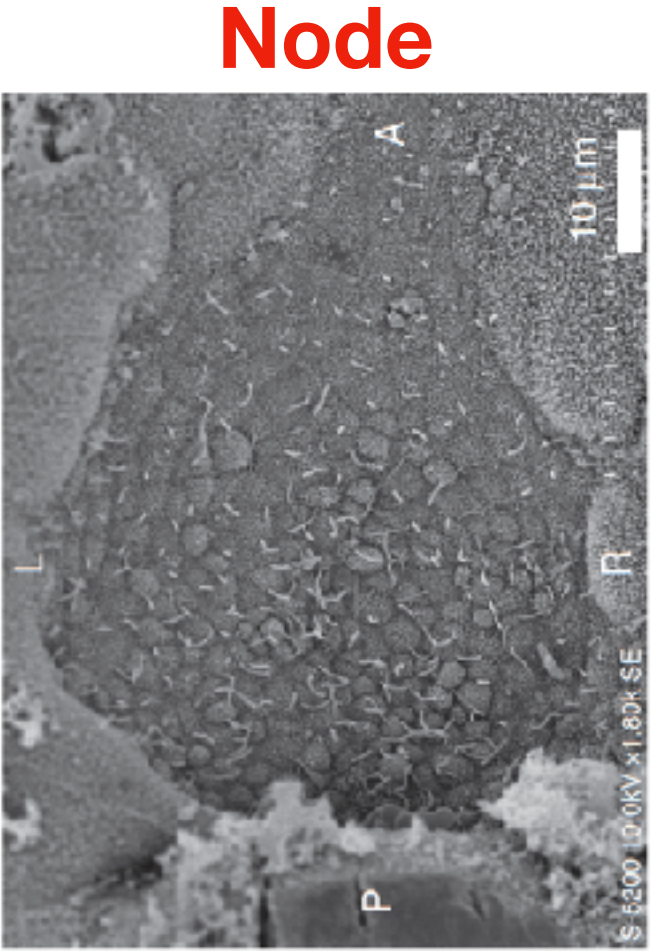
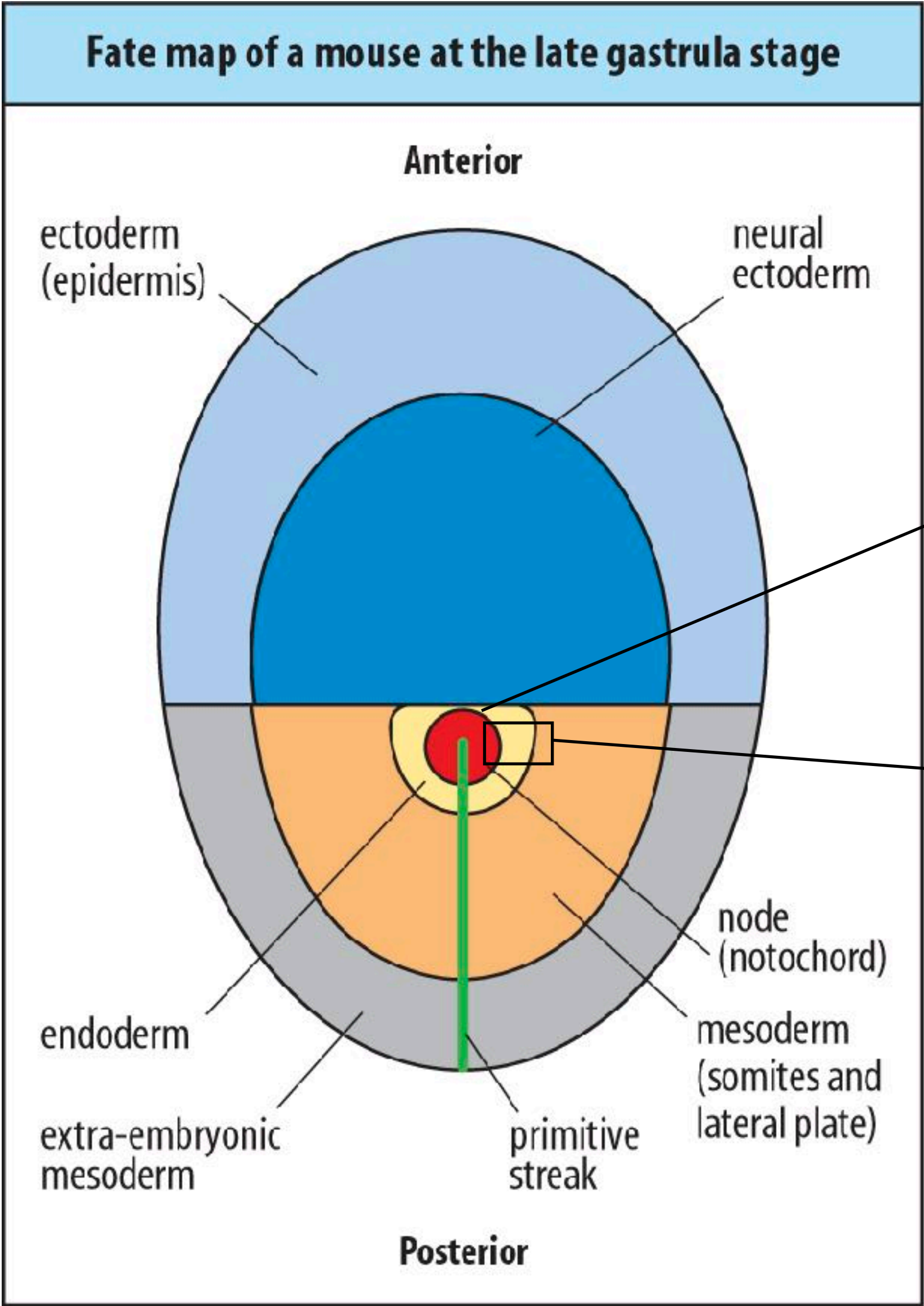
*right isomerism (asplenia)*



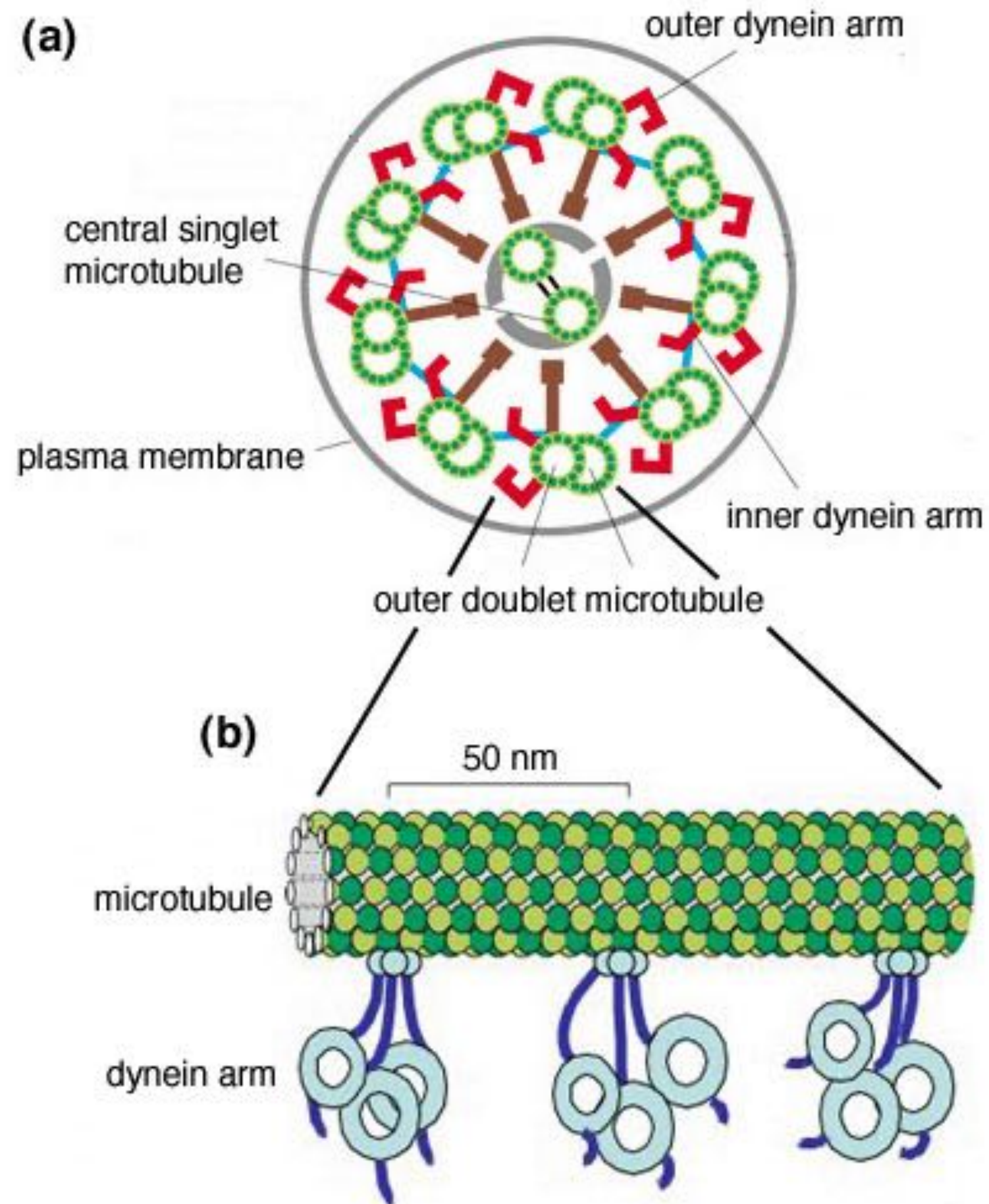
# Early mouse development



# Embryonic origin of the mouse heart



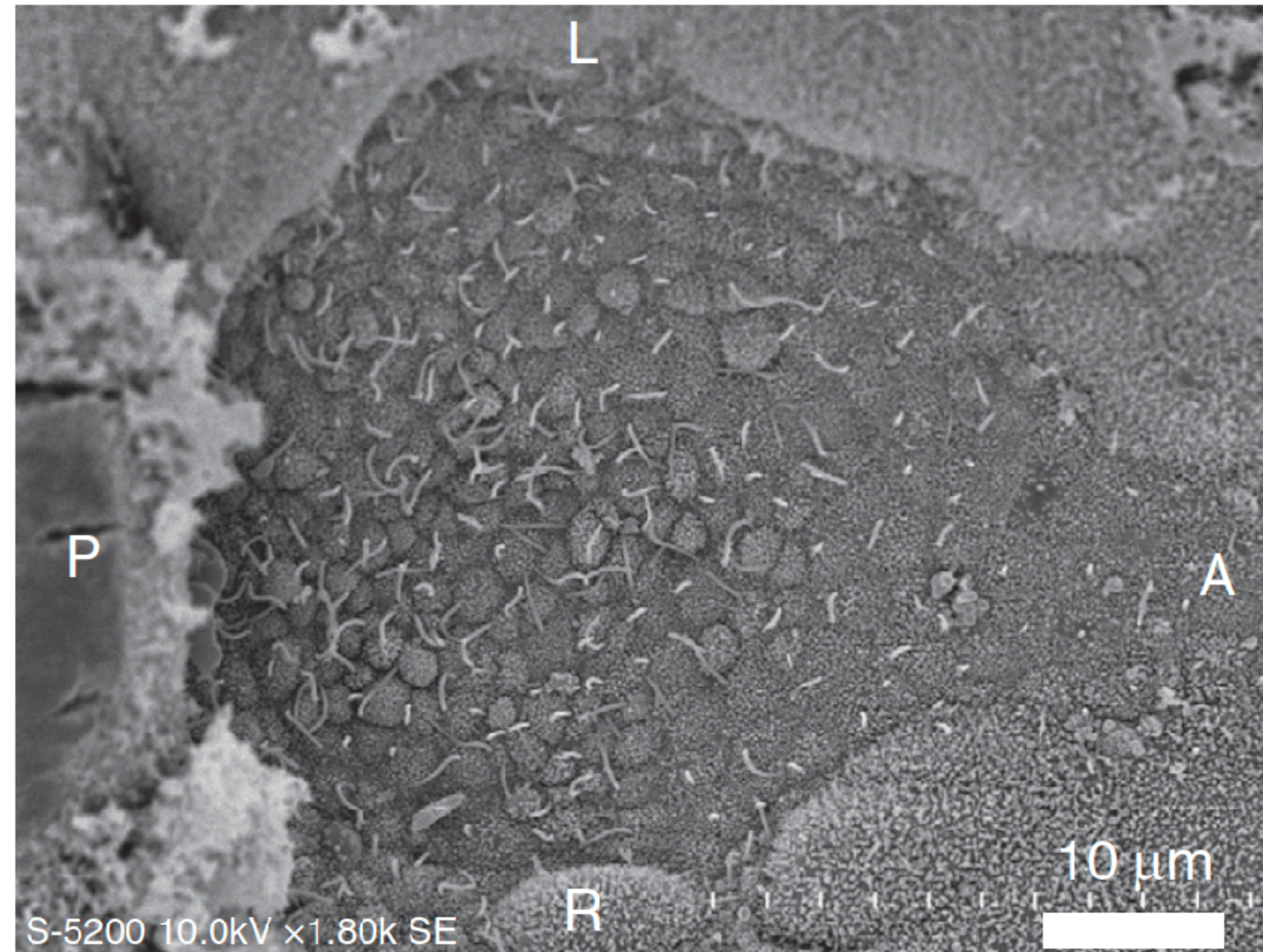
*inversus viscerum* encodes a  
**Dynein** outer arm heavy chain Lrd/Dnah11



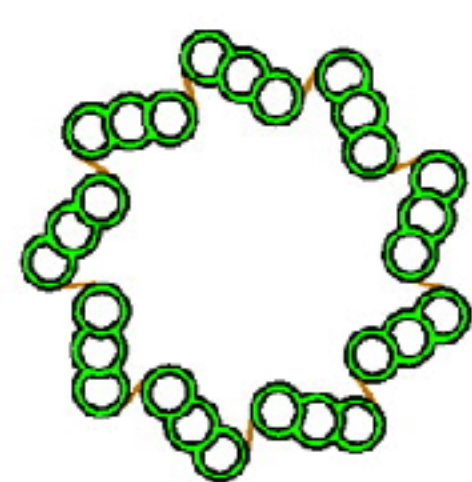
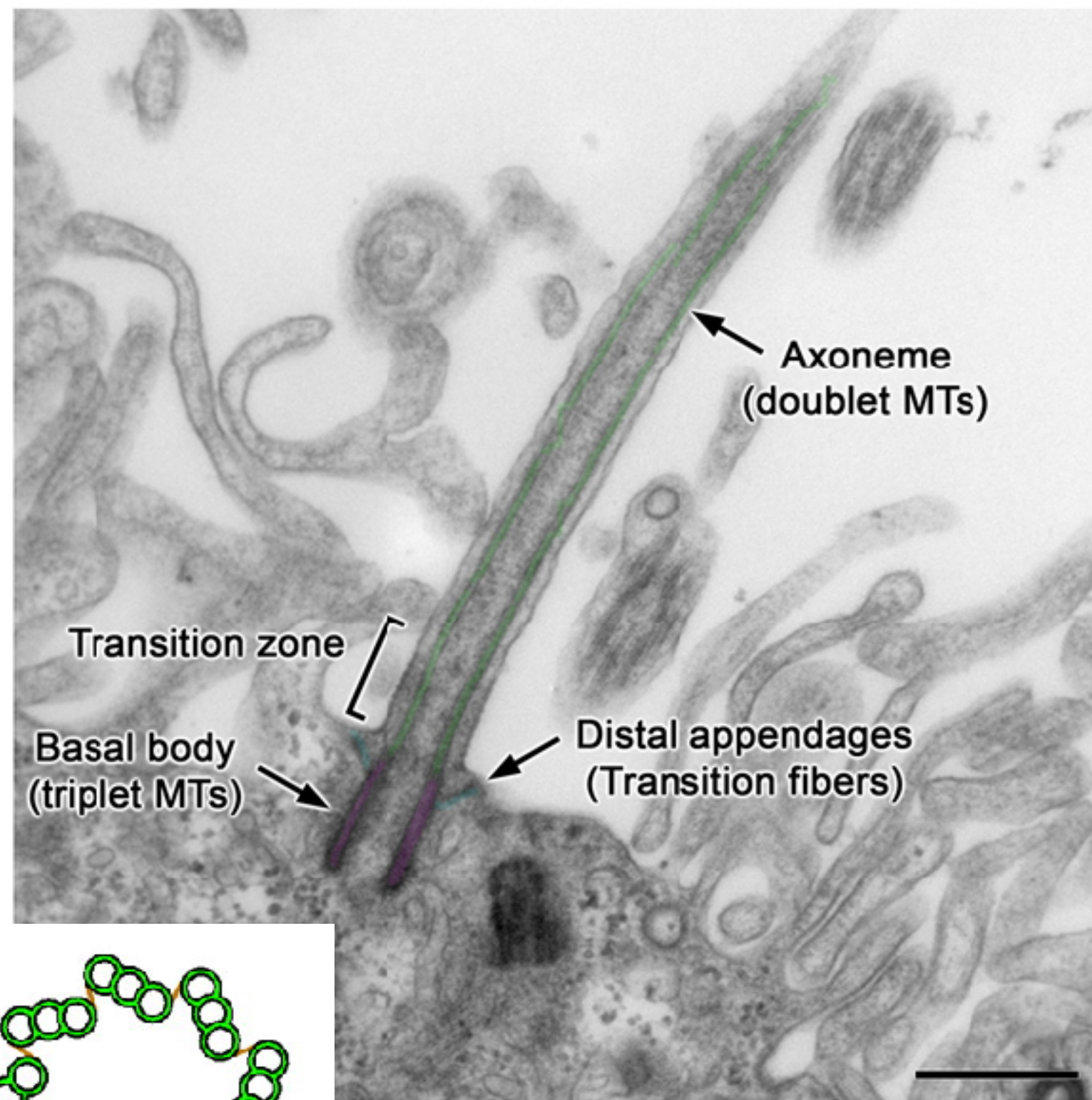
How is axonemal motion generated?

Dnah5 encodes another  
dynein outer arm heavy chain

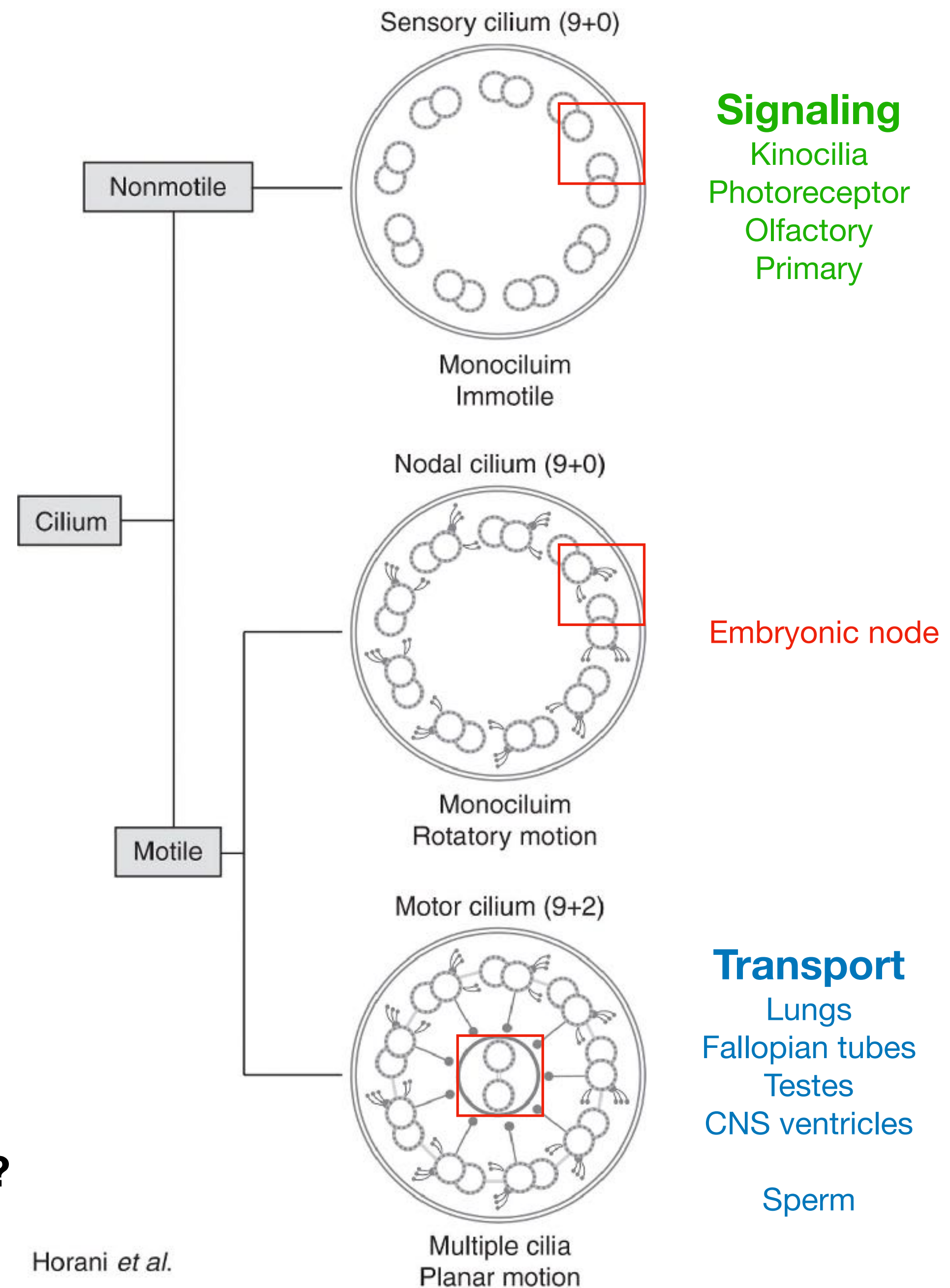
# The mouse node is covered in cilia



# Different cilia



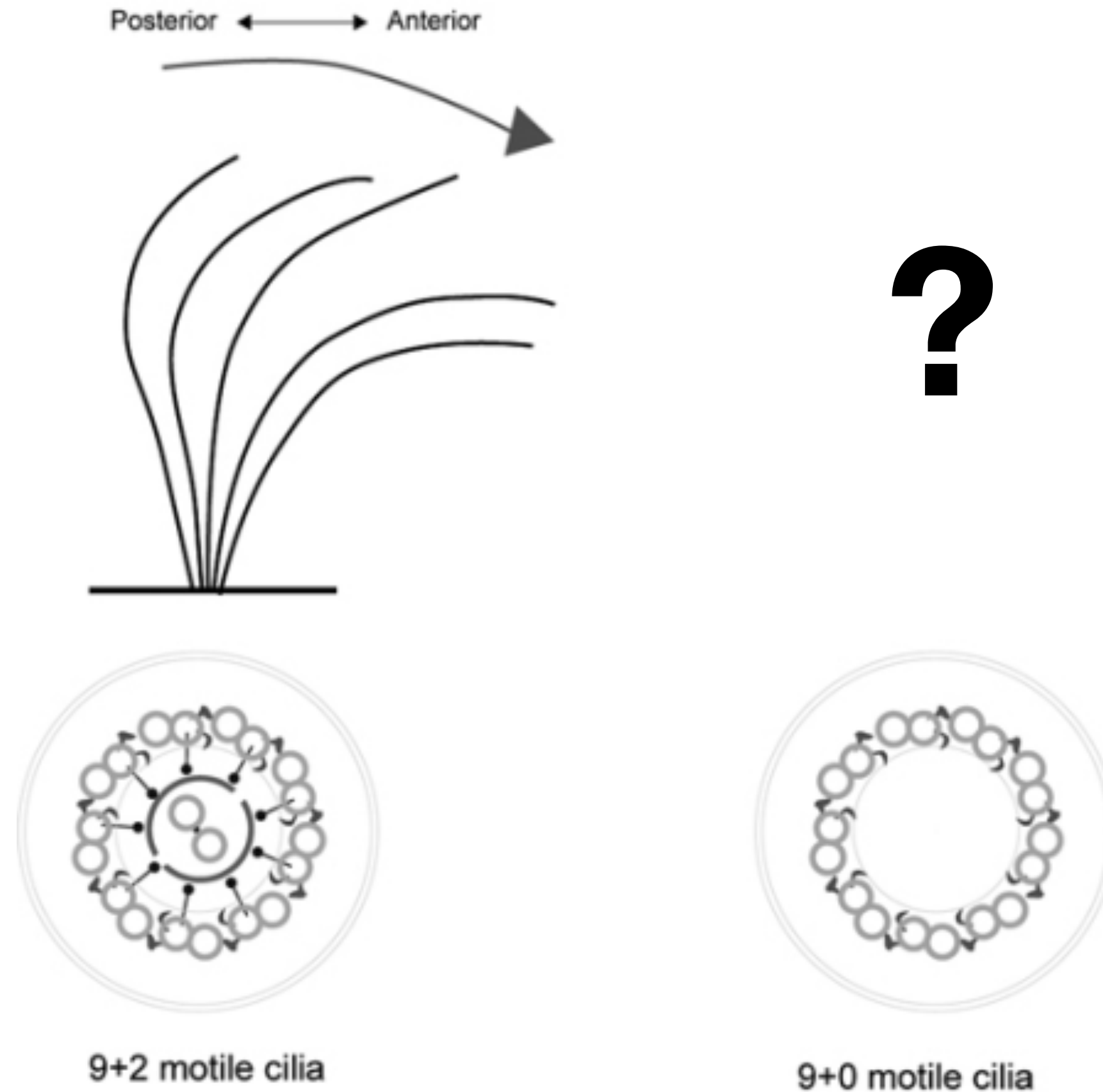
Basal body looks like a ...?



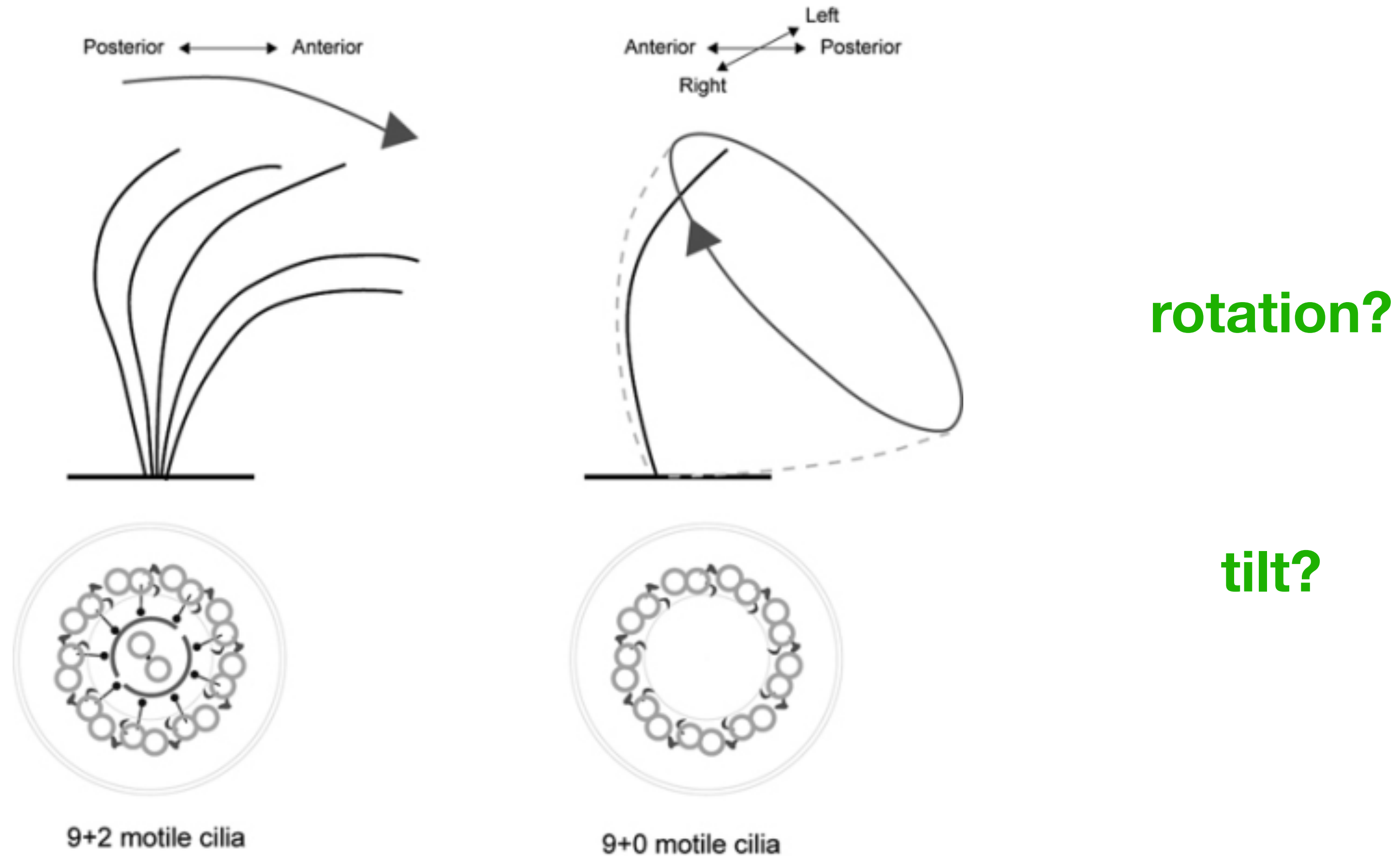


**Group puzzle**

# What motion could a node cilium use to drive persistent fluid flow?

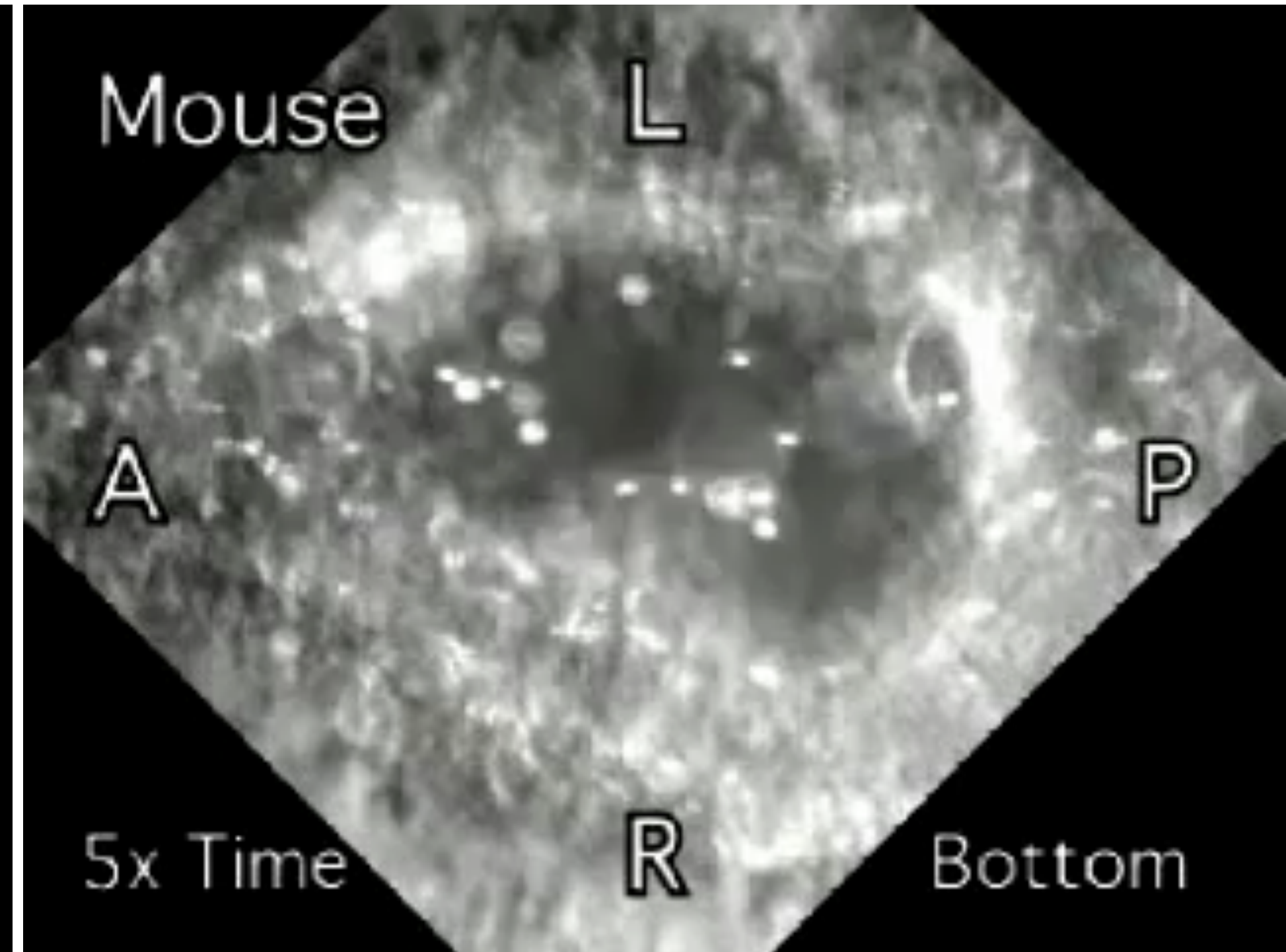
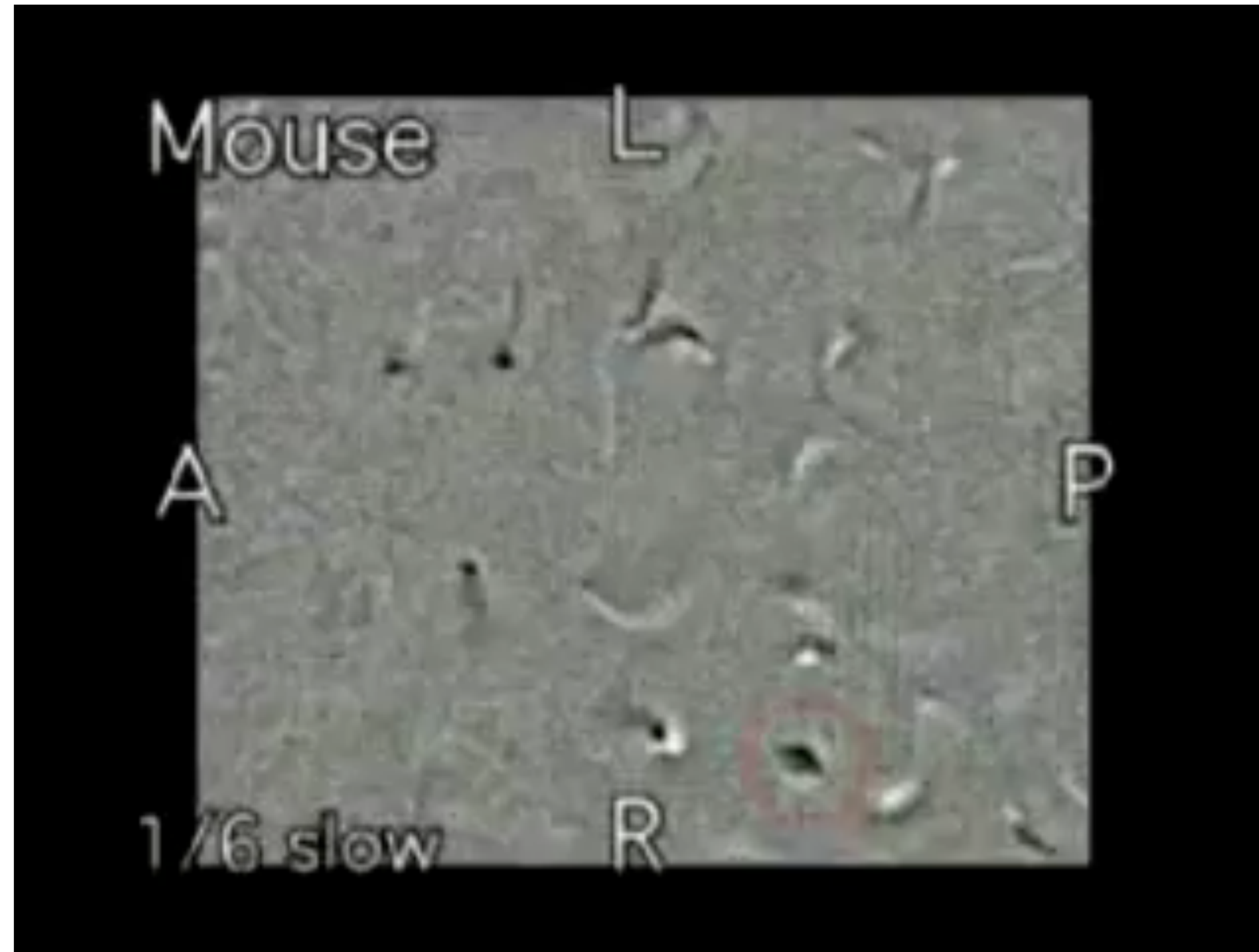


# Paddles and propellers



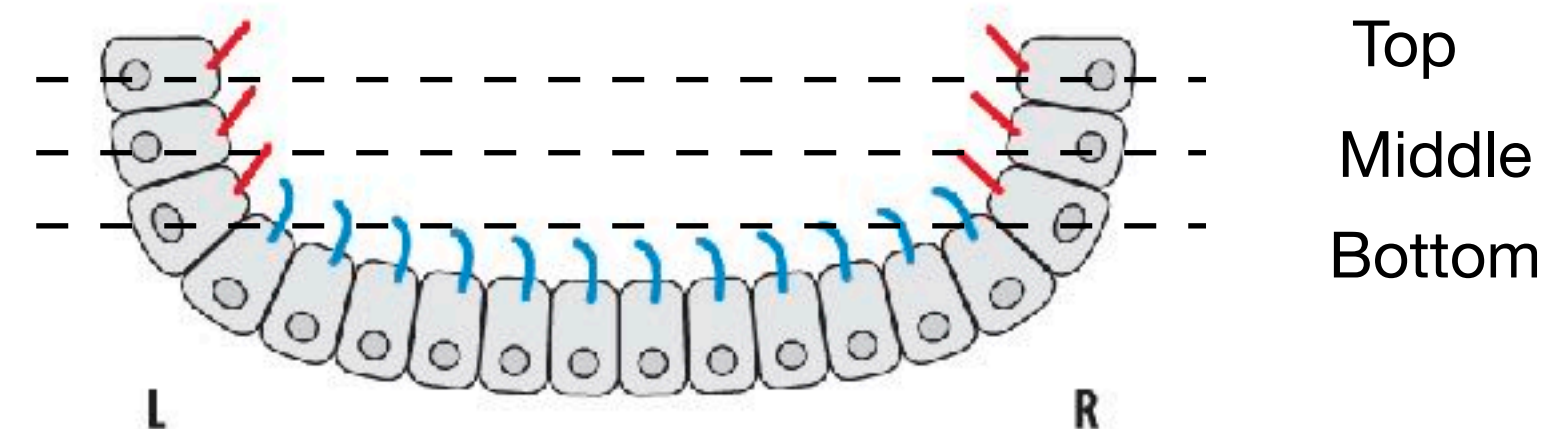
Rsph4a (radial spoke homolog) mutant mice have rotating cilia in their lungs!

# Cilia-induced fluid flow across node



**Cilia rotate clockwise,  
Elevated power stroke,  
Low recovery stroke,  
Point posteriorly**

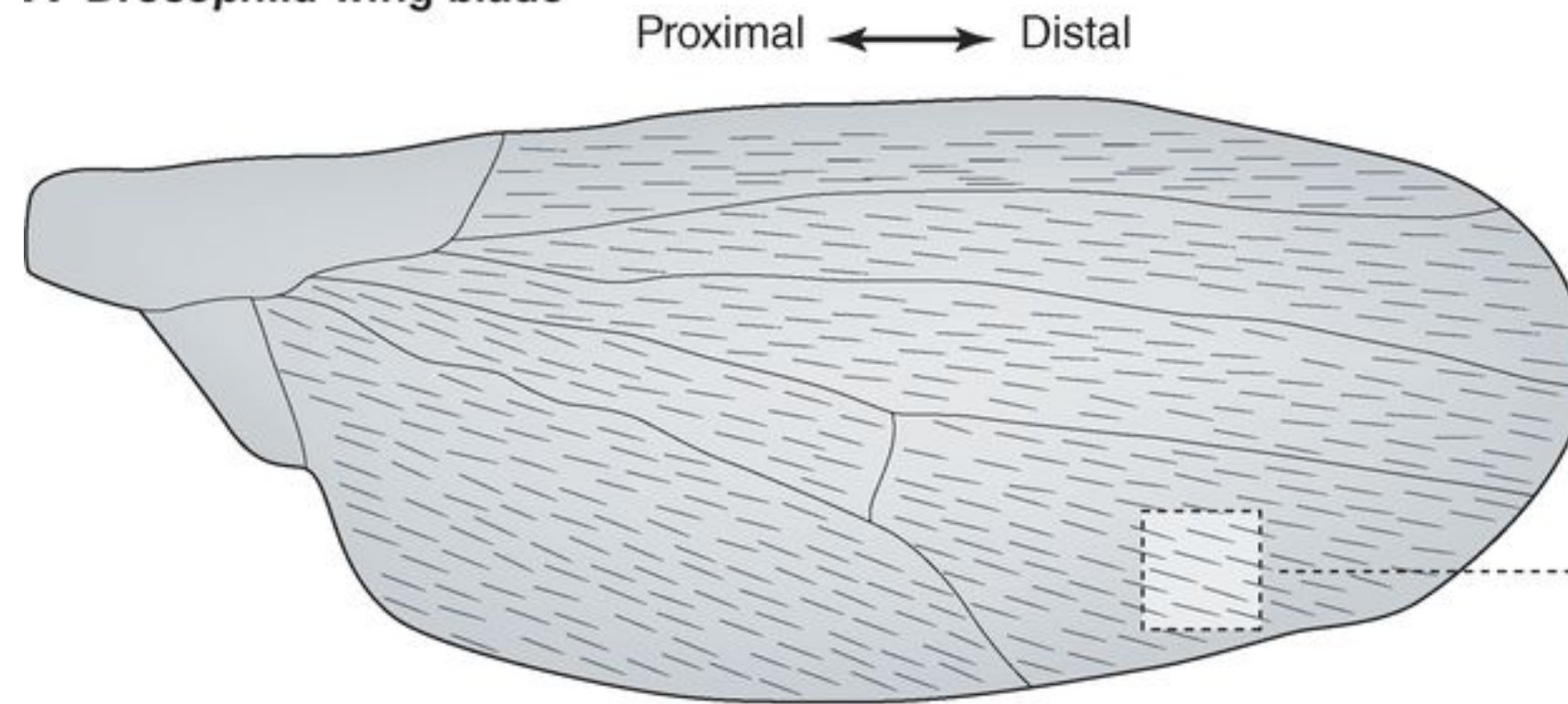
**Beads in flow  
Fast and slow**



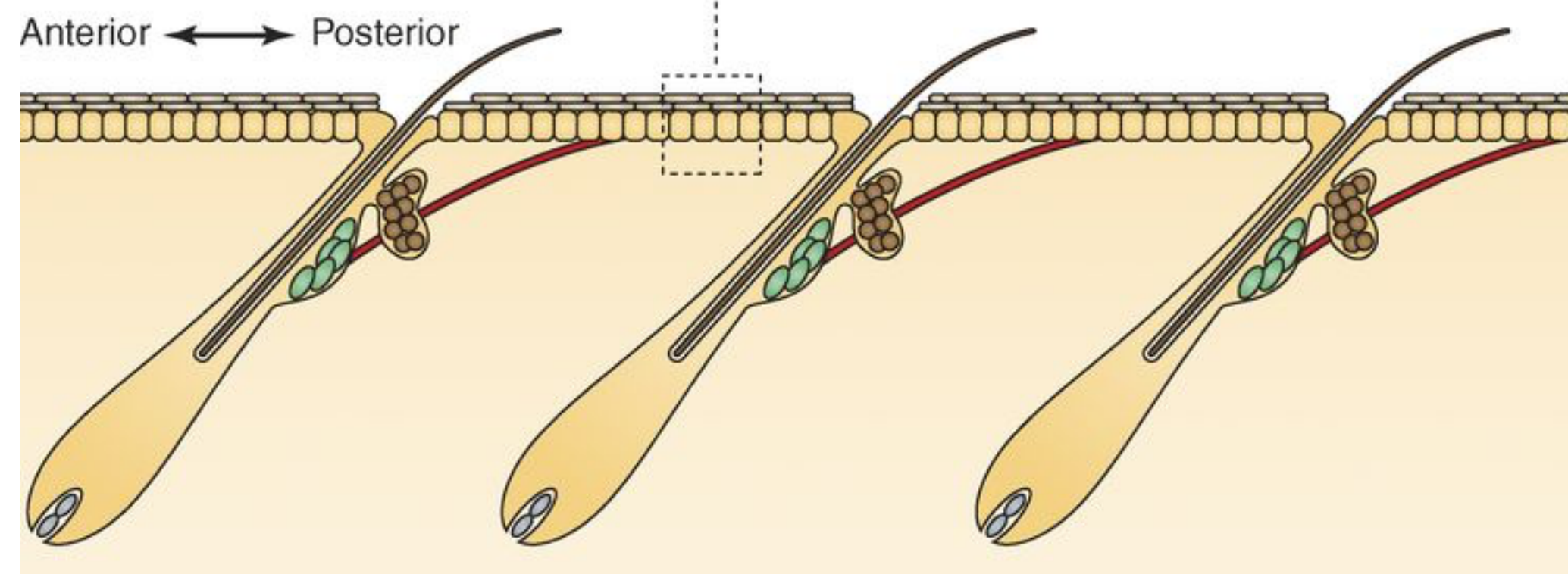
Okada et al, 2005

# Planar polarity in a 2D epithelial sheet

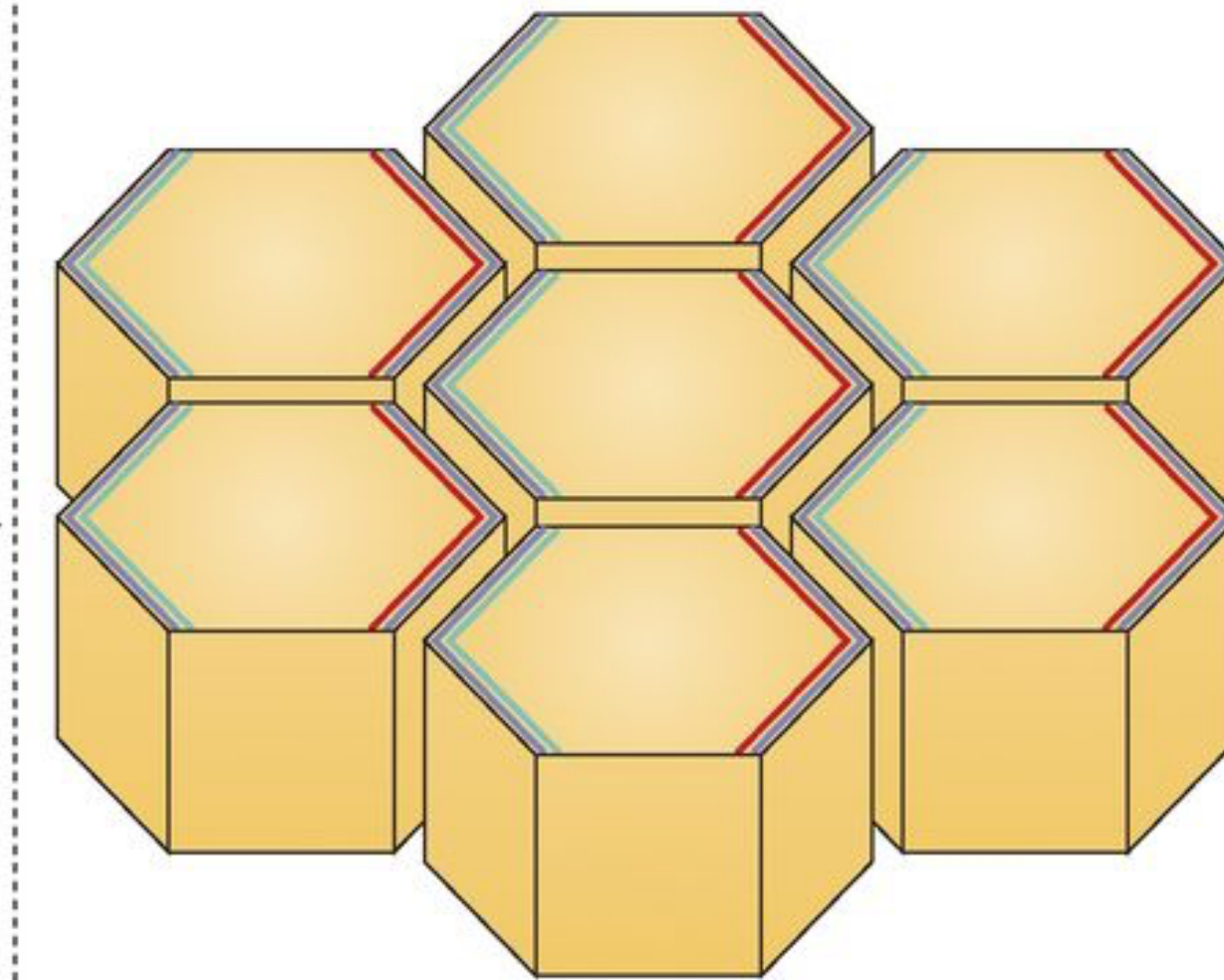
**A** *Drosophila* wing blade



**B** Mouse hair follicles



**C** Core PCP components



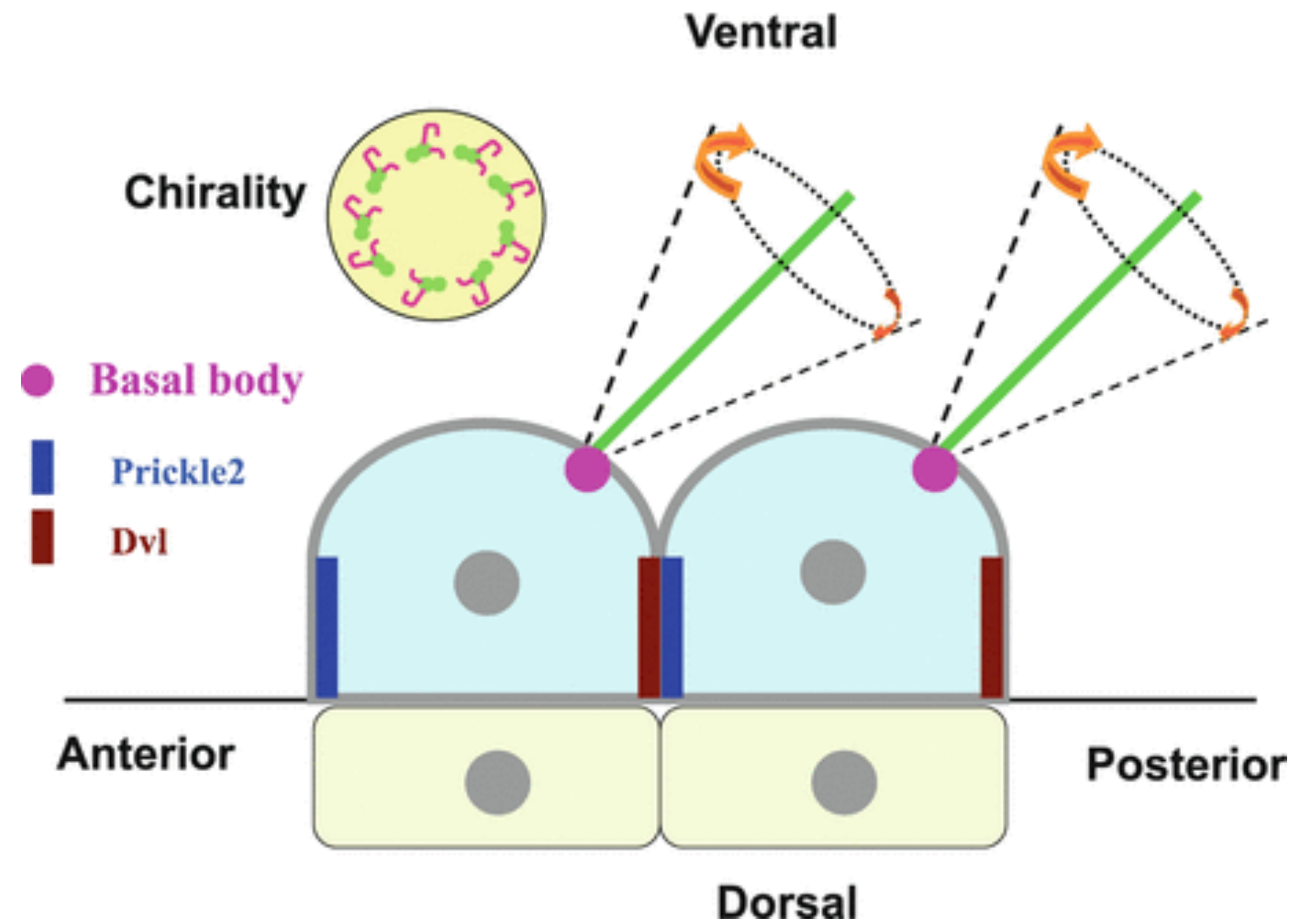
Proximal/anterior  
Van Gogh/Strabismus  
Prickle

Distal/posterior  
Frizzled  
Dishevelled  
Diego

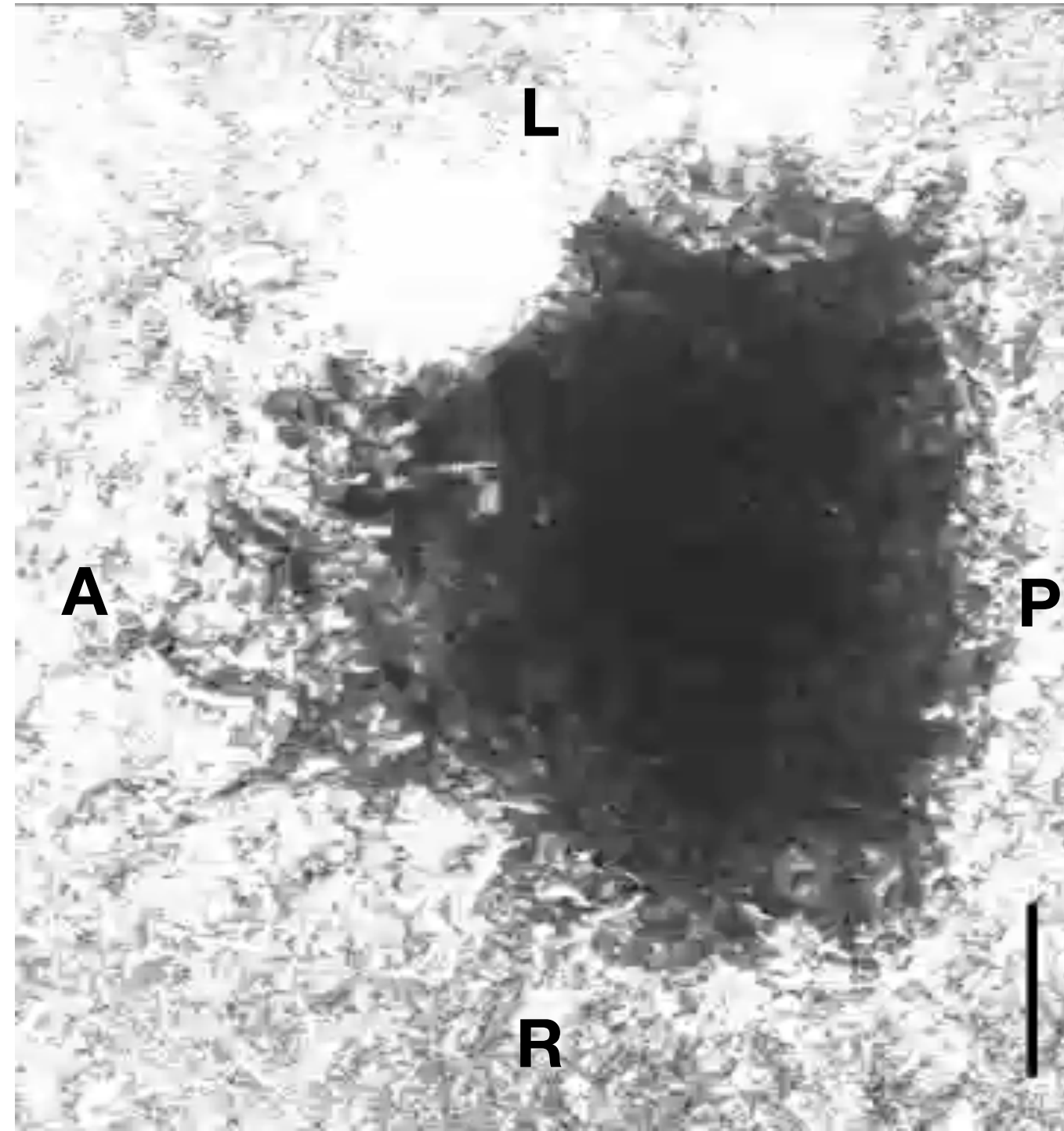
Flamingo/Starry night/Celsr

+ many other examples...

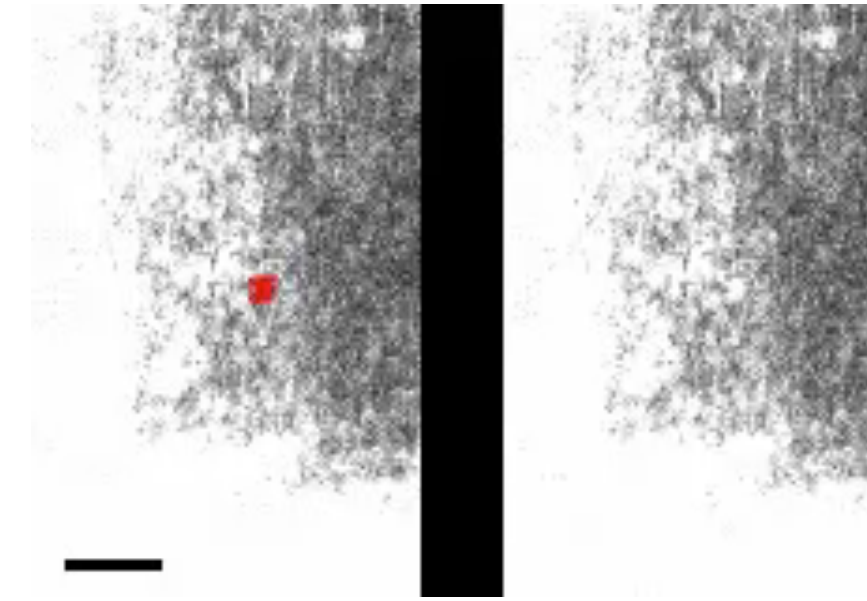
# The node epithelium is polarised early along the A-P axis



# Membrane particles move across node



**Dil labelled membrane**

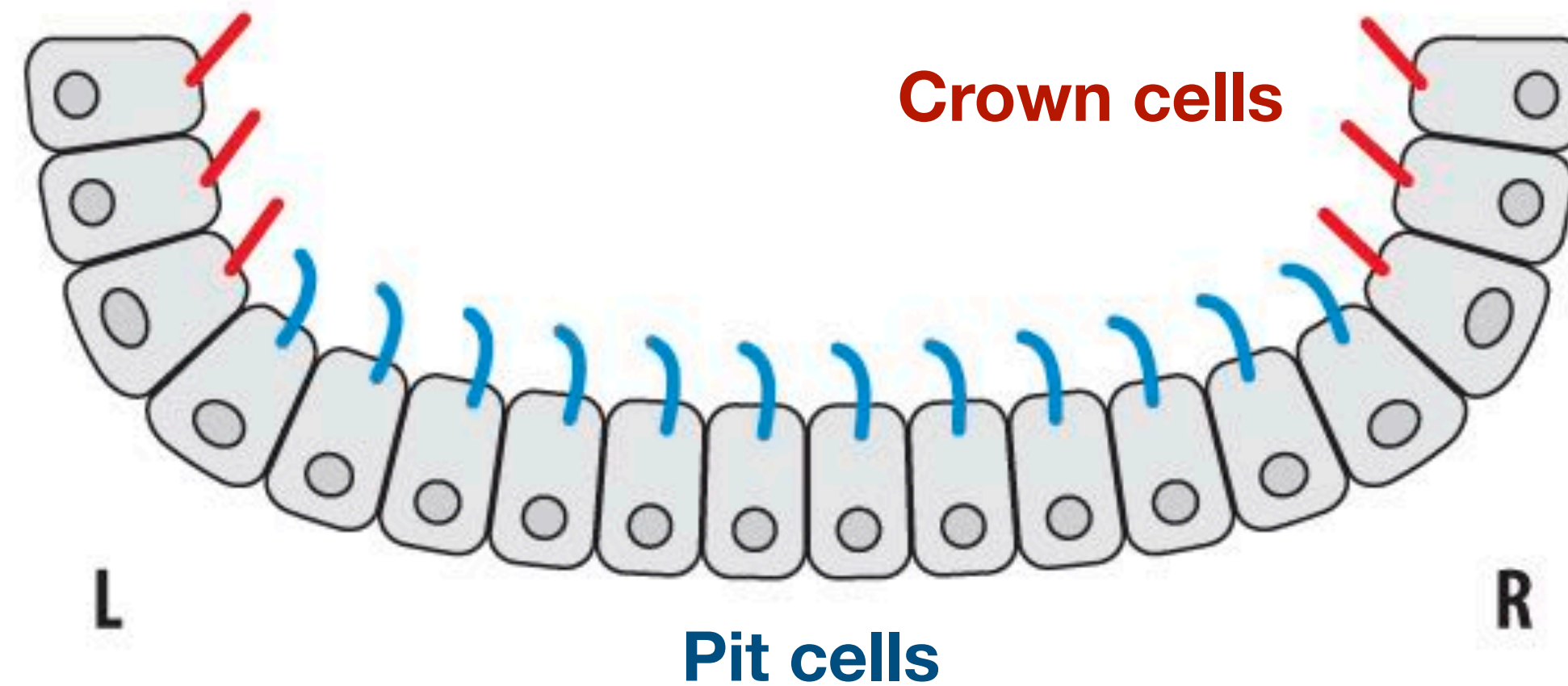
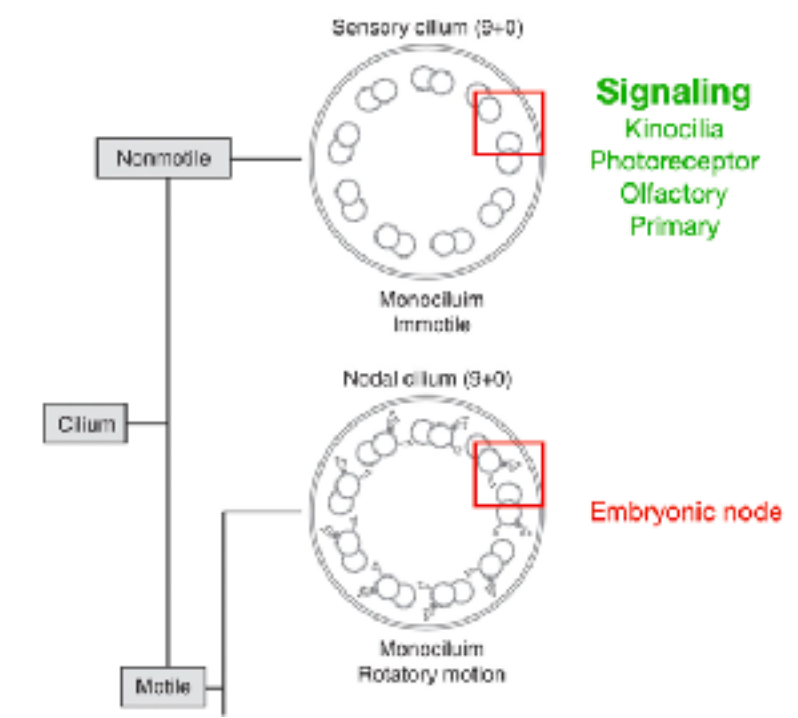


**Release from cilium?**



**Disruption on wall?**

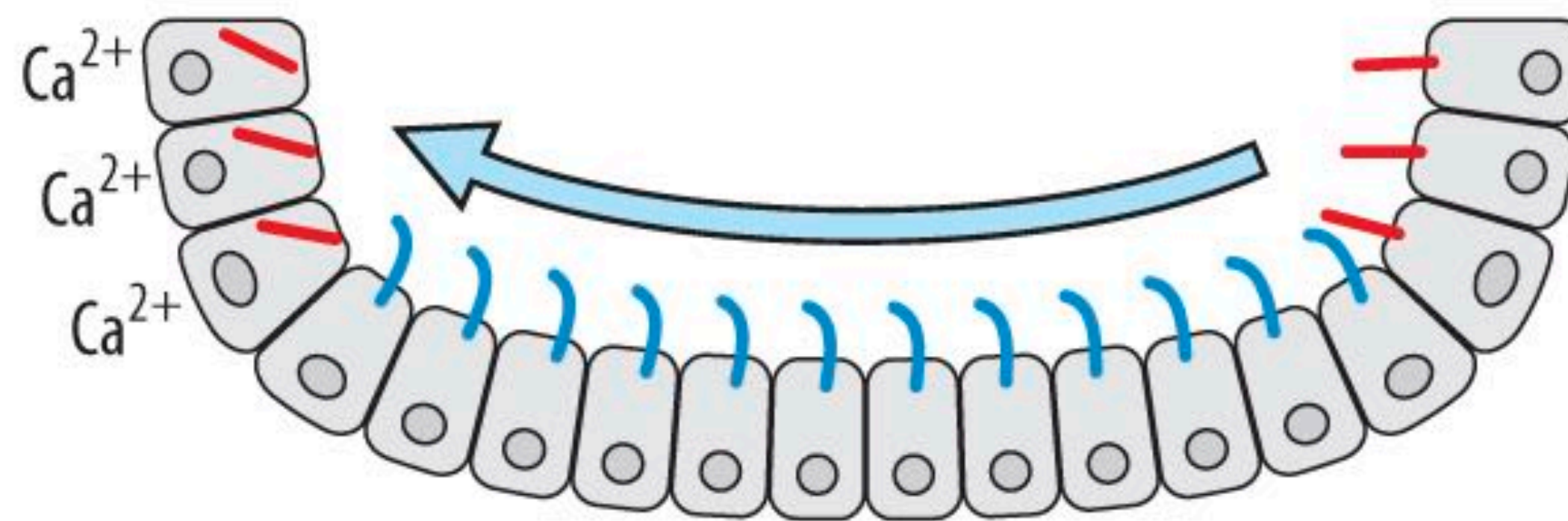
# Two types of cilia inhabit the node



- Polycystin-2** (Pkd2, TRPP2),
- $\text{Ca}^{2+}$ -permeable cation channel
  - polycystic kidney disease in humans
  - laterality defects in mouse
  - defects rescued by Pkd2 in crown cells

## Calcium

- thapsigargin randomises laterality
- $\text{Ca}^{2+}$  oscillations observed



**Nodal  
Lefty**

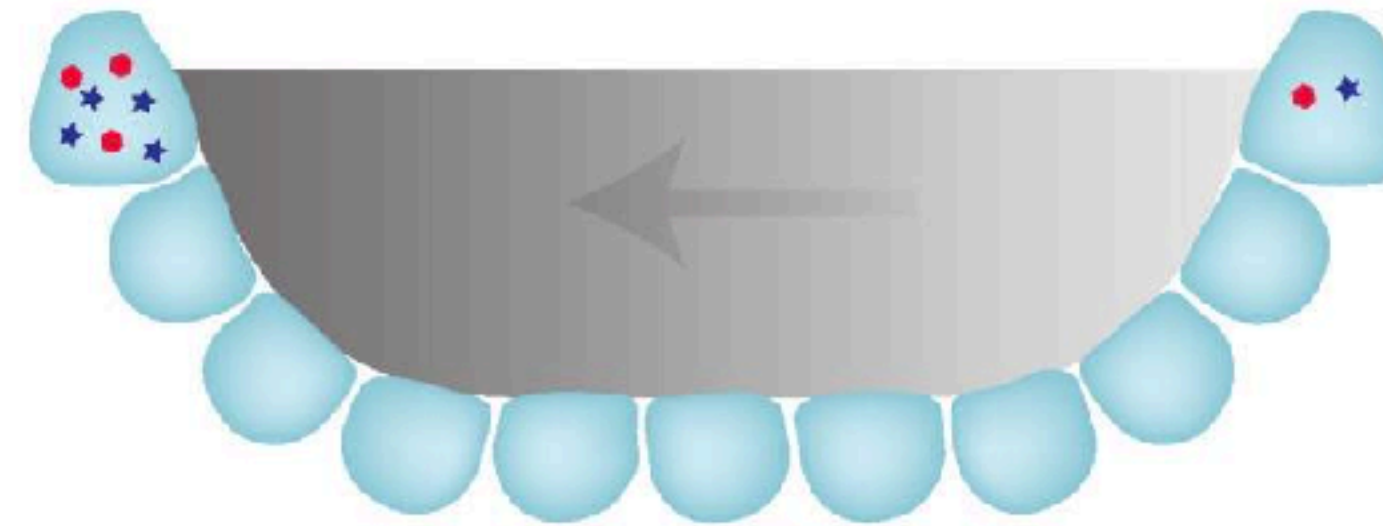




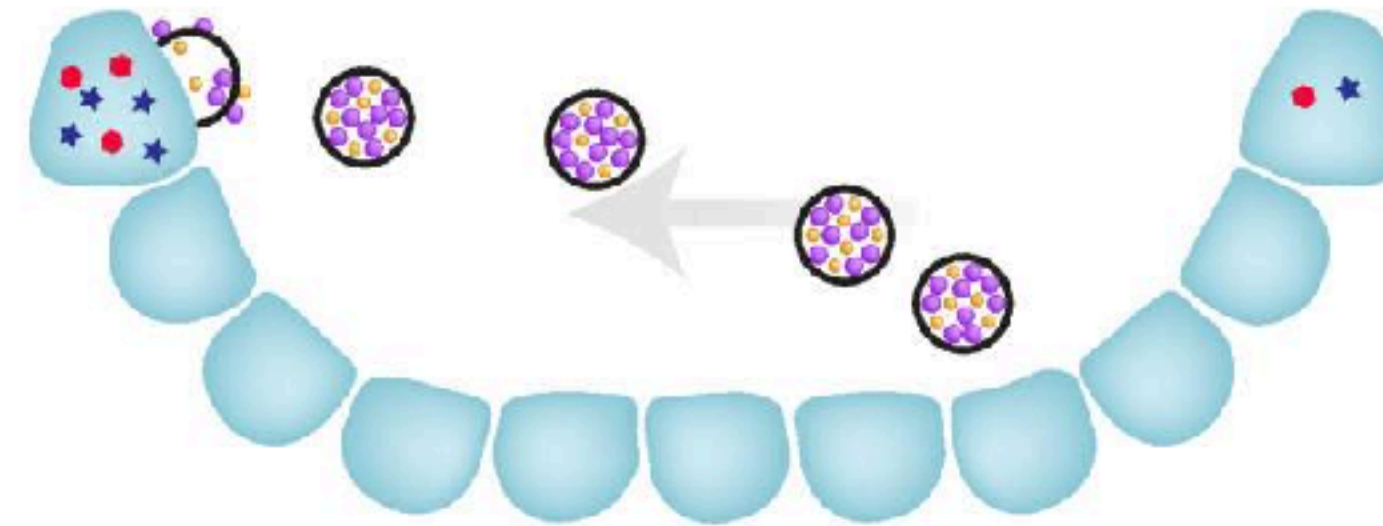
**Group puzzle**

# What is the signal to the cilia in the crown cells?

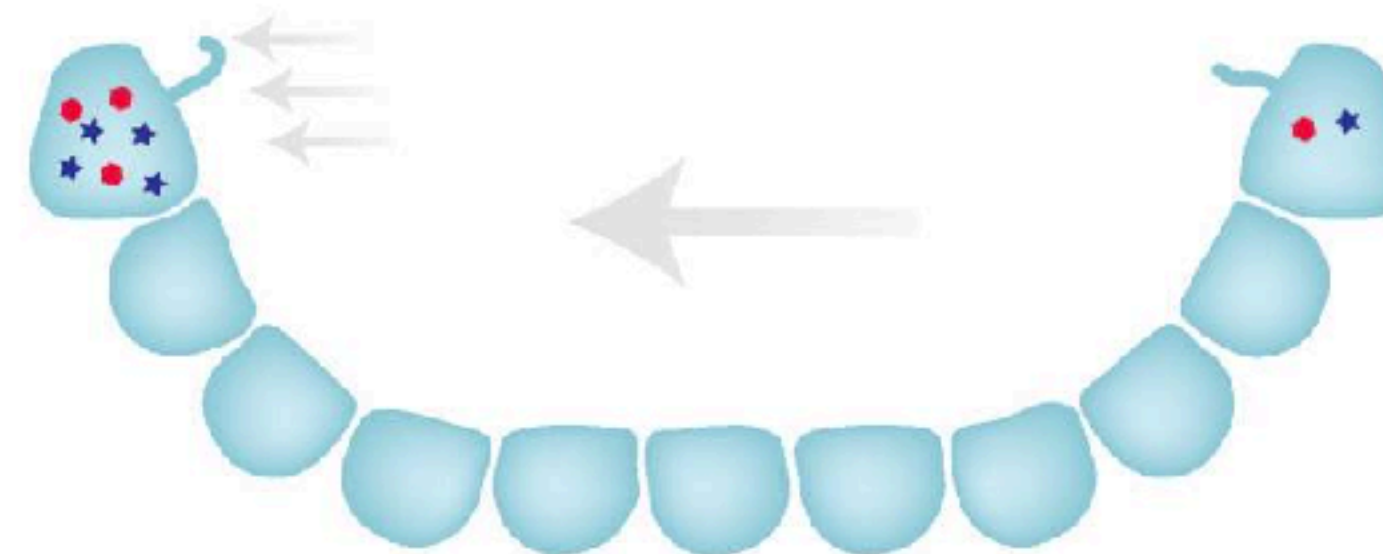
Morphogen gradient



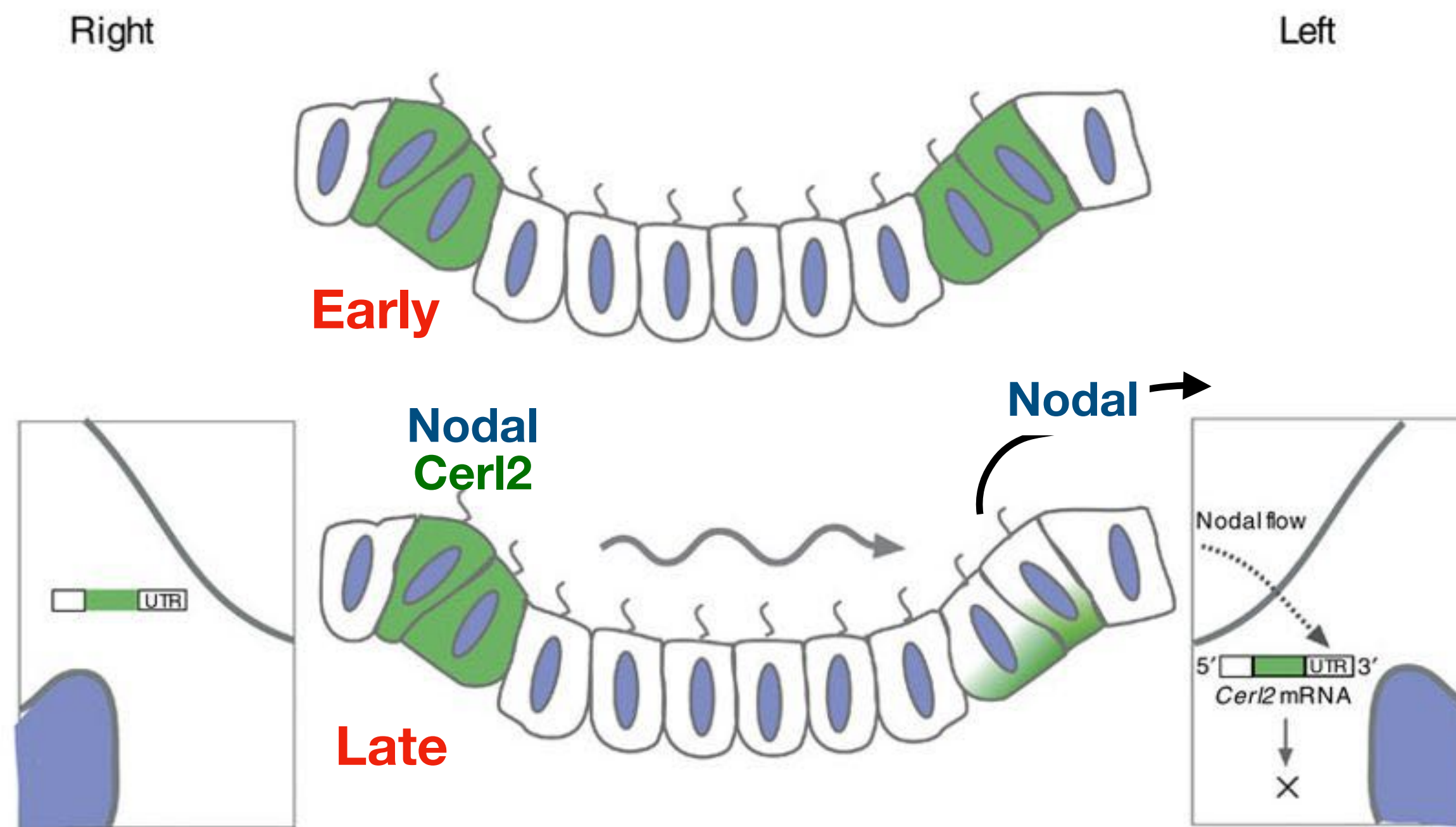
Nodal vesicular parcel (NVP)



Mechanosensation



# Signal processing to release asymmetric Nodal

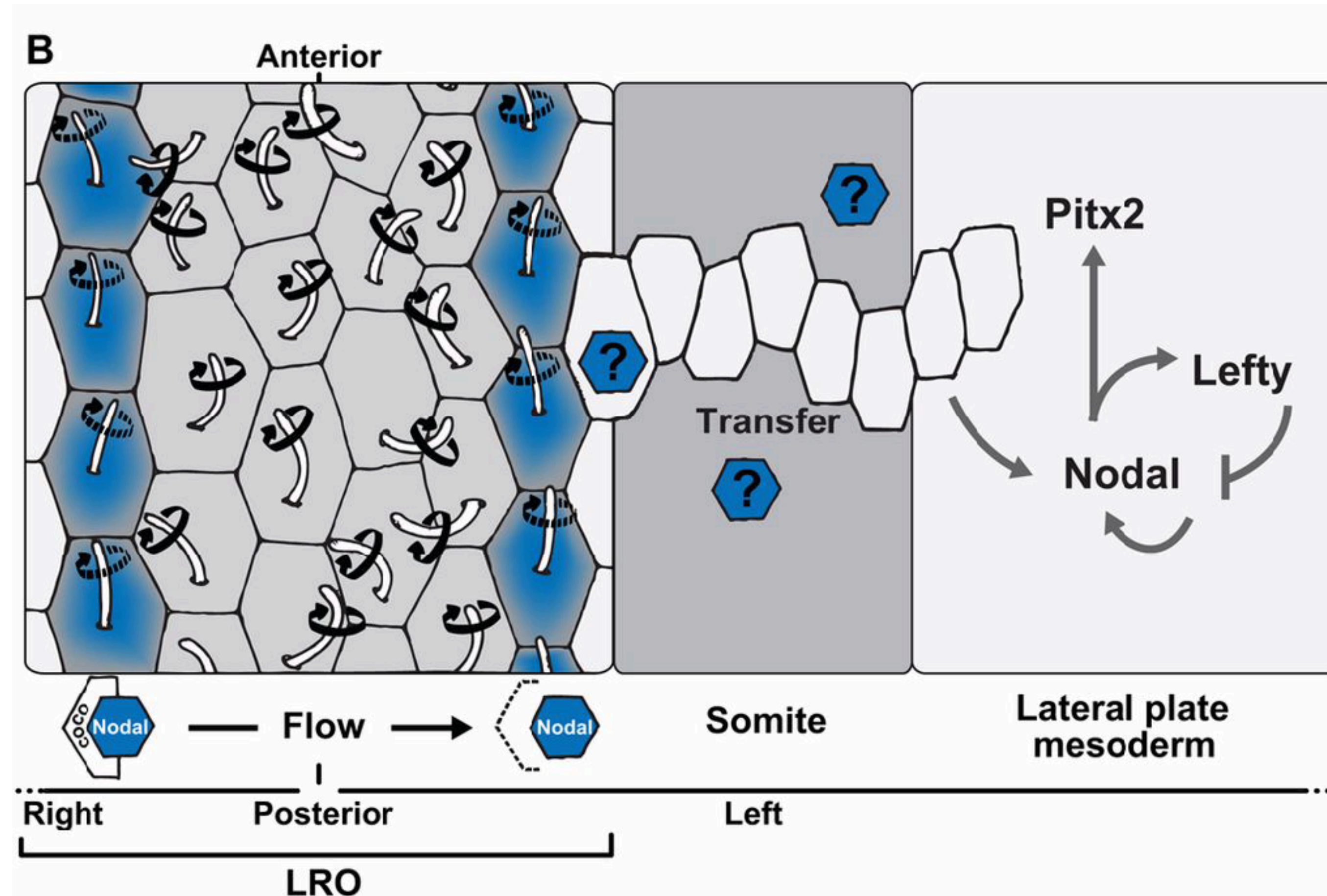


## Cerl2 / Dand5

Earliest asymmetric mRNA  
symmetric in *iv* mice  
Nodal inhibitor(!)  
blocks Nodal on RHS  
Nodal release on LHS



Nodal signals induce the transcription factor **Pitx2**, which activates morphogenetic genes in the LPM



How do node cilia rotate in the clockwise direction?

What is the positional cue that polarizes node cells along AP axis?

Do cilia act as mechano-sensors or chemo-sensors?

How is Cer12 mRNA stability controlled?

How is a small Nodal signal amplified along the LPM?



# Summary

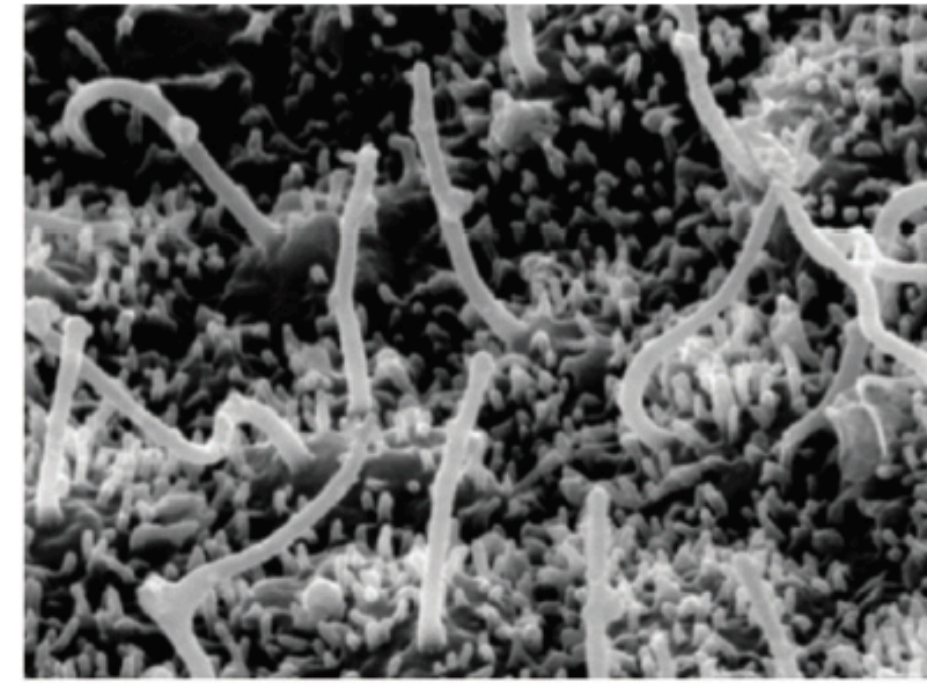
Symmetry breaking



Patterning



Organogenesis

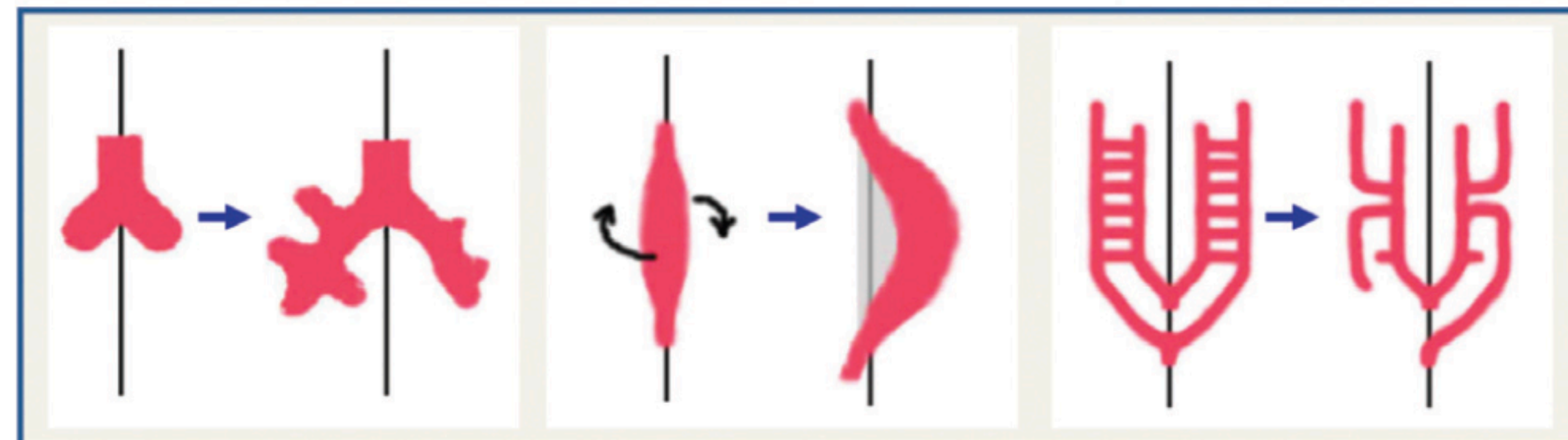


Node cilia

R L



*Nodal*  
*Lefty*



*Pitx2*

# Questions?

